

# PHYSICOCHEMICAL CHARACTERIZATION OF BANANA FRUIT BY UNIVARIATE AND MULTIVARIATE PROCEDURES

## CARACTERIZAÇÃO FÍSICO-QUÍMICA DE FRUTOS DE BANANEIRA POR PROCEDIMENTOS UNI E MULTIVARIADOS

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**ABSTRACT:** Breeding genotypes need to be characterized and evaluated in different soil and climatic conditions. There are few studies on association between characters evaluated and their contribution to banana genetic diversity, being essential to guide genetic breeding programs. This study aimed to evaluate the physicochemical characters of banana genotypes, to estimate associations between characters, to determine the relative importance of characters to study on genetic dissimilarity, and to indicate new genotypes to coastal plain region of Sergipe. Fruits from thirteen genotypes were used: Prata (FHIA-18, BRS Platina, PV94-01, BRS Garantida, YB42-47, Pacovan, and Prata Anã), Maçã (BRS Princesa, BRS Tropical, and Maçã), Ibota (Caipira), Gros Michel (Bucaneiro), and Mysore (Thap Maeo). The experimental design was randomized blocks, with thirteen genotypes and three replications. There is physicochemical variability of fruits between genotypes of same genomic group and subgroup, and between hybrids originated from same progenitor. Content of total sugars has greater contribution to genetic diversity among the genotypes, followed by fruit mass and pulp mass, and starch content in two cycles. Associations between fruit length, fruit diameter, and fruit mass and pulp mass in two production cycles based on phenotypic correlation are highly significant. Hybrids FHIA-18, PA94-01, YB42-47, and BRS Tropical, BRS Princesa, and Bucaneiro cultivars are promising for recommendation on coastal plain regions.

**KEYWORDS:** *Musa* spp. Cultivars. Hybrids.

### INTRODUCTION

Banana tree (*Musa* spp.) is one of the most important fruits being grown in approximately 150 countries (FAO 2016), part of human food in different socioeconomic classes by providing high nutraceutical value (ANJUM et al., 2014; SRIVASTAVA et al., 2015), ease of *in natura* consumption, low cost, and organoleptic characteristics that appeal most of the population.

Brazil has been raising its production and outstanding among the main global producers, occupying the fifth place in 2014 (FAOstat, 2017), with a production in 2016 around 6,962,134 tons in 516,980 hectares (IBGE, 2017).

Genotypes from genetic breeding program need to be characterized and evaluated in different areas of production and soil and climate conditions (SILVA et al., 2016), being relevant to evaluate field agronomic characteristics, as well as to characterize post-harvest attributes, allowing the identification of promising cultivars, for breeding programs or indication for farmers. Studies have been published in different soil and climatic conditions on physicochemical and metabolic characterization of banana fruits (SILVA et al.,

2006; 2016; PIMENTEL et al., 2010; MARQUES et al., 2011; BORGES et al., 2011; 2014; RIBEIRO et al., 2012; GODOY et al., 2013; MENDONÇA et al., 2013; RIBEIRO et al., 2016; ROSA, 2016). However, there are few studies on the association between the evaluated characters and their contribution to genetic diversity, thus being essential to guide genetic breeding programs.

This study aimed to evaluate the physicochemical characters of fruits of banana genotypes, to estimate correlations, to determine the relative importance of the characters to the study on genetic dissimilarity, and to indicate new genotypes to the coastal plain region of Sergipe.

### MATERIAL AND METHODS

Experiment assay was conducted from February 2012 to June 2013 in Jorge Sobral Experimental Field, in Nossa Senhora das Dores, SE, Brazil (10°2'S and 37°11'W), and average altitude of 208 m. Climate of region is semi-humid, with annual average rainfall of 1,161 mm, average air temperature of 25°C, and relative humidity of 77%. Soil of the area was classified as Typical Dystrophic Cohesive Yellow Latosol (Oxisols),

moderate A horizon, mid/loamy texture, plain. It presents average fertility, with low contents of aluminum ( $H + Al = 32.98 \text{ mmolc dm}^{-3}$ ), average acidity ( $pH = 5.52$ ), average levels of calcium and magnesium ( $Ca + Mg = 38.333 \text{ mmolc dm}^{-3}$ ), low contents of phosphorus ( $P = 6.3 \text{ mg dm}^{-3}$ ) and potassium ( $K = 35.105 \text{ mg dm}^{-3}$ ), and low organic matter content ( $OM = 19.8 \text{ g kg}^{-1}$ ). Micropropagated banana seedlings were planted in  $3.00 \text{ m} \times 2.00 \text{ m}$  spacing, under micro sprinklers irrigation system in view of the crop water demand. Control of scrublands, defoliation, of fertilizer foundation and cover, thinning, and other practices have been applied according to recommendations for the culture (BORGES; SOUZA, 2004).

Fruits were used in the second and third production cycles of thirteen banana genotypes, being from groups Prata (FHIA-18, BRS Platina, PA94-01, BRS Garantida, Pacovan, and Prata Anã), Maçã (BRS Princesa, BRS Tropical, YB42-47, and Maçã), Ibota (Caipira), Gros Michel (Bucaneiro), and Mysore (Thap Maeo), harvested when the first bunch featured greenish color with yellow traces, without corners and rounded sides (stage 2), and later placed on the counter at room temperature (average of  $26^{\circ}\text{C}$ ) until its complete maturation (stage 6).

Physical characters of fruit evaluated were: fruit length (cm), fruit diameter (cm), fruit mass (g), pulp mass (g), and pulp/peel ratio, determined by the quotient of mass the parts.

For chemical characters, after obtaining a homogeneous paste by grinding two fruits of each bunch/cluster/cultivar in a blender, we determined: contents of soluble solids (SS), reading in table refractometer (AOAC, 2012), expressed in  $^{\circ}\text{Brix}$ , pH on a sample of 10 g of crushed pulp diluted in 90 mL of distilled water with direct reading on digital pH meter (INSTITUTO ADOLFO LUTZ, 2005), total of titratable acid (TTA) at a sample of 5 g of crushed pulp diluted in 50 mL of distilled water with three drops of phenolphthalein solution to 1% as standard indicator with 0.1 N sodium hydroxide, expressed in % of malic acid (INSTITUTO ADOLFO LUTZ, 2005) and SS/TTA obtained by the ratio between the values of soluble solids (SS) and total titratable acidity (TTA).

Total sugars were quantified by spectrophotometry at a wavelength of 620 nm using a standard curve of fructose ( $100 \text{ mg mL}^{-1}$ ) of 0-100

mg range and expressed in percentage of glucose, according to Trevelyan; Harrison (1952). Reducing sugars were quantified by the reaction of banana pulp solution with Fehling's solution, according to methodology of Instituto Adolfo Lutz (1985). Starch content was obtained by Antrona method proposed by Trevelyan; Harrison (1952), expressed in percentage.

Experimental design was randomized blocks, with thirteen treatments (genotypes) and three replications. Each plot consisted of a hand composed of five fruits per bunch. For statistical analysis of physicochemical variables of the fruits were considered the average values of the second and third production cycles.

Averages of physicochemical variables of the fruits were submitted to analysis of variance, by F test and, when significant, grouped by Scott-Knott test at 5% probability level. Genetic dissimilarity averages were calculated using the Euclidean distance. Main components analysis was performed using the Singh criterion (1981). Were calculated all the possible associations between characters based on Pearson correlation. All analyses were carried out in the statistical program Genes (CRUZ, 2013).

## RESULTS AND DISCUSSION

There were significant differences between genotypes for physical characters of the fruits evaluated (Table 1).

Genotypes were grouped into three groups for the fruit length being the largest obtained by Bucaneiro (21.57 cm). In studies conducted by Carvalho et al. (2011) in Belém, PA, Brazil, Maçã group cultivars: Caipira (11.19 cm), Thap Maeo (10.44 cm), and Tropical (10.55 cm) reached lower values compared with the same cultivars in this study (14.22; 13.74 and 17.14 cm, respectively). In dystrophic red Latosol (Oxisol) of Paraná, Borges et al. (2011) also observed smaller fruit length for the genotypes of Maçã group, FHIA-18, Prata Anã. However, the hybrids PA94-01 and BRS Platina have achieved higher values (19 cm) than those obtained in yellow latosol of the coastal plain region (17.47 and 14.06 cm, respectively). Fruit length is an important physical character for banana classification ensuring homogeneity of the marketing (PBM & PIF, 2006) and, consequently for remuneration (AZEVEDO et al., 2010).

**Table 1.** Average values of physical characteristics of banana tree genotypes in the second and third production cycles – fruit length (FL), fruit diameter (FD), fruit mass (FM), pulp mass (PM), and pulp:peel ratio (PPR)<sup>(1)</sup>.

| Genotypes     | FL (cm) | FD (cm) | FM (g)   | PM (g)   | PPR    |
|---------------|---------|---------|----------|----------|--------|
| FHIA-18       | 17.36 b | 3.71 a  | 130.37 b | 86.53 c  | 1.97 b |
| BRS Platina   | 14.06 c | 3.15 b  | 87.65 c  | 61.81 d  | 2.62 b |
| YB42-47       | 13.67 c | 3.88 a  | 107.51 c | 85.58 c  | 3.92 a |
| PA94-01       | 17.47 b | 3.46 b  | 123.35 b | 86.36 c  | 2.43 b |
| BRS Garantida | 17.69 b | 4.24 a  | 188.02 a | 120.98 b | 1.92 b |
| Pacovan       | 13.67 c | 3.36 b  | 88.53 c  | 59.73 d  | 2.24 b |
| Prata Anã     | 13.87 c | 3.21 b  | 87.09 c  | 60.00 d  | 2.27 b |
| BRS Princesa  | 13.42 c | 3.45 b  | 90.71 c  | 69.57 d  | 3.48 a |
| BRS Tropical  | 17.14 b | 3.93 a  | 143.48 b | 113.98 b | 4.08 a |
| Maçã          | 15.74 c | 3.90 a  | 137.22 b | 112.10 b | 4.05 a |
| Caipira       | 14.22 c | 3.40 b  | 102.20 c | 80.26 c  | 3.80 a |
| Bucaneiro     | 21.57 a | 3.79 a  | 193.96 a | 146.12 a | 3.11 a |
| Thap Maeo     | 13.74 c | 3.73 a  | 117.08 b | 91.58 c  | 3.97 a |
| CV (%)        | 11.45   | 10.51   | 20.85    | 20.04    | 23.00  |

<sup>(1)</sup> Averages followed by the same letter in column belong to the same group according to the clustering method of Scott-Knott at 5% of significance; CV (coefficient of variation).

Regarding the fruit diameter, two groups were formed: first, with higher averages, consisting of FHIA-18, YB42-47, BRS Garantida, BRS Tropical, Maçã, Bucaneiro, and Thap Maeo, ranging from 3.71 to 4.24 cm; and second formed by BRS Platina, PA94-01, Pacovan, Prata Anã, BRS Princesa, and Caipira, with amplitude ranging from 3.15 to 3.46 cm. Differentiated responses between genotypes for this characteristic were also observed by Borges et al. (2011), Carvalho et al. (2011) and Marques et al. (2011). However, Rosa (2016) also observed non-significant difference between Prata Anã, BRS Princesa e BRS Platina. Despite BRS Platina is a hybrid, for this character, in the present work, was not superior to its genitor as reported by Silva et al. (2013). Both fruit diameter and fruit length are important for trade classification (DONATO et al., 2009).

Considering the fruit mass, the first group was constituted by Bucaneiro (193.96 g) and BRS Garantida (188.02 g). The FHIA-18, PA94-01, BRS Tropical, Maçã, and Thap Maeo genotypes presented average values between 117.08 and 143.48 g, and third group, formed by BRS Platina, YB42-47, Pacovan, Prata Anã, BRS Princesa, and Caipira hybrids, ranged from 87.09 to 107.51 g. Productivity, according to Lessa et al. (2012), is associated with fruit mass and number of fruits. For pulp mass, four groups were formed, and the Bucaneiro presented the greater mass (146.12 g),

followed by BRS Garantida (120.98 g), BRS Tropical (113.98 g), and Maçã (112.10 g). FHIA-18, YB42-47, PA94-01, Caipira, and Thap Maeo genotypes the third group; PA42-44, Pacovan, Prata Anã, and BRS Princesa the fourth group. BRS Princesa cultivar obtained smaller fruit mass and pulp mass (90.71 and 69.57 g, respectively) when compared with other genotypes of Maçã group. However, in Fluvents with average fertility in Baixo São Francisco, SE, Lédo et al. (2008) obtained greater fruit mass for BRS Princesa (132.4 g). This fact contributes to the genotype x environment interaction a mentioned by Léon et al. (2016). Probably the environmental conditions, as humid, at Baixo São Francisco, SE was more adequate to BRS Princesa development than the semi-arid region.

First group to pulp/peel ratio, with higher averages, consisted of the genotypes YB42-47, BRS Princesa, BRS Tropical, Maçã, Caipira, Bucaneiro, and Thap Maeo; and the second, by BRS Platina, FHIA-18, PA94-01, Pacovan, Prata Anã, and BRS Garantida. Pulp yield is a feature of economic importance for the industry, since it is directly related to yield in processing fruits (CHITARRA; CHITARRA, 2005).

Overall, it has not been verified the grouping of all Prata Anã hybrids in groups higher than its progenitor regarding physical characters, with the exception of PA94-01 and FHIA-18 for length and fruit mass, corroborating Donato et al.

(2009), observed the same behavior of hybrid BRS Platina and its progenitor, Prata Anã, in relation to BRS FHIA-18, FHIA-18, and FHIA-01 (BRS Maravilha).

Variety observed in the responses of genotypes regarding physical characters, depending on the location and type of soil, is expected. According to León et al. (2016) expression of a phenotype is a function of the genotype, the environment and differential phenotypic response of genotypes to different environments, also known as genotype by environment (G X E) interaction. Kang et al. (1991) cited by Tonk et al. (2011) indicated that selection based on yield only may not always be adequate when genotype x environment interaction is significant. These factors reinforce the need for evaluation of genotypes in different environments and crop conditions for recommendation purposes.

There were significant differences between genotypes for contents of soluble solids, titratable total acidity, and reducing sugars (Table 2). Four groups were formed for soluble solids content, being the first consisting of Pacovan, BRS Princesa, and Prata Anã; the second, by Maçã; the third, by FHIA-18, YB42-47, BRS Garantida, Caipira, and Thap

Maeo; and the fourth, with the lowest averages, by BRS Platina, PA94-01, BRS Tropical, and Bucaneiro.

Genotypes were gathered into two groups regarding total titratable acidity: the first was formed by BRS Tropical, BRS Princesa, Maçã, Pacovan, and YB42-47, ranging from 0.93 to 1.27% malic acid; and the second, by FHIA-18, PA94-01, Thap Maeo, BRS Garantida, Caipira, BRS Platina, Prata Anã, and Bucaneiro, ranging from 0.67 to 0.83% malic acid. High total titratable acidity is a desirable feature for the industry (Godoy, 2010), which emphasizes the suitability of Bucaneiro for processing industry. In a study by Pimentel et al. (2010), conducted in the North of Minas Gerais, Prata Anã presented a higher average of total titratable acidity (0.69%) compared with BRS Platina (0.65%), in contrast with the results in which both genotypes did not differ. Ribeiro et al. (2012), evaluating the behavior of cultivars in organic and conventional management in Cruz das Almas, BA, Brazil, verified Caipira cultivar significantly differed from others in the organic system, with the lowest acidity (0.11%), and the largest in BRS Tropical (0.23%) on the conventional system.

**Table 2.** Average values of physicochemical characteristics of banana genotypes in the second and third production cycles – SS (°Brix), TTA (% malic acid), RS (% glucose), TS (% glucose), and STA (%).

| Genótipos     | SS      | ATT    | RAT     | AR      | TS      | STA    | pH     |
|---------------|---------|--------|---------|---------|---------|--------|--------|
| FHIA-18       | 23.53 c | 0.83 b | 28.83 a | 8.99 b  | 19.49 a | 1.15 a | 4.85 b |
| BRS Platina   | 22.05 d | 0.67 b | 33.54 a | 15.07 a | 20.46 a | 0.92 a | 5.06 a |
| YB42-47       | 24.34 c | 0.93 a | 27.80 a | 7.79 b  | 18.63 a | 1.77 a | 4.74 b |
| PA94-01       | 22.26 d | 0.83 b | 28.86 a | 13.67 a | 18.50 a | 0.92 a | 4.74 b |
| BRS Garantida | 24.05 c | 0.71 b | 34.24 a | 15.61 a | 18.10 a | 2.81 a | 4.74 b |
| Pacovan       | 28.48 a | 0.93 a | 30.62 a | 19.33 a | 16.01 a | 1.96 a | 4.85 b |
| Prata Anã     | 27.58 a | 0.67 b | 41.80 a | 14.10 a | 18.46 a | 2.34 a | 4.66 b |
| BRS Princesa  | 28.29 a | 1.10 a | 27.62 a | 7.18 b  | 18.28 a | 2.00 a | 4.71 b |
| BRS Tropical  | 22.63 d | 1.27 a | 19.00 a | 16.09 a | 17.56 a | 1.87 a | 4.73 b |
| Maçã          | 25.83 b | 1.08 a | 25.33 a | 10.24 b | 20.05 a | 1.65 a | 4.67 b |
| Caipira       | 23.31 c | 0.71 b | 34.46 a | 8.32 b  | 17.26 a | 1.87 a | 5.10 a |
| Bucaneiro     | 20.65 d | 0.67 b | 31.47 a | 14.75 a | 21.68 a | 0.88 a | 5.27 a |
| Thap Maeo     | 23.11 c | 0.79 b | 29.70 a | 10.63 b | 18.82 a | 1.78 a | 4.67 b |
| CV (%)        | 5.62    | 24.21  | 21.87   | 33.23   | 7.05    | 43.57  | 4.09   |

Averages followed by the same uppercase letter in the column belong to the same group according to the clustering method of Scott-Knott at 5% of significance. CV: coefficient of variation; SS: soluble solids; TTA: titratable total acidity; RAT: SS/TTA, RS – reducing sugars; TS: total sugars; and STA: starch

Pacovan, BRS Tropical, BRS Garantida, BRS Platina, Prata Anã, Bucaneiro, and PA94-01 genotypes formed the group with the highest

contents of reducing sugars, ranging from 13.67 to 19.3%, and the second group, formed by Thap Maeo, Maçã, FHIA-18, Caipira, YB42-47, and BRS

Princesa, ranging from 7.18 to 10.24%. Bucaneiro, Caipira and, BRS Platina genotypes presented the highest pH (5.27; 5.10, and 5.06 respectively), and FHIA-18, YB42-47, PA94-01, BRS Garantida, Pacovan, Prata Anã, BRS Princesa, BRS Tropical, Maçã, and Thap Maeo genotypes formed the second group, with pH ranging from 4.66 to 4.85. Results obtained by Bezerra and Dias (2009), Roque et al. (2014) and Castrini et al. (2015), in different environments, did not present great variations as compared with those obtained in the present study. Probably the pH is a characteristic not greatly affected by the genotype x environment interaction.

There were no significant differences between genotypes for SS/TTA ratio, total sugars, and starch. SS/TTA ratio ranged from 19.00 (BRS Tropical) to 41.80 (Prata Anã). In studies conducted by Bezerra and Dias (2009), Prata Anã genotype presented average value (28.13) quite below the one found in this study. According to reports by Chitarra and Chitarra (2005), this variable has been applied to the evaluation of taste in relationship between

soluble solids and titratable acidity. There was no significant difference between genotypes for total sugars and starch, which ranged from 21.68 to 17.26% and from 0.88 to 2.81%, respectively.

Associations between fruit length, fruit diameter, fruit mass and pulp mass in both production cycles based on phenotypic correlation are highly significant (Table 3). There was significant negative correlation of these variables with the pulp/peel ratio.

The pH presented significant negative correlation with soluble solids in second cycle and starch in second and third cycles. According Godoy (2010) pH presented moderate negative correlation with reducing sugars, unlike the titratable total acidity, that had positive correlation with these sugars. Total titratable acidity was highly negatively correlated with SS/TTA, an expected result, and relation between total sugars and starch was significant and negative.

**Table 3.** Phenotypic correlations between physicochemical characters of banana tree genotypes in the second and third production cycles, with the respective significance tests<sup>(2)</sup>.

|        | 2° Production Cycle |         |         |                      |                      |                      |                      |                      |                      |                      |                      |
|--------|---------------------|---------|---------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
|        | FD                  | MFP     | MFWP    | PPR                  | pH                   | SS                   | TTA                  | SS/TTA               | RS                   | TS                   | STA                  |
| LF     | 0.683**             | 0.878** | 0.846** | -0.236 <sup>ns</sup> | 0.364 <sup>ns</sup>  | -0.569*              | 0.035 <sup>ns</sup>  | -0.323 <sup>ns</sup> | 0.126 <sup>ns</sup>  | 0.162 <sup>ns</sup>  | 0.060 <sup>ns</sup>  |
| FD     |                     | 0.845** | 0.830** | -0.002 <sup>ns</sup> | -0.027 <sup>ns</sup> | -0.206 <sup>ns</sup> | 0.306 <sup>ns</sup>  | -0.471*              | 0.099 <sup>ns</sup>  | -0.101 <sup>ns</sup> | 0.275 <sup>ns</sup>  |
| FM     |                     |         | 0.962** | -0.137 <sup>ns</sup> | 0.213 <sup>ns</sup>  | -0.485*              | 0.138 <sup>ns</sup>  | -0.395*              | 0.203 <sup>ns</sup>  | 0.085 <sup>ns</sup>  | 0.228 <sup>ns</sup>  |
| PM     |                     |         |         | 0.104 <sup>ns</sup>  | 0.216 <sup>ns</sup>  | -0.502**             | 0.174 <sup>ns</sup>  | -0.407*              | 0.107 <sup>ns</sup>  | 0.223 <sup>ns</sup>  | 0.081 <sup>ns</sup>  |
| PPR    |                     |         |         |                      | -0.001 <sup>ns</sup> | 0.002 <sup>ns</sup>  | 0.196 <sup>ns</sup>  | -0.079 <sup>ns</sup> | -0.325 <sup>ns</sup> | 0.459*               | -0.384 <sup>ns</sup> |
| pH     |                     |         |         |                      |                      | -0.306 <sup>ns</sup> | -0.376 <sup>ns</sup> | 0.239 <sup>ns</sup>  | 0.205 <sup>ns</sup>  | 0.248 <sup>ns</sup>  | -0.138 <sup>ns</sup> |
| SS     |                     |         |         |                      |                      |                      | 0.174 <sup>ns</sup>  | 0.249 <sup>ns</sup>  | 0.034 <sup>ns</sup>  | -0.373 <sup>ns</sup> | 0.421*               |
| TTA    |                     |         |         |                      |                      |                      |                      | -0.831**             | -0.057 <sup>ns</sup> | -0.149 <sup>ns</sup> | 0.065 <sup>ns</sup>  |
| SS/TTA |                     |         |         |                      |                      |                      |                      |                      | 0.144 <sup>ns</sup>  | 0.011 <sup>ns</sup>  | 0.046 <sup>ns</sup>  |
| RS     |                     |         |         |                      |                      |                      |                      |                      |                      | -0.294 <sup>ns</sup> | 0.165 <sup>ns</sup>  |
| TS     |                     |         |         |                      |                      |                      |                      |                      |                      |                      | -0.521**             |

|     | 3° Production Cycle |         |         |                      |                     |                      |                      |                      |                      |                      |                      |
|-----|---------------------|---------|---------|----------------------|---------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
|     | FD                  | FM      | PM      | PPR                  | pH                  | SS                   | TTA                  | SS/TTA               | RS                   | TS                   | STA                  |
| LF  | 0.533**             | 0.882** | 0.838** | -0.210 <sup>ns</sup> | 0.443*              | -0.413*              | -0.010 <sup>ns</sup> | -0.164 <sup>ns</sup> | 0.327 <sup>ns</sup>  | 0.356 <sup>ns</sup>  | -0.224 <sup>ns</sup> |
| FD  |                     | 0.805** | 0.802** | 0.014 <sup>ns</sup>  | 0.194 <sup>ns</sup> | -0.239 <sup>ns</sup> | 0.344 <sup>ns</sup>  | -0.415*              | -0.041 <sup>ns</sup> | 0.157 <sup>ns</sup>  | 0.303 <sup>ns</sup>  |
| FM  |                     |         | 0.959** | -0.167 <sup>ns</sup> | 0.247 <sup>ns</sup> | -0.371 <sup>ns</sup> | 0.088 <sup>ns</sup>  | -0.258 <sup>ns</sup> | 0.180 <sup>ns</sup>  | 0.341 <sup>ns</sup>  | 0.018 <sup>ns</sup>  |
| PM  |                     |         |         | 0.101 <sup>ns</sup>  | 0.320 <sup>ns</sup> | -0.408*              | 0.202 <sup>ns</sup>  | -0.354 <sup>ns</sup> | 0.052 <sup>ns</sup>  | 0.275 <sup>ns</sup>  | 0.040 <sup>ns</sup>  |
| PPR |                     |         |         |                      | 0.245 <sup>ns</sup> | -0.160 <sup>ns</sup> | 0.360 <sup>ns</sup>  | -0.259 <sup>ns</sup> | -0.516**             | -0.181 <sup>ns</sup> | -0.033 <sup>ns</sup> |

|        |         |                      |                      |                      |                      |                      |
|--------|---------|----------------------|----------------------|----------------------|----------------------|----------------------|
| pH     | -0.445* | -0.232 <sup>ns</sup> | 0.164 <sup>ns</sup>  | 0.001 <sup>ns</sup>  | 0.294 <sup>ns</sup>  | -0.472*              |
| SS     |         | 0.385 <sup>ns</sup>  | -0.115 <sup>ns</sup> | -0.204 <sup>ns</sup> | -0.281 <sup>ns</sup> | 0.233 <sup>ns</sup>  |
| TTA    |         |                      | -0.908**             | -0.090 <sup>ns</sup> | -0.279 <sup>ns</sup> | 0.290 <sup>ns</sup>  |
| SS/TTA |         |                      |                      | -0.061 <sup>ns</sup> | 0.170 <sup>ns</sup>  | -0.242 <sup>ns</sup> |
| RS     |         |                      |                      |                      | -0.147 <sup>ns</sup> | -0.104 <sup>ns</sup> |
| TS     |         |                      |                      |                      |                      | -0.474 <sup>ns</sup> |

<sup>2)</sup> ns – non-significant; \* significant at the 5% level of probability; \*\* significant at the 1% level of probability. LF: fruit length; FD: fruit diameter; FM: fruit mass; PM: pulp mass; PPR: pulp/peel ratio; pH; SS: soluble solids; TTA: titratable total acidity; RS: reducing sugars; TS: total sugars; STA: starch.

Analysis performed criterion by Singh (1981), to estimate the relative contribution of each variable to the study of the genetic dissimilarity, indicated that major components explained 99.17 and 98.96% of total variation between the 13 genotypes in the second and third cycles, respectively (Table 4). Characters total sugars (60.56%), fruit mass (18.34%), starch (10.84%), and pulp mass (9.43%), in the second cycle, and total sugars (55.33%), fruit mass (20.27%), starch (12.67%), and pulp mass (10.69%), in the third cycle, were those that contributed the most to the total diversity between the thirteen genotypes. The

pH, SS, TTA, SS/TTA ratio, and pulp/peel ratio obtained lower relative contribution, in accordance with Godoy (2010). Greatest contribution of the same characters in both cycles can reinforce these characteristics are effective for the analysis of all genotypes. Mattos et al. (2010) observed the first three main components explained 68.30% of the total variation among the 26 banana tree accessions evaluated regarding their agronomic characteristics, and the number of fruits contributed in 83.80% to variation among the fruits. Godoy (2010) also noted high contribution of sugars and low contribution of SS/TTA ratio, pulp yield, and TTA.

**Table 4.** Relative importance (S.j.) of physicochemical characters of fruits for genetic diversity studies in 13 banana genotypes in the second and third production cycles<sup>(4)</sup>.

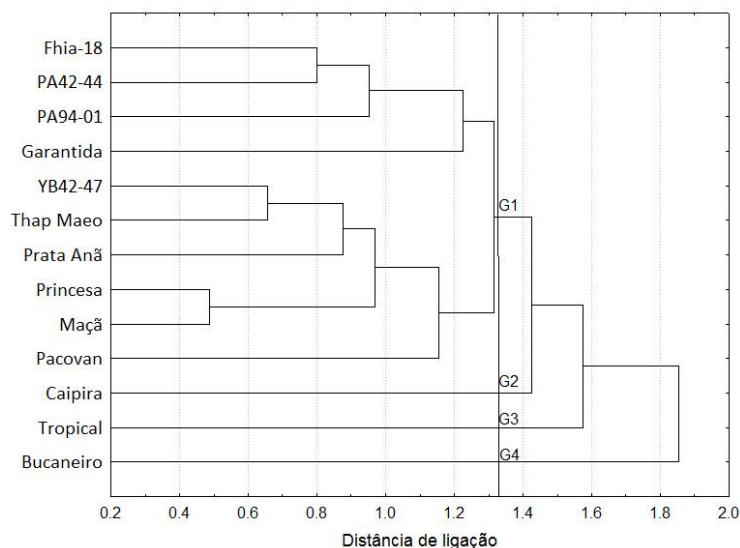
| Characters | 2° Cycle  |         | 3° Cycle  |         |
|------------|-----------|---------|-----------|---------|
|            | S.j       | S.j (%) | S.j       | S.j (%) |
| LF         | 1050.04   | 0.07    | 1006.58   | 0.10    |
| FD         | 31.06     | 0.00    | 13.16     | 0.00    |
| FM         | 289151.04 | 18.34   | 199540.78 | 20.27   |
| PM         | 148770.34 | 9.43    | 105267.60 | 10.69   |
| PPR        | 106.72    | 0.01    | 136.38    | 0.01    |
| pH         | 5.34      | 0.00    | 8.46      | 0.00    |
| SS         | 1085.40   | 0.07    | 871.04    | 0.09    |
| TTA        | 5.10      | 0.00    | 8.84      | 0.00    |
| SS/TTA     | 6560.22   | 0.42    | 6262.62   | 0.64    |
| RS         | 4159.38   | 0.26    | 1798.00   | 0.18    |
| TS         | 955110.42 | 60.56   | 544675.14 | 55.33   |
| STA        | 170968.10 | 10.84   | 124748.56 | 12.67   |

<sup>(4)</sup>S.j.: contribution of variable x to the Euclidean distance between genotypes; LF: length; of the fruit; FD: fruit diameter; FM: fruit mass; PM: pulp mass; PPR: pulp/peel ratio; pH; SS: soluble solids; TTA: titratable total acidity; RS: reducing sugar; TS: total sugar; STA: starch.

Considering the genetic dissimilarity average (1.33) of genotypes in third cycle, four groups were formed (Figure 1): G1 – FHIA-18, BRS Platina, PA94-01, BRS Garantida, YB42-47, Thap Maeo, Prata Anã, BRS Princesa, Maçã, and Pacovan; G2 – Caipira; G3 – BRS Tropical; and G4 – Bucaneiro.

The comparison of the averages, for twelve physicochemical characters under study and groups obtained, by Euclidean distance and UPGMA

clustering method, allowed to infer differences between them. All genotypes of G1 present the genome B. BRS Platina, PA94-01, and FHIA-18 hybrids are genetically closer than their progenitor, Prata Anã. Expected result considering that, overall, hybrids presented better performance than progenitor's to some features. Bucaneiro presented more divergence for being a tetraploid of genome A (G4).



**Figure 1.** Dendrogram of genetic dissimilarity of 13 banana genotypes based on Euclidean distance and UPGMA clustering method in third production cycle. Groups 1 (G1), 2 (G2), 3 (G3), and 4 (G4).

## CONCLUSIONS

There is physicochemical variability of fruits between the genotypes evaluated; the content of total sugars has greater contribution to genetic diversity, followed by fruit mass and pulp mass, and starch content in two production cycles.

Associations between fruit length, fruit diameter, fruit mass and pulp mass in two production cycles based on phenotypic correlation are highly significant.

BRS Platina, PA94-01, and FHIA-18 hybrids are genetically close, and Bucaneiro present

greater divergence for being a tetraploid of genome A.

FHIA-18 and PA94-01 hybrids, YB42-47 and BRS Tropical, BRS Princesa, and Bucaneiro cultivars are promising for recommendation on coastal plain regions.

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**RESUMO:** Genótipos melhorados necessitam ser caracterizados e avaliados em diferentes condições edafoclimáticas. Existem poucos estudos sobre a associação entre os caracteres avaliados e sua contribuição para a diversidade genética da banana, sendo imprescindíveis para orientar programas de melhoramento genético. O objetivo desse trabalho foi avaliar os caracteres físico-químicos de frutos de genótipos de bananeira, estimar as associações entre caracteres, determinar a importância relativa dos caracteres para o estudo da dissimilaridade genética e indicar novos genótipos para a região de Tabuleiros Costeiros de Sergipe. Foram utilizados frutos de treze genótipos: tipo Prata (FHIA-18, BRS Platina, PV94-01, BRS Garantida, YB42-47, Pacovan e Prata-Anã), Maçã (BRS Princesa, BRS Tropical e Maçã), Ibota (Caipira), Gros Michel (Bucaneiro) e o Mysore (Thap Maeo). O delineamento experimental foi em blocos ao acaso, com treze genótipos e três repetições. Existe variabilidade físico-química dos frutos entre os genótipos do mesmo grupo genômico e subgrupo, e entre híbridos originados da mesma genitora. O teor de açúcares totais apresenta maior contribuição para a diversidade genética entre os genótipos, seguido da massa do fruto com e sem casca e teor de amido nos dois ciclos. As associações entre comprimento do fruto, o diâmetro, massa do fruto e da polpa nos dois ciclos de produção com base na correlação fenotípica são altamente significativas. Os híbridos FHIA-18, PA94-01, YB42-47 e as cultivares BRS Tropical, BRS Princesa e Bucaneiro são promissores para recomendação nos Tabuleiros Costeiros.

**PALAVRAS-CHAVE:** *Musa* sp. Cultivares. Híbridos.

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