

Physiological and agronomic performance of common bean treated with multifunctional microorganisms

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ABSTRACT: Multifunctional microorganisms (MM) are able of colonizing root system and shoots and then, provide beneficial effects to the plants. Thus, the objective of this study was test whether multifunctional microorganisms affect gas exchange, macronutrient content, yield components and bean grain yield. A completely randomized design with twenty-six treatments and three replications was used under controlled conditions. Treatments consisted of the application of MM and its combinations in pairs, with nine rhizobacteria isolates BRM 32109, BRM 32110 and 1301 (*Bacillus* sp.), BRM 32111 and BRM 32112 (*Pseudomonas* sp.), BRM 32113 (*Burkholderia* sp.), BRM 32114 (*Serratia* sp.), 1381 (*Azospirillum* sp.) and Ab-V5 (*Azospirillum* brasiliense), an edaphic fungal isolate T-26 (*Trichoderma koningiopsis*), and a control (without MM). The isolates Ab-V5 and BRM 32112, in addition to the combinations BRM 32114 + T-26, 1301 + BRM 32110 and BRM 32114 + BRM 32110 were the highlights treatments, since they provided increases in gas exchange, in the content of macronutrients and in the agronomic performance.

Key words: bioagents; gas exchange; grain yield; nutrients; Phaseolus vulgaris

Desempenho fisiológico e agronômico do feijão-comum tratado com microrganismos

RESUMO: Microrganismos multifuncionais (MM) são capazes de colonizar o sistema radicular e a parte aérea das plantas e proporcionar efeitos benéficos ao sistema. Assim, o objetivo deste trabalho foi testar se os microrganismos multifuncionais afetam as trocas gasosas, o teor de macronutrientes, os componentes do rendimento e o rendimento de grãos do feijãocomum. O delineamento inteiramente casualizado com vinte e seis tratamentos e três repetições foi utilizado em condições controladas. Os tratamentos consistiram na aplicação de MM e em combinações aos pares, com nove isolados de rizobactérias BRM 32109, BRM 32110 e 1301 (*Bacillus* sp.), BRM 32111 e BRM 32112 (*Pseudomonas* sp.), BRM 32113 (*Burkholderia* sp.), BRM 32114 (*Serratia* sp.), 1381 (*Azospirillum* sp.) e Ab-V5 (*Azospirillum brasiliense*), um isolado fúngico edáfico T-26 (*Trichoderma koningiopsis*) e um controle (sem MM). Os isolados Ab-V5 e BRM 32112, além das combinações BRM 32114 + T-26, 1301 + BRM 32110 e BRM 32110 foram os tratamentos de destaque, pois proporcionaram aumentos nas trocas gasosas, no teor de macronutrientes e no desempenho agronômico.

Palavras-chave: bioagentes; trocas gasosas; produtividade; nutrientes; Phaseolus vulgaris

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Introduction

Common bean (*Phaseolus vulgaris* L.) is one of the most important legumes for food consumption in Brazil and worldwide (Carbonell et al., 2019; Ganascini et al., 2019). Common bean grains is considered a source of basic protein for direct consumption, low cost, excellent source of fiber, iron and amino acids (Li et al., 2017). Brazil is the world's largest producer of this legume with a cultivated area of 2,926.2 thousand ha, productivity of 1,074 kg ha⁻¹ and production of 3,144.2 thousand tons (Conab, 2021), in addition to being cultivated in almost all Brazilian states (Carbonell et al., 2019).

Smallholder's farms dominate the Brazilian production of common beans (about 67%), but there is a tendency to increase the participation of medium and large farmers, mainly in the Cerrado region (irrigated winter cultivation) (Ganascini et al., 2019). According to Carbonell et al. (2019), winter cultivation is characterized by the intensive use of synthetic inputs, like chemical fertilizers that provide significant increases in grain yield, however, it also increases production costs.

In view of the serious consequences of the intensive use of agricultural inputs has caused to the agro-ecosystems, such as air and water pollution, soil contamination and human health problems, the idea of environmental conservation and the rational use of its resources has become increasingly relevant (Ekin, 2019). As an example, there are the specific interactions between plants and multifunctional microorganisms, which have a positive impact on the ecosystems, such as nitrogen fixation, phosphate solubilization and microbiolization, production of phytohormones, biofilm, siderophores, biological control and maintenance of the food chain and biogeochemical cycles (Braga et al., 2016).

In this context, rhizobacteria, able of rapidly colonize root system of plants, represent strategic organisms in the soilplant system because in addition to preventing the invasion of pathogens through the production of secondary metabolites (Oliveira et al., 2016) also help the plants to growth and develop better (Ahemad & Kibret, 2014). Among the positive effects of these bacteria there are: N_2 fixation (Damam et al., 2016), production of phytohormones (Sureshbabu et al., 2016), increased phosphate solubilization (Paiter et al., 2019), stimulus to the greater root development of plants (Aquino et al., 2018), increased absorption of water and nutrients (Sureshbabu et al., 2016), control of phytopathogens (Singh et al., 2019), among others that result in greater plant development and greater grain yield in contrast to nontreated plants (Ekin, 2019).

In addition to rhizobacteria, the soil fungus, *Trichoderma* spp., also plays a major role in plant growth (Aquino et al., 2018). Its effect has been related to the protection of plants against primary and secondary rhizosphere pathogens, production of plant growth hormones, increased absorption and translocation of mineral nutrients, in addition to increased solubility and availability of many nutrients (Montesinos et al., 2019).

Additionally, the co-inoculation with rhizobacteria has been proposed as a technology able of enhancing the promotion of growth in plants and, consequently, benefiting the N_2 fixation process and stimulating the increase in the absorption of water and nutrients, compared to the application of individual isolates (Korir et al., 2017). However, despite of the potential for the use of multifunctional microorganisms in agricultural crops, mainly in co-inoculation, there are still few works published in the crop of common bean.

Thus, we tested whether multifunctional microorganisms affect gas exchange, macronutrient content, yield components and grain yield of common bean plants.

Materials and Methods

Characterization of the environment

The experiment was conducted in a greenhouse, at the headquarters of Embrapa Rice and Bean, Santo Antônio de Goiás, GO. Soil from the arable layer (0.00-0.20 m) of a dystrophic Oxisol, medium texture, was used. The chemical characteristics of the soil were determined with the following results pH (H_2O) = 5.6; Ca² + = 28.4 mmol_c dm-³; Mg²⁺ = 12.5 mmol_c dm-³; H + Al³⁺ = 28 mmol_c dm-³; P = 11 mg dm-³; K + = 218 mg dm-³; Cu²⁺ = 0.8 mg dm-³; Zn²⁺ = 4.0 mg dm-³; Fe³⁺ = 7.3 mg dm-³; Mn²⁺ = 18 mg dm-³ and organic matter = 37.7 g kg⁻¹.

Three weeks before the sowing of common bean, 78 pots with a capacity of 7 kg were completely filled with soil and fertilized with 10 g of NPK, formulation 5-30-15. During the entire conduct of the experiment, soil moisture was monitored daily, and kept close to field capacity conditions.

Experimental design and treatments

A completely randomized design with 26 treatments and three replications was used. Treatments consisted of the application of microorganisms and their combinations in pairs in common bean, with nine rhizobacteria isolates and one fungus isolated from soil (Table 1). Microorganisms were applied at three times in the crop, microbiolization of the seeds and sprayed of microorganisms solution at 7 (soil) and 21 (plants) days. In the control treatment, water was used both for microbiolization, as well as spraying at 7 and 21 days.

The main characteristics of the rhizobacteria isolates BRM 32109 (*Bacillus* sp.), BRM 32110 (*Bacillus thunringiensis*), BRM 32111 (*Pseudomonas fluorescens*), BRM 32112 (*Pseudomonas* sp.), BRM 32113 (*Burkholderia pyrrocinia*), BRM 32114 (*Serratia* sp.), 1301 (*Bacillus* sp.), 1381 (*Azospirillum* sp.), Ab-V5 (*Azospirillum brasilense*) and the fungus T-26 (*Trichoderma koningiopsis*) are described in Table 2. The microorganisms used are stored and preserved in the Multifunctional Microorganisms collection of Embrapa Rice and Bean.

Microbiolization of seeds

Each bacterium was grown in solid medium (nutrient agar), suspensions with the bacteria were prepared in liquid medium 523 (nutrient broth) (Kado & Heskett, 1970), in a shaking incubator, for 24 hours at 28 °C. The suspension concentration

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Treatments	Microorganisms	Treatments	Microorganisms
1	BRM 32109 (<i>Bacillus</i> sp.)	14	1301 + 32114
2	BRM 32110 (Bacillus thunringiensis)	15	1381 + 32114
3	BRM 32111 (Pseudomonas fluorescens)	16	1381 + 1301
4	BRM 32112 (Pseudomonas sp.)	17	32110 + Ab-V5
5	BRM 32113 (Burkholderia pyrrocinia)	18	32114 + Ab-V5
6	BRM 32114 (<i>Serratia</i> sp.)	19	1301 + Ab-V5
7	T-26 (Trichoderma koningiopsis)	20	1381 + Ab-V5
8	Ab-V5 (Azospirillum brasilense)	21	Ab-V5 + T-26
9	1381 (<i>Azospirillum</i> sp.)	22	32110 + T-26
10	1301 (<i>Bacillus</i> sp.)	23	32114 + T-26
11	32114 + 32110	24	1301 + T-26
12	1301 + 32110	25	1381 + T-26
13	1381 + 32110	26	Control

Table 2. Collection code, geographical origin, biochemical characteristics and taxonomic classification of the nine rhizobacteria isolates used in this study.

Biochemical ^D									
Code ^A	Origin ^B	Color ^c	AIAF	Cel. ^G	Phos ^H	Sid ⁱ	Biofilm ^J	Taxonomic ^E	
BRM 32109	GO/Brazil	White	-	+	+	+	+	Bacillus sp.	
BRM 32110	PA/Brazil	White	-	+	+	-	+	Bacillus thunringiensis	
BRM 32111	PA/Brazil	Yellow	-	+	+	+	+	Pseudomonas fluorescens	
BRM 32112	GO/Brazil	Yellow	-	+	+	+	+	Pseudomonas sp.	
BRM 32113	PA/Brazil	Pink	+	+	-	+	+	Burkholderia pyrroiynia	
BRM 32114	PA/Brazil	Pink	+	+	+	+	+	Serratia sp.	
Ab-V5	PR/Brazil	Yellow	+	-	-	+	+	Azospirillum brasilense	
1381		White	+	-	-	-	-	Azospirillum sp.	
1301		White	-	-	+	-	-	Bacillus sp.	

^ANumeric code of rhizobacteria and fungus isolates in the Embrapa Rice and Bean collection of Multifunctional Microorganisms.

^BGeographic source of each isolate.

^{C, D, E}Colony color, biochemical characterization and taxonomic classification of each isolate, described by Nascente et al. (2017).

^fProducer of indolacetic acid, Gproducer of cellulase, Hproducer of phosphatase, Iproducer of siderophores, ^JProducer of biofilm. The methodology is described in Nascente et al. (2017).

of each bacterium was adjusted in a spectrophotometer to A540 = 0.5, which corresponds to 1×10^8 colony-forming units (CFU) per mL. The *Trichoderma koningiopsis* was grown in PDA culture medium, then multiplied in rice grain substrate and the suspension concentration corresponds to 1×10^8 ml⁻¹ of conidia. The seeds of common bean were immersed in each suspension of microorganisms, and the seeds of the control treatment were immersed in distilled water, for a period of 2 hours under constant agitation at 25°C.

Watering and spraying were carried out with the suspensions of microorganisms in the same concentrations used to microbiolize the seeds. Thus, at 7 days after sowing (DAS), the soil was watered with 100 ml of suspension in each treatment and at 21 DAS the plants were sprayed with 30 ml of suspension for each treatment, following the methodology of Silva et al. (2020). The plants were sprayed using a manual sprayer with pressure supplied by a CO_2 pressure source and a conical nozzle (TX-VS2).

Management of common bean plants

Ten common bean seeds cultivar BRS FC 402 were sown per pot. Ten days after germination, the plants were thinned, leaving three plants per pot. Cultural practices were carried out according to the recommendations and needs of the crop.

Assessments

Gas exchange measurements: photosynthetic rate (µmol $CO_2 m^{-2} s^{-1}$), transpiratory rate (mmol $H_2O m^{-2} s^{-1}$), stomatal conductance (mol $H_2O m^{-2} s^{-1}$), internal CO_2 concentration (vpm) and leaf temperature (°C) were determined in common bean plants at 37 DAS (beginning of flowering), using a portable gas meter in the IRGA infrared region (LCpro +, ADC BioScientific), between 08:00 and 10:00 am. The readings were taken in the central third of the first completely expanded leave (from top to bottom). The equipment was regulated to use concentrations of 370 - 400 mol mol⁻¹ CO_2 , and 1200 µmol [quanta] m⁻² s⁻¹ of photosynthetically active photon flux density (DFFFA). The minimum equilibrium time established for the readings was 2 minutes.

Yield components and grain yield: harvesting was carried out when common bean plants reached physiological maturity, 83 DAS. Thus, the number of pods per pot, the number of grains per pod was counted and the mass of 100 grains and grain yield were determined. The values obtained were corrected for 13% humidity. These determinations were made in each per pot.

Nutrient content: after drying and weighing the dry biomass of the shoots and roots, representative aliquots were removed for grinding and determination of the levels of macronutrients N, P and K.

Statistical analysis

It was determined normality of errors (Shapiro-Wilk test) and homogeneity of variance (Bartlett test) of the data. Then, the data were subjected to analysis of variance and, when significance was detected, the means were compared by the LSD test ($p \le 0.05$). The SISVAR statistical package was used to process these data.

Additionally, a multivariate principal component analysis (PCA) was performed to describe the correlation between response variables (gas exchange, accumulation of nutrients in the shoots and roots, yield components and grain yield) and isolated and combined microorganisms. The main components (PCs) were loaded with response variables when the correlation test produced $r \ge 0.50$. The first five PCs responsible for 68.2% of the data variation were maintained. Biplots (two-dimensional graph) using these five PCs that correlate isolated and combined microorganisms and response variables were built with the "FactoExtra" package on the R platform.

Results and Discussion

Gas exchange evaluation

The effect of inoculation on common bean plants on photosynthesis (A), transpiration (E), stomatal conductance (gs), internal CO_2 concentration (Ci) and leaf temperature (Tleaf) is shown in Figure 1.

The increase in E (16.5% on average) and gs (25.0% on average) in common bean plants inoculated with Ab-V5 (E and gs); and the combinations BRM 32114 + Ab-V5 (E and gs), 1381 + 1301 (gs) and 1301 + T-26 (E) indicate that there was a greater flow of water and nutrients from the soil to the plant, since the supply water for the crop was abundant (Figure 1). This is advantageous since the higher nutrient content within the plant culminates in a more adequate metabolism and, consequently, accumulation of photoassimilates and growth gain.

According to Silva et al. (2017), a higher transpiratory rate is attributed to a greater stomatal opening, which in turn can be induced by multifunctional microorganisms. On the other hand, greater stomatal opening decreases stomatal resistance to CO_2 diffusion, favoring its mesophilic conductance and, consequently, the production of photoassimilates. Although no difference in A was observed between inoculated and noninoculated plants, the photosynthetic rate of common bean plants is considered high, with emphasis on plants inoculated with Ab-V5, BRM 32113 and the combinations 1301 + T-26 and 1301 + BRM 32110 (Figure 1A).

Changes in the intercellular concentration of CO_2 (Ci), found in common bean plants inoculated with the combinations Ab-V5 + T-26, 1301 + Ab-V5, BRM 32110 + Ab-V5 and BRM 32114 + T-26 (Figure 1E), suggest that there may have been an increase in Rubisco's carboxylation activity by regulating



* Treatments followed by the same letter in columns do not differ from each other by the LSD test ($p \ge 0.05$). ** Treatments: (1) BRM 32109; (2) BRM 32110; (3) BRM 32111; (4) BRM 32112; (5) BRM 32113; (6) BRM 32114; (7) T-26; (8) Ab-V5; (9) 1381; (10) 1301; (11) BRM 32114 + BRM 32110; (12) BRM 32114 + Ab-V5; (13) BRM 32114 + T-26; (14) BRM 32110 + Ab-V5; (15) BRM 32110 + T-26; (16) Ab-V5 + T-26; (17) 1381 + 1301; (18) 1381 + Ab-V5; (19) 1301 + Ab-V5; (20) 1381 + BRM 32114; (21) 1301 + BRM 32114; (22) 1381 + BRM 32110; (23) 1301 + BRM 32110; (24) 1381 + T-26; (25) 1301 + T-26; (26) control (without microorganism).

Figure 1. (A) Photosynthesis; (B) Transpiration; (C) Stomatal conductance; (D) Leaf temperature and (E) Internal CO2 concentration in common bean plants treated with multifunctional microorganisms, isolated or in combination.

the stomatal opening that favors the fixation of mesophilic CO_2 (Shi et al., 2010). These benefits acquired with the use of multifunctional microorganisms can result in greater production of photoassimilates, which can be allocated to heterotrophic organs of common bean plants to sustain growth or be converted into reserve products or have been exuded by the roots and used by the rhizobacteria themselves. In upland rice, Nascente et al. (2017) found values significantly higher than the control treatment for the variables photosynthesis, transpiration and stomatal conductance, while there was no statistical difference for internal CO_2 concentration and leaf temperature.

Macronutrients in shoots and roots

Common bean plants treated with isolates BRM 32112, 1381, BRM 32110, BRM 32114, BRM 32109, BRM 32111, Ab-V5, BRM 32113, T-26 and 1301, in addition to the combinations BRM 32114 + T-26, 1301 + BRM 32114, 1301 + Ab-V5, 1381 +1301, BRM 32110 + T-26, BRM 32114 + BRM 32110 and 1301 + BRM 32110 presented the highest levels of N in their shoots. The average increase was around 3.70% (Figure 2A).

In relation to macronutrient P, common bean plants treated with isolates BRM 32111, BRM 32112, BRM 32114, 1381, BRM 32110 and Ab-V5 and with the combination BRM 32114 + BRM 32110, showed significantly higher levels compared to control plants (Figure 2B). The average increase was of the order of 9.23%. As for macronutrient K, common bean plants treated with the BRM 1301 + Ab-V5 combination and the BRM 32113 and 1301 isolates showed significantly higher levels (11.02%) when compared to the control plants (Figure 2C). Regarding the levels of macronutrients in the roots of common bean plants, the levels of N and K were similar between treatments; while the P content was significantly higher in common bean plants treated with the combinations 1381 + 1301, BRM 32114 + T-26 and 1301 + BRM 32110, compared to control plants (Figures 2D, 2E and 2F). The average increase in the P content was around 14.68%.

Multifunctional microorganisms can benefit plants through their ability to solubilize some nutrients in the soil solution and, thus, make them available to the plant to increase the growth rate (Chaudhary & Sindhu, 2016). As observed in the present study, multifunctional microorganisms induced the accumulation of N, P, K in the shoots and K in the roots, positively influencing the growth of common bean plants. These results may be related, since the transpiration of the plant is fundamental for the absorption of water and mineral nutrients from the soil. The results show that the multifunctional microorganisms Ab-V5, BRM 32114 + Ab-V5 and 1301 + T-26 provided increases in transpiration of common bean plants.

Regarding P, our results corroborate those of Braga Júnior et al. (2017), where several rhizobacteria tested,



* Treatments followed by the same letter in columns do not differ from each other by the LSD test ($p \ge 0.05$). ** Treatments: (1) BRM 32109; (2) BRM 32110; (3) BRM 32111; (4) BRM 32112; (5) BRM 32113; (6) BRM 32114; (7) T-26; (8) Ab-V5; (9) 1381; (10) 1301; (11) 32114 + 32110; (12) 32114 + Ab-V5; (13) 32114 + T-26; (14) 32110 + Ab-V5; (15) 32110 + T-26; (16) Ab-V5 + T-26; (17) 1381 + 1301; (18) 1381 + Ab-V5; (19) 1301 + Ab-V5; (20) 1381 + 32114; (21) 1301 + 32114; (22) 1381 + 32110; (23) 1301 + 32110; (24) 1381 + T-26; (25) 1301 + T-26; (26) control (without microorganism).

Figure 2. (A) Nitrogen (shoot); (B) Phosphorus (shoot); (C) Potassium (shoot) and (D) Nitrogen (root); (E) Phosphorus (root); (F) Potassium (root) in common bean plants treated with multifunctional microorganisms, isolated or in combination.

alone, provided higher phosphorus content in the shoots of inoculated plants than in uninoculated plants, both under natural phosphate fertilization and without using fertilizer. Therefore, the isolates BRM 32110, BRM 32111, BRM 32112 and BRM 32114, in addition to the combinations, BRM 32114 + BRM 32110, 1301 + BRM 32110 and BRM 32114 + T-26 favored the solubilization of P in the soil solution and, consequently, accumulation of P in the shoots and root of common bean plants. This can be explained due to the ability of these microorganisms to solubilize and mineralize inorganic phosphorus through organic acids, release of H⁺ protons, aid of ATPases and production of chelators.

In addition, these microorganisms can favor the increase in the accumulation of nutrients in plants, promoting root development, which allows greater access and absorption of nutrients and improving the absorption mechanisms (Ahemad & Kibret, 2014). Isolates 1381 and BRM 32113 and the combination 1301 + Ab-V5 provided the highest K accumulation in the shoots. This suggests that the high accumulation of K in common bean plants may have been due to the improvement of the K absorption mechanism or to the greater development of roots by microorganisms.

Yield components and grain yield

For yield components, the increase in the number of pods per plant (47.4%) was highlighted in plants inoculated with Ab-V5, 1381+ BRM 32114 and BRM 32114 + BRM 32110, followed by BRM 32111, BRM 32114 + T -26, 1381 + AB-V5, 1381 + BRM 32110 and 1301 + BRM 32110 (Figure 3A). The increase in the number of grains per pod obtained in common bean plants inoculated with the isolate BRM 32112 was double compared to the control (Figure 3B). In relation to the increase in the mass of 100 grains (10.8%), most treatments based on bioagents showed a beneficial effect, with emphasis on common bean plants inoculated with the isolate BRM 32114 and the combination BRM 32110 + T -26 (Figure 3C).

In this study, the bioagents that stood out to stimulate increased grain yield in common bean plants were Ab-V5, 1381+ BRM 32114 and BRM 32114 + BRM 32110 (Figure 3D). Common bean plants inoculated with bioagents that differed significantly from non-inoculated plants showed an average grain yield increase equal to 66.5%. In crops, such as common bean, where higher grain yield are the most important goal, the main criteria for selecting bioagents are those that lead to greater grain yield.

These results of higher grain yield in the microorganisms treatments instead of no-treated plants, corroborate those found by Yadegari et al. (2008), who reported that treatment with rhizobacteria significantly increased the number of pods per plant, the number of grains per pod, mass of 100 grains, mass of grains per plant and mass of pods per plant, with the co-inoculation with *Rhizobium* and rhizobacteria showed a significant increase in common bean yield and yield components.

The study shows that the highlighted microorganisms were the isolated Ab-V5 (*Azospirillum brasilense*) and BRM 32112 (*Pseudomonas* sp.), had a positive influence of *Azospirillum brasilense* on stomatal conductance, transpiration, number of pods per pot, grain yield and N and content P in the shoots of common bean plants and *Pseudomonas* sp. in the number of grains per pot, grain yield and N and P content in the shoots of common bean plants. Bacteria of the genus *Azospirillum* sp. are able to promote plant growth through biosynthesis and release of amino acids, indoacetic acid, cytokinins, gibberellins and other polyamines, favoring root growth and, consequently, intensifying the absorption of water and nutrients by plants. In addition to fixing atmospheric nitrogen (N₂) through the biological nitrogen fixation process (Zeffa et



* Treatments followed by the same letter in columns do not differ from each other by the LSD test (p ≥ 0.05). ** Treatments: (1) BRM 32109; (2) BRM 32110; (3) BRM 32111; (4) BRM 32112; (5) BRM 32113; (6) BRM 32114; (7) T-26; (8) Ab-V5; (9) 1381; (10) 1301; (11) 32114 + 32110; (12) 32114 + Ab-V5; (13) 32114 + T-26; (14) 32110 + Ab-V5; (15) 32110 + T-26; (16) Ab-V5 + T-26; (17) 1381 + 1301; (18) 1381 + Ab-V5; (19) 1301 + Ab-V5; (20) 1381 + 32114; (21) 1301 + 32114; (22) 1381 + 32110; (23) 1301 + 32110; (24) 1381 + T-26; (25) 1301 + T-26; (26) control (without microorganism).

Figure 3. (A) Number of pods per pot; (B) Number of beans per pod; (C) Mass of 100 grains and (D) Grain yield common bean plants treated with multifunctional microorganisms, alone or in combination.

al., 2019). The group of *Pseudomonas* sp. they promote plant growth either directly through phosphorus solubilization and production of siderophores or indirectly, with their ability to control plant diseases (Widnyana & Javandira, 2016).

The combinations BRM 32114 + T-26 (Serratia sp. + Trichoderma koningiopsis), BRM 1301 + BRM 32110 (Bacillus sp. + Bacillus thunringiensis) and BRM 32114 + BRM 32110 (Serratia sp. + Bacillus thunringiensis) are also featured in this study. These combinations positively influenced the number of pods per pot, grain yield and N content in the shoots. In addition, the BRM 32114 + T-26 and BRM 1301 + BRM 32110 combinations influenced the P content in the root and the internal CO₂ concentration, while the BRM 32114 + BRM 32110 influenced the P content in the shoots of common bean plants.

The genus *Serratia* sp. associated with plants, promotes plant growth by various mechanisms, such as indolacetic acid production, nitrogen fixation, 1-aminocyclopropane-1-carboxylate deaminase activity, phosphate solubilization, siderophores production and systemic induction host plant resistance (Martínez et al., 2018). *Bacillus* sp. can promote plant growth through the production of phosphatase and phosphorus solubilization and is also frequently used in biological control (Widnyana & Javandira, 2016). *Trichoderma* sp. it is known to be efficient in biological control, but recently it has been identified as a promoter of plant growth through the production of phytohormones such as auxins, cytokinins, abscisic acid, gibberellins, among others (Guzmán et al., 2018).

Principal component analysis

Regarding the analysis of the main components, it was found that the variability of treatments (isolated and combined microorganisms) observed in gas exchange (photosynthesis, transpiratory rate, stomatal conductance, internal CO_2 concentration and leaf temperature), in the yield components (number of pods per pot, number of grains per pot and mass of 100 grains) and in grain yield and nutrient accumulation (N, P and K), both in shoots and in the roots, was better explained by five main components (PCs), representing 72.26% of the data variation, that is, PC1 (22.08%), PC2 (18.94%), PC3 (14.86%), PC4 (8.89%) and PC5 (7.49) (Figure 4).

The factor map (biplot) shows groups of variables (arrows) indicating positive and negative correlations with each main component (PC), with the length of the arrow indicating the magnitude of each response for each PC (Figure 4). PC1 was positively correlated with grain yield, number of pods per pot and stomatal conductance. On the other hand, PC2 was positively correlated with transpiration, stomatal conductance, internal CO₂ concentration and leaf temperature. While PC3 was positively correlated with photosynthesis and K at the roots while PC5 correlated positively with P at the roots.

The variables of K in the roots, grain yield, number of pods per pot and K in the shoots (69.6, 66.3, 65.7, and 64.2%, respectively) presented the greatest contribution to PC1. For PC2, transpiration, stomatal conductance, internal CO_2



Treatments isolates: (1) BRM 32109; (2) BRM 32110; (3) BRM 32111; (4) BRM 32112; (5) BRM 32113; (6) BRM 32114; (7) T-26; (8) Ab-V5; (9) 1381; (10) 1301 (red), in combination: (11) 32114 + 32110; (12) 32114 + Ab-V5; (13) 32114 + T-26; (14) 32110 + Ab-V5; (15) 32110 + T-26; (16) Ab-V5 + T-26; (17) 1381 + 1301; (18) 1381 + Ab-V5; (19) 1301 + Ab-V5; (20) 1381 + 32114; (21) 1301 + 32114; (22) 1381 + 32110; (23) 1301 + 32110; (24) 1381 + T-26; (25) 1301 + T-26 (blue); control (26) without microorganisms (green).

Figure 4. Principal component analysis (PCA) explaining correlations among variables and among 26 treatments with individual and combined microorganisms. Five main components (PCs) accounted for 72.26% of the data variation.

concentration and temperature (67.9, 64.6, 62.4, and 62.0%, respectively) provided the greatest contribution. While for PC3, N and P in the shoots (91.0 and 81.7%), for PC4, photosynthesis and K in the roots (40.8 and 40.4%) and for PC5, P in the roots (50.9%) were the variables that provided the greatest contributions.

Based on the representational quality of treatments with isolated microorganisms and in combination for the variables analyzed, treatments 8 (Ab-V5) and 20 (1381 + BRM 32114) showed the greatest positive correlation for the mass of 100 grains and the number of pods per pot, respectively (Figure 4). The transpiration and stomatal conductance variables were positively correlated with treatments 14 and 19 (BRM 32110 + Ab-V5 and 1301 + Ab-V5). The accumulation of P in the shoots was positively correlated with treatment 4 (BRM 32112) and the accumulation of P in the roots with treatment 6 (BRM 32113) and the accumulation of P at the roots with treatment 13 (BRM 32114 + T-26).

Therefore, it appears that multifunctional microorganisms significantly influenced the development of common bean plants such as gas exchange, yield components and grain yield and accumulation of macronutrients in the shoots and roots, and this resulted in more productive plants.

Conclusions

This study describes aspects of the effects of multifunctional microorganisms on common bean plants, providing a better understanding of the physiological and agronomic changes that occur in inoculated plants. Events such as gas exchange and macronutrient content in the shoots and roots were increased so that the plants could achieve increased yield components and grain yield.

The outstanding bioagents were *Azospirillum brasilense* (Ab-V5); *Pseudomonas* sp. (BRM 32112), in addition to the combinations *Serratia* sp. + *Trichoderma koningiopsis* (BRM 32114 + T-26), *Bacillus* sp. + *Bacillus thunringiensis* (1301 + BRM 32110) and *Serratia* sp. + *Bacillus thunringiensis* (BRM 32114 + BRM 32110). In addition, the results show an innovation in relation to a microorganism, however, complementary studies under field conditions are needed. The genus *Trichoderma koningiopsis* is important for the control of nematodes, but little studied as a growth promoter; however, the results show this genus as a highlight, providing an increase in gas exchange, accumulation of nutrients and components of yield and grain yield.

Compliance with Ethical Standards

Author contribution: Conceptualization: RCC, NAS, FMCC, LAC; Data curation: RCC, NAS, SJFA; Formal analysis: RCC, FLLM, SMA, PRAC; Funding acquisition: NAS, FMCC; Investigation: RCC, NAS, FMCC, LAC; Methodology: RCC, NAS, FMCC, LAC, SJFA; Project administration: RCC, NAS; Resources: NAS, FMCC, LAC; Software: No programs were developed; Supervision: NAS, FMCC, LAC; Validation: RCC, NAS, FMCC, LAC; Visualization: RCC, FLLM, SMA, NAS, FMCC, LAC; Writing – original draft: RCC, SMA, NAS; ML; Writing – review & editing: RCC, NAS, FMCC, LAC.

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