

Morpho-physiological and anatomical characteristics of *Urochloa brizantha* cv. Marandu in silvopastoral and monoculture systems

Luciano Fernandes Sousa^{1*}, Guilherme Rocha Moreira², José Pires de Lemos Filho³, Domingos Savio Campos Paciullo⁴, João Maurício Bueno Vendramini⁵, Robert Emilio Mora Luna⁶ and Rogério Martins Maurício⁷

¹Escola de Medicina Veterinária e Zootecnia, Universidade Federal do Tocantins, Br-153, Km 112, Zona Rural, 77804-970, Araguaína, Tocantins, Brazil.

²Departamento de Estatística e Informática, Universidade Federal Rural de Pernambuco, Recife, Pernambuco, Brazil. ³Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Minas Gerais, Brazil. ⁴Centro Nacional de pesquisa em gado de Leite, Empresa Brasileira de Pesquisa Agropecuária, Juiz de fora, Minas Gerais, Brazil. ⁵Range Cattle Research and Education Center, Institute of Food and Agricultural Sciences, University of Florida, Ona, Florida, USA. ⁶Departamento de Ciencias Animales, Facultad de Agronomía e Ingeniería Forestal, Pontificia Universidad Católica de Chile, Santiago, Chile. ⁷Departamento de Engenharia de Biosistemas, Universidade Federal de São João Del Rey, São João Del Rey, Minas Gerais, Brazil. *Author for correspondence. E-mail: luciano.sousa@uft.edu.br

ABSTRACT. To gain insights into the forage morphological and anatomical characteristics in a silvopastoral system (SPS) with Bolsa de Pastor (*Zeyheria tuberculosa*) and palisadegrass 'Marandu' (*Urochloa brizantha*) monoculture (MONO). The SPS was established through natural regeneration of the tree species. Treatments were a SPS and MONO distributed in a completely randomized design with six replicates and repeated measures were the harvest periods. Response variables were morpho-physiological and anatomical characteristics: green: dead material ratio, leaf blade: stem+sheath ratio, leaf area index, chlorophyll and carotenoid concentrations, proportions of non-lignified and achlorophyllous areas, lignified areas in stems, proportions of non-lignified and achlorophyllous areas, lignified and chlorophyllous areas in leaves, as well as cell length in longitudinal section of stem. Morpho-physiological patterns were altered ($p < 0.05$) under natural shading conditions due to higher photosynthetic efficiency in the SPS. There was no effect ($p > 0.05$) of the systems on anatomical patterns, proportions of non-lignified and achlorophyllous, lignified and chlorophyllous tissues, these proportions were influenced only by the periods of the year, both for stems and leaves. Cells of the internodes of the grasses of the studied systems had the same length. The SPS alters morpho-physiological characteristics of palisadegrass and increases the concentration of chlorophyll *a* and *b*.

Keywords: chlorophyll; pastures; plant physiology; shading.

Received on June 1, 2021.
Accepted on February 14, 2022.

Introduction

In Brazil, livestock systems are predominantly based on grass monocultures and relies on fertilizer and herbicides to be feasible. Conversely, the lack of inputs and mismanagement may result in soil degradation and destruction of the remaining native vegetation (Dias-Filho, 2014). Silvopastoral systems (SPS) are an alternative method to restore degraded pastures and diversify the income in livestock systems (Jose & Dollinger, 2019; Murgueitio, Chará, Barahona, & Rivera, 2019). Silvopastoral system consist of combinations of trees and shrubs with pastures and animals in the same area, simultaneously or staggered over time (Jose, Walter, & Mohan Kumar, 2019). Palisadegrass (*Brachiaria brizantha* [Hochst. ex A. Rich.] R. Webster cv. Marandu) is the most cultivated warm-season perennial grass in Brazil (Jank, Barrios, Do Valle, Simeão, & Alves, 2014; Ministério da Agricultura, Pecuária e do Abastecimento [MAPA], 2018). Palisadegrass is widely used due to its adaptation on different soil and climatic conditions, and is commonly used in SPS (Gomes et al., 2019; Oliveira et al., 2021).

According to Malaviya, Baig, Kumar, and Kaushal (2020) and Cruz et al. (2021), leaves of plants growing under conditions of low light, such as forages grown in SPS have greater concentration of chlorophylls than those growing in full sunlight. Moreover, such environmental conditions may modify other chemical, morphological, and

productive characteristics of forage tissues under either natural (Gomes et al., 2019, 2022; Santos et al., 2018) or artificial (Guenni, Romero, Guédez, Bravo de Guenni, & Pittermann, 2018; Pang et al., 2019a; Pang et al., 2019b) shading conditions. These changes in morpho-physiological and anatomical patterns can modify herbage mass and nutritive value, which may affect stocking rates, voluntary intake, and animal performance of livestock grazing in SPS (Paciullo et al., 2021; Silva et al., 2021a; Silva et al., 2021b; Sousa et al., 2015).

The objective of this study was to evaluate morpho-physiological and anatomical characteristics of palisadegrass, under the influence of the tree species Bolsa de Pastor (*Zeyheria tuberculosa*) in a tropical region.

Material and methods

Experimental area and location

The experiment was conducted in a SPS located in Lagoa Santa, state of Minas Gerais, Brazil (19°35'36"S, 43°51'56"W; 747 m altitude). The soil at the experimental site is classified as ferralsols in the European Soil Classification System (Tóth et al., 2008) and the chemical properties before the initiation of the study were: pH (H₂O) = 4.72 cmol_c·dm⁻³, P = 1.99 (mg·dm⁻³), Ca²⁺ = 1.61 cmol_c·dm⁻³, Mg²⁺ = 1.46 cmol_c·dm⁻³, K = 1.25 cmol_c·dm⁻³, SB = 4.41 cmol_c·dm⁻³, CEC = 10.69 cmol_c·dm⁻³, OM = 33.82 g·kg⁻¹.

Treatments consisted of the SPS or a monoculture of palisadegrass (MONO) distributed in a completely randomized design with six replicates. Six paddocks (16 m² each; experimental unit) were established and fenced. The Bolsa de Pastor trees were established in 1982 from natural regeneration of trees. The excessive trees and undesirable species were eliminated and the remaining tree stand was oriented to maintain a minimum distance of 4 m between trees, which resulted in a density of 160 trees·ha⁻¹ (Viana, Maurício, Matta-Machado, & Pimenta, 2002). The vegetation of the area used for the MONO was removed and the area was prepared for sowing. Based on soil analysis results, phosphate rock and dolomitic limestone were manually applied at 1.2 and 1.5 t·ha⁻¹, respectively. Palisadegrass was planted in 1990 and seeds were manually sown at a seeding rate of 10 kg·ha⁻¹ viable seeds and 2 cm depth. From the establishment until the beginning of this study, pastures were used to feed beef and dairy cattle. Rainfall data were collected at 10 km from the experimental area. Seven harvestings were made over one year, as shown in Table 1.

Table 1. Experimental periods and rainfall (mm) during the trial.

Period	Rainfall pattern	Date	Days	Rainfall (mm)
P1	Wet	(Nov. 24 to Dec. 25)	31	325.0
P2	Wet	(Dec. 26 to Jan. 25)	31	185.5
P3	Wet	(Jan. 26 to Feb. 25)	31	259.7
P4	Wet	(Feb. 26 to Mar. 28)	31	157.3
P5	Transition from wet to dry season	(Mar. 29 to Apr. 28)	31	88.4
P6	Dry	(Apr. 29 to Jul. 28)	92	11.7
P7	Transition from dry to wet season	(Jul. 29 to Nov. 23)	118	99.1
Total	----	----	365	1126.7

Characteristics evaluated

In November 2008, forage was manually harvested with a cleaver at 30 cm stubble height at the initiation of the experimental period. Canopy height within the sampling area was measured according to Almeida, Maraschin, Harthmann, Ribeiro Filho, and Setelich (2000) in four points per experimental unit using a graduated ruler. Three 1 m² samples per experimental unit were harvested using a metal frame (Paladines, 1992), thereafter with a 31 days regrowth interval during the growing season. In the dry season, forage was harvested twice in July and November due to limited herbage mass. The remaining forage was harvested at the same stubble height and removed from the experimental unit, after sample collection. The harvested forage was weighed, dried in a forced-air oven at 55°C (Association of Official Analytical Chemists [AOAC], 1990) and used for forage mass calculation. Two subsamples were taken and according to Chacón, Stobbs, and Haydock (1977), green and dead material, leaf blade (Lb), and stem + sheath (SS) were quantified, and green: dead and Lb:SS ratios were calculated.

Sections with 2 cm² of the middle third of the leaf blade from fully expanded green leaves were harvested, weighed, and extracted in 80% acetone for measuring the chlorophyll and carotenoid concentrations. Extracts obtained were filtered through fast filter paper and stored in test tubes in a dark environment. Subsequently, the optical density of filtrates was read at 663, 645, and 470 nm using a Genesys 10S[®] ultraviolet/visible spectrophotometer (Thermo Fisher

Scientific, USA). Values obtained at these density readings were used for determining the chlorophyll (*a* and *b*) and carotenoids concentrations, according to the method described by Lichtenthaler (1987).

Photosynthetically active radiation (PAR) was measured with a quantummeter (LI-1400 DataLogger, Li-Cor Biosciences, USA). Six readings per experimental unit were taken every hour after forage harvesting from 07:00 to 18:00h. Leaf area index (LAI) was estimated indirectly using a LAI-2000 plant canopy analyzer (Li-Cor Biosciences, USA). Measurements in the SPS were taken at 08:00h, at the forage sampling points from a reference point outside the canopy of trees and from five readings below the SPS canopies in their two layers (close to the ground level and above the forage). In contrast, MONO measurements were taken at 08:00h, at the forage sampling points, totaling one reading above and five readings under the forage canopy, taken at the ground level. From these readings, the device estimated the leaf area index of pastures through procedures described by Welles and Norman (1991).

In each plot, representative tillers of the population were collected to measure the different proportions of lignified (LIG), chlorophyllous (CHLO) and non-lignified (N-LIG) and achlorophyllous (ACHLO) plant tissues (Figure 1). These proportions were measured in cross sections of leaves (middle-third of the first leaf of the second node) and longitudinal and cross sections of stems (middle third of the internode between the second and third nodes). Slides were prepared using a Leica RM-2145 rotary microtome (Leica Microsystems, Germany). Subsequently, they were dissolved in water and stained with 0.0125% aqueous solution of basic fuchsin, 0.5% basic safranin and 1.0% Astra blue solution (Bukatsch, 1972), which allowed distinguishing tissues with different characteristics. Once the double staining was performed, slides were washed in running water, oven-dried at 40°C, subjected to conventional slide mounting and covered with coverslips.

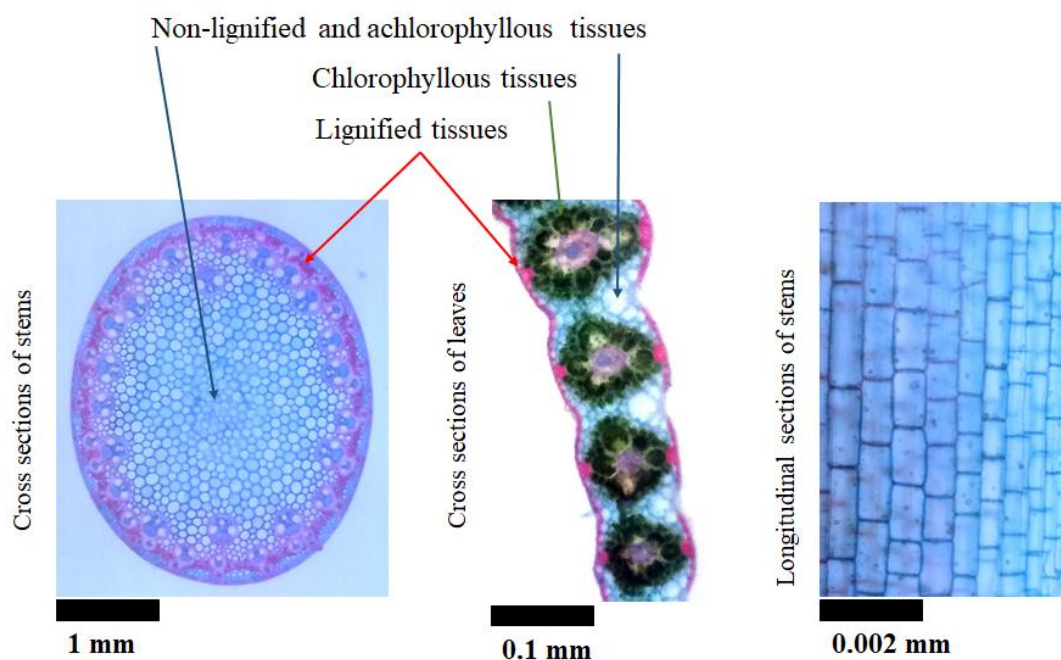


Figure 1. Illustration of the photos used to measure anatomical characteristics.

Then, photomicrographs of the sections were taken using an analog microscope. Photomicrographs were then developed, scanned and digitized, and tissue areas with different characteristics in the leaf and stem cross-sections, and cell length in the longitudinal section of the stem were measured by ImageJ® software.

Statistical analysis

A completely randomized design with six replicates and repeated measures was adopted to analyze the chemical composition, and productive and physiological characteristics of forages. The repeated measures consisted of the seven harvesting periods (P1 to P7): four during the wet season (P1 to P4), one during the transition period (wet-dry) (P5), one during the dry period (P6), and one during the transition period (dry-wet) (P7).

Statistical analysis was performed using the SAS software (SAS Institute Inc., 2004). Data relative to chemical composition, the productive and physiological characteristics of forages were analyzed according to

a repeated-measures model with PROC MIXED of SAS. The sphericity test was applied to check if this assumption was met.

If the sphericity test was not significant ($p > 0.05$) (Situation 1), the univariate structure was chosen (same as PROC GLM); on the other hand, if the sphericity test was significant ($p < 0.05$) (Situation 2), an analysis of variance was applied using mixed models analysis of variance (PROC MIXED procedure), and the variance-covariance matrix was estimated using a restricted maximum likelihood function -2RLL (-2 Res Log Likelihood) and AIC (Akaike's Information Criterion) with -2RLL, AIC values closer to zero indicating a better fit (Wolfinger, 1993). The significance level adopted for analysis of variance was set at 0.05 (probability of Type I error).

Comparisons of means were performed using the Scott-Knott test, as suggested by Conrado, Ferreira, Scapim, and Maluf (2017), with a significance level of 0.05 (probability of Type I error). Regression studies between rainfall and production data were performed to contribute to the discussion of results. The significance level adopted for the analysis of variance was set at 0.05 (probability of Type I error).

Statistical analyses were run in SAS statistical software (SAS Institute Inc., 2004) according to the model as follows:

Statistical model of situation 1: $Y_{ijk} = \mu + S_i + e_{(a)i} + P_j + PS_{ji} + e_{(b)ijk}$, where: " Y_{ijk} " is the observation in the i -th system, j -th harvesting period and k -th replicate; " μ " is the overall mean; " S_i " is the effect of the i -th system, $i = 1, 2$; " $e_{(a)i}$ " is the type A error; " P_j " is the effect of the j -th harvesting period, $j = 1, 2, 3, \dots, 7$; " PS_{ij} " is the effect of interaction between harvesting period \times system; " $e_{(b)ijk}$ " is the type B error.

Statistical model of situation 2: $Y_{ijk} = \mu + S_i + \delta_{ik} + e_{(a)ik} + P_j + PS_{ji} + e_{(b)ijk}$, where, " Y_{ijk} " is the observation in the i -th system, j -th harvesting period and k -th replicate; " μ " is the overall mean; " S_i " is the effect of the i -th system, $i = 1, 2$; " δ_{ik} " is the random effect of the k -th experimental unit on the i -th system; $e_{(a)i}$ is the type A error associated with the i -th system and k -th replicate.; " P_j " is the fixed effect of the j -th harvesting period, $j = 1, 2, 3, \dots, 7$; " PS_{ij} " is the effect of interaction between harvesting period \times system; " $e_{(b)ijk}$ " is the random error associated with the i -th system, j -th harvesting period and k -th replicate.

Results and discussion

Photosynthetically active radiation data

There was an effect of period on shading percentage ranging from 47.2 to 60.2 (Table 2). The greatest proportion of shading occurred between P4 and P6, which seems contradictory, because *Z. tuberculosa* was deciduous at this period (April to July), reducing canopy density. However, PAR in full sun was lower at this period due to the translation movement of the earth, which increases the Earth's tilt with respect to the sun. Consequently, sunlight passes through a thicker layer of atmosphere.

According to Guenni, Seiter, and Figueroa (2008), palisadegrass subjected to 71% artificial shading showed lower biomass compared to 43 and 0%. Shading values in the SPS were close to 55% in most harvesting periods. Although PAR strongly influences forage yield, other factors also contribute to forage production. Sousa et al. (2010) reported that forage mass of palisadegrass under 74% natural shading was reduced by only 15%, demonstrating that other aspects may interfere with plant response to PAR. The greater humidity provided by the SPS may result in more beneficial conditions for forage production compared to MONO (Baliscei et al., 2013). However, several studies have shown that severe PAR restriction decreased forage mass in tropical pastures (Abraham et al., 2014; Lelis et al., 2018; Santiago-Hernández et al., 2016).

Table 2. Photosynthetically active radiation (PAR) and shading percentage in silvopastoral (SPS) and monoculture (MONO) systems during different experimental periods.

System	PAR ($\mu\text{mol}\cdot\text{photons}\cdot\text{sec}^{-1}\cdot\text{m}^{-2}$)							Mean	p-value ¹	SEM
	Period									
	P1	P2	P3	P4	P5	P6	P7			
SPS	715.4aD	600.8aC	571.3aC	473.2aB	369.9aA	367.6aA	652.1aD	535.8	System < 0.001	
MONO	1354.9bC	1304.8bC	1290.6bC	1119.2bB	928.3bA	845.6bA	1258.8bC	1157.5	Period = 0.002	42.696
Mean	1035.2	846.6	931.0	796.2	649.1	606.6	955.5	846.6	Interaction < 0.001	
Shading ² (%)	47.20	53.95	55.73	57.72	60.15	56.53	48.20	54.21		

Means followed by different lowercase letters in the same column are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); Means followed by different uppercase letters in the same row are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); P1 to P4 = wet periods; P5 = transition period (wet-dry); P6 = dry period and P7 = transition period (dry-wet); SEM = Standard error of the mean; ¹Probability of type I error by Fisher' test; ²(PAR in MONO system - PAR in SPS system) \times 100/PAR in MONO system.

Morpho-physiological characteristics

The Lb:SS ratio (Table 3) did not vary ($p > 0.05$) between treatments during the wet season. However, the SPS had greater Lb:SS ratio ($p < 0.05$) than MONO in the transition periods and dry season. Greater Lb:SS ratio in the dry season can be partially explained by less severe water deficit conditions in the SPS compared to the MONO, due to a favorable micro-climate provided by trees in these systems (Baliscei et al., 2013). Furthermore, leaf blade growth rate may be greater in the MONO due to greater sunlight exposure (Taiz, Zeiger, Møller, & Murphy, 2015).

Period had a significant effect ($p < 0.05$) on Lb:SS ratio. In both treatments, the Lb:SS ratio was lower ($p < 0.05$) in the wet season than in the other periods (Table 3). Effect of period on Lb:SS ratio likely occurred due to reduced leaf growth under water deficit conditions, since the leaf is the main transpiration organ and, consequently, responsible for water loss in plants (Taiz et al., 2015).

Table 3. Leaf blade: stem+sheath and green: dead material ratios, canopy height, leaf area index of forage in silvopastoral (SPS) and monoculture (MONO) systems during different experimental periods.

System	Leaf blade: stem+sheath ratio							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	5.34aB	5.22aB	5.02aB	4.62aB	2.12bA	2.79bA	2.33bA	3.92	System = 0.032 Period < 0.001 Interaction = 0.023	0.451
MONO	5.66aB	5.03aB	4.89aB	4.54aB	1.65aA	1.36aA	1.65aA	3.57		
Mean	5.40	5.13	4.96	4.58	1.89	2.08	1.99	3.74		
	Green: dead ratio									
SPS	15.5bC	12.5bC	10.1bC	9.89bC	5.38aB	1.27aA	5.03aB	8.54	System = 0.021 Period = 0.012 Interaction < 0.001	0.667
MONO	10.9aC	8.68aC	7.12aC	6.55aC	6.35aB	1.27aA	4.95aB	6.55		
Mean	13.23	10.61	8.63	8.22	5.87	1.27	4.99	7.54		
	Canopy height (cm)									
SPS	62.0bB	63.3bB	65.0bB	66.7bB	65.0bB	55.3bA	53.7bA	61.6	System = 0.352 Period = 0.024 Interaction = 0.542	2.453
MONO	50.0aB	50.0aB	49.7aB	48.3aB	47.6aB	42.7aA	43.0aA	47.3		
Mean	56.0	56.7	57.3	57.5	56.3	49.0	48.3	54.5		
	Leaf area index									
SPS	3.33bB	3.47bB	3.97bB	3.02bB	3.36bB	1.35aA	1.53aA	2.86	System < 0.001 Period = 0.003 Interaction < 0.001	0.272
MONO	2.01aB	2.51aB	2.15aB	2.09aB	2.18aB	1.39aA	1.51aA	1.98		
Mean	2.67	2.99	3.06	2.56	2.77	1.37	1.52	2.42		

Means followed by different lowercase letters in the same column are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); Means followed by different uppercase letters in the same row are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); P1 to P4 = wet periods; P5 = transition period (wet-dry); P6 = dry period and P7 = transition period (dry-wet); SEM = Standard error of the mean; ¹Probability of type I error by Fisher's test.

During the wet season, the green: dead ratio in the SPS was higher ($p < 0.05$) than in MONO (Table 3). In the transition periods and dry season, the green: dead ratio of the SPS did not differ ($p > 0.05$) from the MONO. Harvesting period had a significant effect ($p < 0.05$) on the green: dead ratio. In both treatments, green: dead ratios were significantly greater ($p < 0.05$) in the wet season than in transition periods (P5 and P7), with greatest values in the dry season (P6). Greater values of green: dead ratio in the SPS during the growing season can be explained by the effect of shading on morphogenic characteristics, such as the number of days of full leaf activity (Santos et al., 2018), and also, shading induces a reduction in dead biomass (Gómez, Guenni, & Bravo de Guenni, 2013). This indicates higher tissue senescence rate in MONO. Furthermore, shading can inhibit bud activity and affect the formation of new leaves and tillers, leading to a reduced leaf metabolism, which increases the maintenance of semi-senescent tissues (Frank & Hofmann, 1994). Formation and maintenance of the living parts of plants depend on several genetic and environmental factors. Among environmental variables, PAR and water may be influential factors (Taiz et al., 2015). These factors were limiting during the dry season (Tables 1 and 2), which impairs the formation and maintenance of living plant tissues, reducing the green: dead ratio.

Canopy height was greater in the SPS ($p < 0.05$) (Table 3). Period had a significant effect ($p < 0.05$) on canopy height, with higher values ($p < 0.05$) in the wet season and transition period (wet-dry) (P5) than in the dry season and transition period (dry-wet) (P7). It is expected that palisadegrass growing under shading conditions would have greater canopy height than under full sunlight. Stem elongation is a compensatory mechanism for the reduced light intensity (Paciullo et al., 2017). This is also explained by the higher availability of water during the experiment, in periods of higher canopy height.

During the wet season and transition period (wet-dry), LAI in the SPS was greater ($p < 0.05$). However, in the dry season and transition period (dry-wet), the LAI of the SPS did not differ ($p > 0.05$) from the MONO (Table 3). Period had a significant effect ($p < 0.05$) on LAI and was greater ($p < 0.05$) in the wet season and transition period (wet-dry) than in the dry season and the transition period (dry-wet).

Leaf area index can interfere with critical ecological aspects, such as inter- and intra-specific competition between plants, carbon retention, and soil conservation. It can also be a crucial component of agroecosystem biogeochemical cycles (Bréda, 2003). Differences in LAI values found in this study are possibly related to changes in leaf appearance and elongation rates, which are influenced by the quantity and quality of light, among other environmental factors. These rates determine some structural canopy characteristics such as the number of leaves per tiller, leaf size, and tiller density, which are responsible for determining the LAI of the canopy (Bahmani, Thom, Matthew, Hooper, & Lemaire, 2003). Higher values of LAI in the SPS may be justified by thinner and longer leaves resulting from the limited light incidence (Santos et al., 2018).

When the LAI value is optimal, the interception of approximately all incident radiation with minimal self-shading will provide the maximum crop growth rate, defined as dry matter weight accumulated per unit area per unit time (Ludlow, Wilson, & Heslehurst, 1974). According to Portes, Ferreira, Peixoto, and Melo (2017), the optimal LAI for palisadegrass monoculture cultivated in the wet season is 6.51, but this LAI is only reached at 97 days of growth, when palisadegrass contains reduced nutritive value (Quintino et al., 2013). The 32-day LAI (range of 29 - 35, on average), proposed by Portes et al. (2017), corresponds to an LAI of 2.01, which is similar to that found in MONO during the wet season, but below the values reported in the SPS (Table 3).

Concentration of chlorophyll *a* in the SPS was greater ($p < 0.05$) (Table 4). In the SPS, the chlorophyll *a* was higher ($p < 0.05$) in the wet season and the transition period (dry-wet) than in the other periods (Table 4). There were no significant effects of period ($p > 0.05$) on chlorophyll *a* concentration between forages harvested, regardless of the period. Concentration of chlorophyll *b* in the SPS was greater than in MONO throughout the experimental period (Table 4). Period had a significant effect ($p < 0.05$) on chlorophyll *b*, which showed greater ($p < 0.05$) values in the wet season and transition period (dry-wet) than in the dry season and in the transition period (wet-dry).

Table 4. Concentrations of chlorophyll *a* and *b*, chlorophyll *a/b* ratio and carotenoid concentration in *U. brizantha* leaves in silvopastoral (SPS) and monoculture (MONO) systems during different experimental periods.

System	Chlorophyll <i>a</i> ($\mu\text{g}\cdot\text{cm}^{-2}$)							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	2.48bB	2.49bB	2.55bB	2.50bB	1.92bA	1.93bA	2.48bB	2.34	System < 0.001 Period = 0.025 Interaction = 0.028	0.139
MONO	1.29aA	1.28aA	1.27aA	1.21aA	1.29aA	1.27aA	1.18aA	1.26		
Mean	1.89	1.89	1.91	1.86	1.61	1.60	1.83	1.80		
System	Chlorophyll <i>b</i> ($\mu\text{g}\cdot\text{cm}^{-2}$)							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	0.87bB	0.86bB	0.90bB	0.88bB	0.73bA	0.74bA	0.93bB	0.84	System < 0.001 Period = 0.041 Interaction < 0.001	0.036
MONO	0.40aB	0.40aB	0.40aB	0.38aB	0.50aA	0.49aA	0.44aB	0.45		
Mean	0.64	0.63	0.65	0.63	0.62	0.62	0.69	0.64		
System	Chlorophyll <i>a/b</i> ratio							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	2.85aA	2.90aA	2.83aA	2.84aA	2.63aB	2.61aB	2.67aB	2.76	System = 0.042 Period = 0.003 Interaction = 0.034	0.083
MONO	3.18bA	3.20bA	3.18bA	3.18bA	2.58aB	2.59aB	2.68aB	2.94		
Mean	3.02	3.05	3.01	3.01	2.61	2.60	2.67	2.85		
System	Carotenoids ($\mu\text{g}\cdot\text{cm}^{-2}$)							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	0.59bB	0.61bB	0.63bB	0.35bA	0.36bA	0.39bA	0.64bB	0.48	System < 0.001 Period = 0.012 Interaction = 0.378	0.019
MONO	0.44aB	0.41aB	0.38aB	0.26aA	0.26aA	0.30aA	0.50aB	0.37		
Mean	0.52	0.51	0.51	0.26	0.29	0.35	0.57	0.43		

Means followed by different lowercase letters in the same column are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); Means followed by different uppercase letters in the same row are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); P1 to P4 = wet periods; P5 = transition period (wet-dry); P6 = dry period and P7 = transition period (dry-wet); SEM = Standard error of the mean; ¹Probability of type I error by Fisher's test.

Response to the amount of light is the result of a sequence of environmental signals and their respective receptors. In the specific case of photosynthesis, light signal is received by pigments (chlorophyll and carotenoids), and the response is directly proportional to the amount of energy received (PAR) up to the light saturation point of a given leaf (Taiz et al., 2015). Cruz et al. (2021) reported a linear increase in the concentration of chlorophyll *a* and *b* in leaves as shading percentage increased. Plants growing under conditions of low radiation develop more

grana (Shao et al., 2014), which are a set of stacked membranous discs (thylakoid) containing chlorophyll and located on chloroplasts (Taiz et al., 2015).

Another theory is that in tropical plants exposed to high light conditions, chlorophyll is continuously synthesized and destroyed (photo-oxidation); however, under higher light intensities, a greater degradation occurs, and equilibrium is reached at a lower concentration. Therefore, shaded leaves have greater chlorophyll concentrations than those exposed to full sunlight (Malaviya et al., 2020). Souza et al. (2016) observed an increase in the concentration of chlorophyll *a* in palisadegrass in response to 50% natural shading. Corroborating with this, Mishra, Tiwari, and Bhatt (2010) reported an increase in total chlorophyll concentration due to shading in tropical forages.

According to Shao et al. (2014), the concentrations of chlorophylls *a* and *b* increase disproportionately in response to natural shading, with a relatively greater increase in chlorophyll *b*. Thus, the greater relative proportion of chlorophyll *b* may be advantageous under shading since it allows for greater light absorption efficiency due to the energy uptake of chlorophyll *b* at other wavelengths (typical of naturally shaded environments). It effectively participates in photochemical reactions, ensuring greater values of photosynthetic rate and forage accumulation. These results are in accordance with Gomes et al. (2019), who stated that one of the physiological characteristics of shaded leaves is the lower amount of secondary pigments and the lower chlorophyll *a/b* ratio compared to leaves in full sun.

Chlorophyll *a/b* ratio in SPS was lower ($p < 0.05$) than in MONO during the wet season (Table 4). The chlorophyll *a/b* ratio generally tends to decrease with reduced light intensity due to greater relative proportion of chlorophyll *b* in shaded environments. This because chlorophyll *b* molecule is degraded more slowly in shaded plants than chlorophyll *a* (Taiz et al., 2015). In the transition periods, and the dry season, the chlorophyll *a/b* ratio did not differ ($p > 0.05$) between systems (Table 4). Nonetheless, chlorophyll *a/b* ratio was lower ($p < 0.05$) in the wet season compared to the transition period and dry season (Table 4), possibly due to factors such as lower PAR incidence and a higher proportion of shading (Table 2) in the transition period and dry season. Such factors may increase chlorophyll *b* concentration by reducing chlorophyll *a/b* ratio.

Concentration of carotenoids was greater ($p < 0.05$) in the SPS (Table 4). In both treatments, carotenoids were greater ($p < 0.05$) in the first three periods of the wet season and in the transition period (dry-wet) than in the last period of the wet season, the transition period (wet-dry) and the dry season (Table 4). Carotenoids are essential components in the photosynthetic antenna complex, contributing to the absorption of incident radiation and dissipation of excess absorbed energy, among other functions (Taiz et al., 2015) and, in general, forage grasses and cover crops used as forage are rich sources of carotenoids (Maxin, Cornu, Andueza, Laverroux, & Graulet, 2020; Nozière et al., 2006). Under water or light stress conditions, carotenoids concentration in the leaves tends to increase (Shao et al., 2014), which corroborates the results found in this experiment.

Anatomical characteristics

There was no effect of treatment ($p > 0.05$) on the proportion of N-LIG and ACHLO tissues (Table 5). However, the proportion of N-LIG and ACHLO tissues was lower in the dry season than in the wet season, which may be explained by the increased proportion of LIG areas (Table 5). Increase in lignin concentration of perennial grasses in the dry season is an adaptive strategy for greater conservation of tissue moisture and chemical energy photoassimilated in the wet season (Gomes et al., 2019). According to some authors (Santos et al., 2016; Sousa et al., 2010), morphostructural alterations of some tropical forages is one of the responses associated with shading; however, literature evaluating anatomical modifications in the proportion of tissues is still scarce.

Leaves have three different areas: N-LIG and ACHLO areas, LIG areas, and CHLO areas. Forage in MONO had a similar proportion of N-LIG and ACHLO areas compared to forage in the SPS, with differences ($p > 0.05$) only in the proportion of areas during the P1 in the SPS (Table 6). This indicates that treatments did not influence the increase or reduction of these areas. Regarding LIG areas, no significant differences were detected between harvesting periods and forage production systems ($p > 0.05$; Table 6).

Proportion of CHLO area in the SPS did not vary during the experimental period, except for the P1 period (greatest rainfall). The proportion of CHLO in MONO pastures was lower in the transition period and dry season than in the wet season (Table 6). Valente et al. (2016) reported anatomical differences in leaves of tropical forage exposed to full sunlight or shaded, but these modifications were conditional to shading percentage. This study was conducted with a 54% shading, proportion lower than the one tested

by the aforementioned authors (69%). This difference possibly explains the non-anatomical differences in this study. The lack of modifications in the proportion of areas in leaves (N-LIG and ACHLO; LIG, and CHLO) strengthens the idea that there is no change in the chemical composition of the leaf blade tissue. Thus, changes in the nutritive value of forage are more related to the proportion of leaves in the canopy (Lee, 2018).

Table 5. Proportion of non-lignified and lignified areas in stem cross-sections of forage in silvopastoral (SPS) and monoculture (MONO) systems during different experimental periods.

Systems	Non-lignified and achlorophyllous area (%)							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	70.1aA	71.6aA	66.9aB	66.4aB	68.4aB	61.8aC	62.8aC	66.9	System = 0.563 Period = 0.041 Interaction = 0.039	1.52
MONO	68.9aA	68.3bA	66.6aA	66.9aA	66.5aA	62.8aB	64.8aB	66.4		
Mean	69.5	70.0	66.8	66.7	67.5	62.3	63.8	66.6		
Lignified area (%)										
SPS	29.9aA	27.4bA	33.1aA	33.6aA	31.6aA	38.2aB	37.2aB	33.2	System = 0.452 Period = 0.029 Interaction = 0.047	1.617
MONO	31.2aA	32.7aA	33.4aB	33.1aB	33.5aB	37.3aC	35.2aC	33.6		
Mean	30.5	30.0	33.3	33.4	32.5	37.8	36.2	33.4		

Means followed by different lowercase letters in the same column are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); Means followed by different uppercase letters in the same row are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); P1 to P4 = wet periods; P5 = transition period (wet-dry); P6 = dry period and P7 = transition period (dry-wet); SEM = Standard error of the mean; ¹Probability of type I error by Fisher's test.

Table 6. Proportion of non-lignified and achlorophyllous areas, lignified areas and chlorophyllous areas in leaf cross-sections of forage in silvopastoral (SPS) and monoculture (MONO) systems during different experimental periods.

System	Non-lignified and achlorophyllous area (% total leaf cross-sectional area)							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	36.7aA	46.5aB	43.7aB	46.7aB	46.2aB	50.7aB	46.4aB	45.4	System = 0.659 Period = 0.041 Interaction = 0.025	1.754
MONO	40.0aA	45.7aA	51.4aA	43.5aA	43.2aA	52.4aA	43.5aA	45.7		
Mean	38.3	46.1	47.5	45.1	44.7	52.1	44.9	45.5		
Lignified area (% total leaf cross-sectional area)										
SPS	26.1	23.2	20.5	23.2	21.9	22.9	23.0	23.1a	System = 0.543 Period = 0.729 Interaction = 0.417	1.137
MONO	23.4	25.7	25.4	24.2	25.7	24.7	24.2	24.8a		
Mean	24.7	24.4	23.0	23.7	23.8	24.3	23.6	23.9		
Chlorophyllous area (% total leaf cross-sectional area)										
SPS	37.3aA	30.8aB	31.7aB	30.8aB	29.3aB	28.4aB	30.6aB	30.7	System = 0.562 Period = 0.046 Interaction = 0.032	1.440
MONO	38.6aA	32.9aB	31.6aB	30.4aC	30.8aC	22.9aD	30.4aC	31.2		
Mean	38.0	31.8	31.7	30.6	30.0	23.7	30.5	31.0		

Means followed by different lowercase letters in the same column are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); Means followed by different uppercase letters in the same row are significantly different by Scott-Knott test at a significance level of 0.05 (probability of Type I error); P1 to P4 = wet periods; P5 = transition period (wet-dry); P6 = dry period and P7 = transition period (dry-wet); SEM = Standard error of the mean; ¹Probability of type I error by Fisher's test.

Reduced proportions of CHLO area during the wet-dry transition periods and dry season, can be because these periods have climate restrictions related to photosynthetic activity and plant development, in MONO and SPS. Plants under shading have increased concentration of total chlorophylls (Taiz et al., 2015) as observed in this study. Moreover, the number of cells and the amount of CHLO tissues also increased with natural shading. The literature corroborates these results, emphasizing that this aspect is typical of tropical Poaceae plants, the so-called anatomophysiological plasticity (Costa et al., 2015; Paciullo et al., 2011; Souza et al., 2016).

The stem cell length did not differ ($p > 0.05$) between treatments, and was not affected by the System \times Period interaction ($p > 0.05$) (Table 7). Stem etiolation possibly occurred due to greater meristem cell proliferation at stem nodes of forages in the SPS. Anatomical studies on longitudinal sections of the stem in shaded plants are essential to elucidate the process of etiolation in tropical grasses (Lelis et al., 2018; Sousa et al., 2010). This process may be linked to the balance of plant hormones with stem cell elongation, including auxin (Bartel, 1997; Mutai, Njuguna, & Ghimire, 2017) and hormones, such as gibberellins and cytokines (Zaman, Kurepin, Catto, & Pharis, 2016), which trigger cell division and differentiation in plants (Taiz et al., 2015).

Table 7. Mean cell length in the cross-sectional area of the middle third of the forage stem in silvopastoral (SPS) and monoculture (MONO) systems during different experimental periods.

Systems	Mean cell length in the cross-sectional area of the stem (μm)							Mean	p-value ¹	SEM
	P1	P2	P3	P4	P5	P6	P7			
SPS	1.85	1.87	1.85	1.87	1.88	1.87	1.86	1.87	System = 0.342 Period = 0.582 Interaction = 0.198	0.115
MONO	1.71	1.86	1.77	1.72	1.92	1.71	1.72	1.73		
Mean	1.78	1.87	1.81	1.80	1.90	1.74	1.79	1.80		

P1 to P4 = wet periods; P5 = transition period (wet-dry); P6 = dry season and P7 = transition period (dry-wet); SEM = Standard error of the mean.

¹Probability of type I error by Fisher's test.

Conclusion

The SPS alters morpho-physiological characteristics of palisadegrass and increases the concentration of chlorophyll *a* and *b*. The proportion of chlorophyllous cells in forage leaves in the SPS remains constant throughout the year, even in the transition and dry periods, which does not occur in the MONO. Etiolation in the SPS is due to cell multiplication and not to the increase in stem cell length.

Acknowledgements

This work was funded by the *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES)*, *Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq)*, *Fundação de Amparo a Pesquisa do Estado de Minas Gerais (PAPEMIG)*, Graduate Program of the Federal University of Minas Gerais - Veterinary School and the Grants Program of the Federal University of Tocantins.

References

- Abraham, E. M., Kyriazopoulos, A. P., Parissi, Z. M., Kostopoulou, P., Karatassiou, M., Anjalanidou, K., & Katsouta, C. (2014). Growth, dry matter production, phenotypic plasticity, and nutritive value of three natural populations of *Dactylis glomerata* L. under various shading treatments. *Agroforestry Systems*, *88*(2), 287-299. DOI: <https://doi.org/10.1007/s10457-014-9682-9>
- Almeida, E. X., Maraschin, G. E., Harthmann, O. E. L., Ribeiro Filho, H. M. N., & Setelich, E. A. (2000). Oferta de forragem de Capim-Elefante Anão "Mott" e a dinâmica da pastagem. *Revista Brasileira de Zootecnia*, *29*(5), 1281-1287. DOI: <https://doi.org/10.1590/s1516-35982000000500004>
- Association of Official Analytical Chemists [AOAC]. (1990). Animal feed. In K. Helrich (Ed.), *Association of Official Analytical Chemists [AOAC]* (Vol. 1, 15th ed., p. 69-90). Arlington, VA, USA: AOAC.
- Bahmani, I., Thom, E. R., Matthew, C., Hooper, R. J., & Lemaire, G. (2003). Tiller dynamics of perennial ryegrass cultivars derived from different New Zealand ecotypes: Effects of cultivar, season, nitrogen fertiliser, and irrigation. *Australian Journal of Agricultural Research*, *54*(8), 803-817. DOI: <https://doi.org/10.1071/AR02135>
- Balisci, M. A., Barbosa, O. R., Souza, W., Costa, M. A. T., Krutzmann, A., & Queiroz, E. O. (2013). Microclimate without shade and silvopastoral system during summer and winter. *Acta Scientiarum - Animal Sciences*, *35*, 49-56. DOI: <https://doi.org/10.4025/actascianimsci.v35i1.15155>
- Bartel, B. (1997). Auxin biosynthesis. *Annual Review of Plant Physiology and Plant Molecular Biology*, *48*, 51-66. DOI: <https://doi.org/10.1146/annurev.arplant.48.1.51>
- Bréda, N. J. J. (2003). Ground-based measurements of leaf area index: A review of methods, instruments and current controversies. *Journal of Experimental Botany*, *54*(392), 2403-2417. DOI: <https://doi.org/10.1093/jxb/erg263>
- Bukatsch, F. (1972). Bemerkungen zur Doppelfärbung Astrablau- Safranin. *Mikrokosmos*, *61*, 255.
- Chacón, E., Stobbs, T. H., & Haydock, K. P. (1977). Estimation of leaf and stem contents of oesophageal extrusa samples from cattle. *Journal of the Australian Institute of Agricultural Science*, *43*(1-2), 73-75.
- Conrado, T. V., Ferreira, D. F., Scapim, C. A., & Maluf, W. R. (2017). Adjusting the Scott-Knott cluster analyses for unbalanced designs. *Crop Breeding and Applied Biotechnology*, *17*, 1-9. DOI: <https://doi.org/10.1590/1984-70332017v17n1a1>

- Costa, J. P. R., Caputti, G. P., Galzerano, L., Silva, W. L., Ruggieri, A. C., & Malheiros, E. B. (2015). Relative chlorophyll contents in the evaluation of the nutritional status of nitrogen from xaraes palisade grass and determination of critical nitrogen sufficiency index. *Acta Scientiarum - Animal Sciences*, 37(2), 109-114. DOI: <https://doi.org/10.4025/actascianimsci.v37i2.24854>
- Cruz, P. J. R., Borges, C. E., Santos, M. V., Magalhães, M. A., Martuscello, J. A., Fonseca, D. M., & Silva, L. D. (2021). Shade effects on the hybrid Mavuno brachiariagrass (*Urochloa* spp.) as potential grass in agroforestry systems. *Agroforestry Systems*, 95(6), 1105-1108. DOI: <https://doi.org/10.1007/s10457-021-00636-4>
- Dias-Filho, M. B. (2014). Diagnóstico das Pastagens no Brasil. In *Embrapa Amazônia Oriental, Documentos* (Vol. 402). Belém, Pará, Brasil. Retrieved from <https://www.infoteca.cnptia.embrapa.br/bitstream/doc/986147/1/DOC402.pdf>
- Frank, A. B., & Hofmann, L. (1994). Light quality and stem numbers in cool-season forage grasses. *Crop Science*, 34(2), 468-473. DOI: <https://doi.org/10.2135/cropsci1994.0011183X003400020030x>
- Gomes, F. J., Cavalli, J., Pedreira, B. C., Pedreira, C. G. S., Holschuch, S. G., & Pereira, D. H. (2022). Forage nutritive value of Marandu palisade grass under clipping in a silvopastoral system. *Agroforestry Systems*, 96, 79-88. DOI: <https://doi.org/10.1007/s10457-021-00696-6>
- Gomes, F. J., Pedreira, C. G. S., Bosi, C., Cavalli, J., Holschuch, S. G., Mourão, G. B., ... Pedreira, B. C. (2019). Shading effects on marandu palisadegrass in a silvopastoral system: Plant morphological and physiological responses. *Agronomy Journal*, 111(5), 2332-2340. DOI: <https://doi.org/10.2134/agronj2019.01.0052>
- Gómez, S., Guenni, O., & Bravo de Guenni, L. (2013). Growth, leaf photosynthesis and canopy light use efficiency under differing irradiance and soil N supplies in the forage grass *Brachiaria decumbens* Stapf. *Grass and Forage Science*, 68(3), 395-407. DOI: <https://doi.org/10.1111/gfs.12002>
- Guenni, O., Romero, E., Guédez, Y., Bravo de Guenni, L., & Pittermann, J. (2018). Influence of low light intensity on growth and biomass allocation, leaf photosynthesis and canopy radiation interception and use in two forage species of *Centrosema* (DC.) Benth. *Grass and Forage Science*, 73(4), 967-978. DOI: <https://doi.org/10.1111/gfs.12368>
- Guenni, O., Seiter, S., & Figueroa, R. (2008). Growth responses of three *Brachiaria* species to light intensity and nitrogen supply. *Tropical Grasslands*, 42(2), 75-87.
- Jank, L., Barrios, S. C., Do Valle, C. B., Simeão, R. M., & Alves, G. F. (2014). The value of improved pastures to Brazilian beef production. *Crop and Pasture Science*, 65(11), 1132-1137. DOI: <https://doi.org/10.1071/CP13319>
- Jose, S., & Dollinger, J. (2019). Silvopasture: a sustainable livestock production system. *Agroforestry Systems*, 93(1), 1-9. DOI: <https://doi.org/10.1007/s10457-019-00366-8>
- Jose, S., Walter, D., & Mohan Kumar, B. (2019). Ecological considerations in sustainable silvopasture design and management. *Agroforestry Systems*, 93(1), 317-331. DOI: <https://doi.org/10.1007/s10457-016-0065-2>
- Lee, M. A. (2018). A global comparison of the nutritive values of forage plants grown in contrasting environments. *Journal of Plant Research*, 131(4), 641-654. DOI: <https://doi.org/10.1007/s10265-018-1024-y>
- Lelis, D. L., Rennó, L. N., Chizzotti, M. L., Pereira, C. E. R., Silva, J. C. P., Moreira, L. G. T., ... Chizzotti, F. H. M. (2018). Photosensitization in naïve sheep grazing signal grass (*Brachiaria decumbens*) under full sunlight or a silvopastoral system. *Small Ruminant Research*, 169(January), 24-28. DOI: <https://doi.org/10.1016/j.smallrumres.2018.08.018>
- Lichtenthaler, H. K. (1987). Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Methods in Enzymology*, 148, 350-382. DOI: [https://doi.org/10.1016/0076-6879\(87\)48036-1](https://doi.org/10.1016/0076-6879(87)48036-1)
- Ludlow, M. M., Wilson, G. L., & Heslehurst, M. R. (1974). Studies on the productivity of tropical pasture plants. V. Effect of shading on growth, photosynthesis and respiration in two grasses and two legumes. *Australian Journal of Agricultural Research*, 25(3), 425-433. DOI: <https://doi.org/10.1071/AR9740425>
- Malaviya, D. R., Baig, M. J., Kumar, B., & Kaushal, P. (2020). Effects of shade on Guinea grass genotypes *Megathyrus maximus* (Poales: Poaceae). *Revista de Biologia Tropical*, 68(2), 563-572. DOI: <https://doi.org/10.15517/RBT.V68I2.38362>
- Ministério da Agricultura, Pecuária e do Abastecimento [MAPA]. (2018). *Controle da Produção de Sementes e Mudas*. Retrieved from <http://indicadores.agricultura.gov.br/sigefsementes/index.htm>

- Maxin, G., Cornu, A., Andueza, D., Laverroux, S., & Graulet, B. (2020). Carotenoid, tocopherol, and phenolic compound content and composition in cover crops used as forage. *Journal of Agricultural and Food Chemistry*, *68*(23), 6286-6296. DOI: <https://doi.org/10.1021/acs.jafc.0c01144>
- Mishra, A. K., Tiwari, H. S., & Bhatt, R. K. (2010). Growth, biomass production and photosynthesis of *Cenchrus ciliaris* L. under *Acacia tortilis* (Forssk.) Hayne based silvopastoral systems in semi arid tropics. *Journal of Environmental Biology*, *31*(6), 987-993.
- Murgueitio, E., Chará, J., Barahona, R., & Rivera, J. (2019). Development of sustainable cattle rearing in silvopastoral systems in Latin America. *Cuban Journal of Agricultural Science*, *53*, 65-71.
- Mutai, C., Njuguna, J., & Ghimire, S. (2017). Brachiaria Grasses (*Brachiaria* spp.) harbor a diverse bacterial community with multiple attributes beneficial to plant growth and development. *Microbiology Open*, *6*(5), e00497. DOI: <https://doi.org/10.1002/mbo3.497>
- Nozière, P., Graulet, B., Lucas, A., Martin, B., Grolier, P., & Doreau, M. (2006). Carotenoids for ruminants: From forages to dairy products. *Animal Feed Science and Technology*, *131*(3-4), 418-450. DOI: <https://doi.org/10.1016/j.anifeedsci.2006.06.018>
- Oliveira, J. de C., Azevedo, A. M., Ribeiro, J. M., Freitas, I. C., Dias, R. F., Duarte, A. C. S., ... Frazão, L. A. (2021). Sampling representativeness of soil carbon and physiological parameters of marandu palisadegrass in a tropical silvopastoral system. *Scientia Agricola*, *78*(suppl e20200176), 1-9. DOI: <https://doi.org/10.1590/1678-992x-2020-0176>
- Paciullo, D. S. C., Fernandes, P. B., Carvalho, C. A. B., Morenz, M. J. F., Lima, M. A., Maurício, R. M., & Gomide, C. A. M. (2021). Pasture and animal production in silvopastoral and open pasture systems managed with crossbred dairy heifers. *Livestock Science*, *245*(3), 104426. DOI: <https://doi.org/10.1016/j.livsci.2021.104426>
- Paciullo, D. S. C., Fernandes, P. B., Gomide, C. A. M., Castro, C. R. T., Souza Sobrinho, F., & Carvalho, C. A. B. (2011). The growth dynamics in Brachiaria species according to nitrogen dose and shade. *Revista Brasileira de Zootecnia*, *40*(2), 270-276. DOI: <https://doi.org/10.1590/s1516-35982011000200006>
- Paciullo, D. S. C., Gomide, C. A. M., Castro, C. R. T., Maurício, R. M., Fernandes, P. B., & Morenz, M. J. F. (2017). Morphogenesis, biomass and nutritive value of *Panicum maximum* under different shade levels and fertilizer nitrogen rates. *Grass and Forage Science*, *72*(3), 590-600. DOI: <https://doi.org/10.1111/gfs.12264>
- Paladines, O. (1992). Medida de la producción primaria de los potreros bajo condiciones de pastoreo. In *Metodologías de Pastizales* (Serie Meto, p. 39-81). Quito, EC: Convenio Ecuatoriano-Alemán.
- Pang, K., Van Sambeek, J. W., Navarrete-Tindall, N. E., Lin, C.-H., Jose, S., & Garrett, H. E. (2019a). Responses of legumes and grasses to non-, moderate, and dense shade in Missouri, USA. II. Forage quality and its species-level plasticity. *Agroforestry Systems*, *93*, 25-38. DOI: <https://doi.org/10.1007/s10457-017-0068-7>
- Pang, K., Van Sambeek, J. W., Navarrete-Tindall, N. E., Lin, C.-H., Jose, S., & Garrett, H. E. (2019b). Responses of legumes and grasses to non-, moderate, and dense shade in Missouri, USA. I. Forage yield and its species-level plasticity. *Agroforestry Systems*, *93*, 11-24. DOI: <https://doi.org/10.1007/s10457-017-0067-8>
- Portes, T. A., Ferreira, A. M., Peixoto, M. M., & Melo, H. C. (2017). Growth and senescence of *Urochloa brizantha* under Brazilian Cerrado conditions. *African Journal of Agricultural Research*, *12*(34), 2625-2632. DOI: <https://doi.org/10.5897/ajar2017.12366>
- Quintino, A. C., Abreu, J. G., Almeida, R. G., Macedo, M. C. M., Cabral, L. S., & Galati, R. L. (2013). Production and nutritive value of piatã grass and hybrid sorghum at different cutting ages. *Acta Scientiarum - Animal Sciences*, *35*(3), 243-249. DOI: <https://doi.org/10.4025/actascianimsci.v35i3.18016>
- Santiago-Hernández, F., López-Ortiz, S., Ávila-Reséndiz, C., Jarillo-Rodríguez, J., Pérez-Hernández, P., & Guerrero-Rodríguez, J. (2016). Physiological and production responses of four grasses from the genera *Urochloa* and *Megathyrsus* to shade from *Melia azedarach* L. *Agroforestry Systems*, *90*, 339-349. DOI: <https://doi.org/10.1007/s10457-015-9858-y>
- Santos, D. C., Guimarães Júnior, R., Vilela, L., Pulrolnik, K., Bufon, V. B., & França, A. F. S. (2016). Forage dry mass accumulation and structural characteristics of Piatã grass in silvopastoral systems in the Brazilian savannah. *Agriculture, Ecosystems and Environment*, *233*, 16-24. DOI: <https://doi.org/10.1016/j.agee.2016.08.026>

- Santos, M. V., Ferreira, E. A., Cruz, P. J. R., Ribeiro, V. H. V., Alencar, B. T. B., Cabral, C. M., ... Aspiazú, I. (2018). Leaf anatomy of “Marandu” grass cultivated in plant arrangements in agrosilvopastoral systems. *Pesquisa Agropecuaria Brasileira*, 53(12), 1320-1328. DOI: <https://doi.org/10.1590/S0100-204X2018001200004>
- SAS Institute Inc. (2004). *SAS/Stat® 9.1. User’s Guide*. Cary, NC: SAS.
- Shao, Q., Wang, H., Guo, H., Zhou, A., Huang, Y., Sun, Y., & Li, M. (2014). Effects of shade treatments on photosynthetic characteristics, chloroplast ultrastructure, and physiology of *de Anoctochilus roxburghii*. *Plos One*, 9(2), e85996. DOI: <https://doi.org/10.1371/journal.pone.0085996>
- Silva, I. A. G., Dubeux Jr, J. C. B., Melo, A. C. L., Cunha, M. V., Santos, M. V. F., Apolinário, V. X. O., & Freitas, E. V. (2021a). Tree legume enhances livestock performance in a silvopasture system. *Agronomy Journal*, 113, 358-369. DOI: <https://doi.org/10.1002/agj2.20491>
- Silva, I. A. G., Dubeux Jr, J. C. B., Santos, M. V. F., Mello, A. C. L., Cunha, M. V., Apolinário, V. X. O., & Freitas, E. V. (2021b). Tree canopy management affects dynamics of herbaceous vegetation and soil moisture in silvopasture systems using arboreal legumes. *Agronomy*, 11(8), 1509. DOI: <https://doi.org/10.3390/agronomy11081509>
- Sousa, L. F., Maurício, R. M., Moreira, G. R., Gonçalves, L. C., Borges, I., & Pereira, L. G. R. (2010). Nutritional evaluation of “Braquiaraõ” grass in association with “Aroeira” trees in a silvopastoral system. *Agroforestry Systems*, 79, 189-199. DOI: <https://doi.org/10.1007/s10457-010-9297-8>
- Sousa, L. F., Maurício, R. M., Paciullo, D. S. C., Silveira, S. R., Ribeiro, R. S., Calsavara, L. H., & Moreira, G. R. (2015). Forage intake, feeding behavior and bio-climatological indices of pasture grass, under the influence of trees, in a silvopastoral system. *Tropical Grasslands-Forrajes Tropicales*, 3(3), 129-141. DOI: [https://doi.org/10.17138/TGFT\(3\)129-141](https://doi.org/10.17138/TGFT(3)129-141)
- Souza, I. A., Ribeiro, K. G., Rocha, W. W., Araújo, S. A. C., Pereira, O. G., & Cecon, P. R. (2016). Forage mass, chemical composition and leaf chlorophyll index of signal grass and organic matter in soil under increasing levels of nitrogen. *Semina: Ciências Agrárias*, 37(3), 1505-1514. DOI: <https://10.5433/1679-0359.2016v37n3p1505>
- Taiz, L., Zeiger, E., Møller, I. M., & Murphy, A. (2015). Plant Physiology and Development. In *Journal of Chemical Information and Modeling* (6th ed., p. 761). Sunderland, MA: Sinauer Associates.
- Tóth, G., Montanarella, L., Stolbovoy, V., Máté, F., Bódis, K., Jones, A., ... Van Liedekerke, M. (2008). *Soils of the European Union*. Ispra, Varese, Italy.: European Commission. Joint Research Centre. Institute for Environment and Sustainability. DOI: <https://doi.org/10.2788/87029>
- Valente, T. N. P., Lima, E. S., Gomes, D. I., Santos, W. B. R., Cesário, A. S., & Santos, S. C. (2016). Anatomical differences among forage with respect to nutrient availability for ruminants in the tropics: A review. *African Journal of Agricultural Research*, 11(18), 1585-1592. DOI: <https://doi.org/10.5897/ajar2016.10828>
- Viana, V. M., Maurício, R. M., Matta-Machado, R., & Pimenta, I. A. (2002). Manejo de la regeneración natural de especies arbóreas nativas para la formación de sisemas silvopastoriles en la zonas de bosques secos de sureste de Brasil. *Agroforestería En Las Américas*, 9(33-34), 48-52.
- Welles, J. M., & Norman, J. M. (1991). Instrument for indirect measurement of canopy architecture. *Agronomy Journal*, 83(5), 818-825. DOI: <https://doi.org/10.2134/agronj1991.00021962008300050009x>
- Wolfinger, R. (1993). Covariance structure selection in general mixed models. *Communications in Statistics - Simulation and Computation*, 22(4), 1079-1106. DOI: <https://doi.org/10.1080/03610919308813143>
- Zaman, M., Kurepin, L. V., Catto, W., & Pharis, R. P. (2016). Evaluating the use of plant hormones and biostimulators in forage pastures to enhance shoot dry biomass production by perennial ryegrass (*Lolium perenne* L.). *Journal of the Science of Food and Agriculture*, 96(3), 715-726. DOI: <https://doi.org/10.1002/jsfa.7238>