



Journal of Essential Oil Research

ISSN: 1041-2905 (Print) 2163-8152 (Online) Journal homepage: http://www.tandfonline.com/loi/tjeo20

Scents from Brazilian Cerrado: *Psidium myrsinites* DC. (Myrtaceae) leaves and inflorescences essential oil

Fernando C.M. de Medeiros, Cláudio H.S. Del Menezzi, Humberto R. Bizzo & Roberto F. Vieira

To cite this article: Fernando C.M. de Medeiros, Cláudio H.S. Del Menezzi, Humberto R. Bizzo & Roberto F. Vieira (2015) Scents from Brazilian Cerrado: *Psidium myrsinites* DC. (Myrtaceae) leaves and inflorescences essential oil, Journal of Essential Oil Research, 27:4, 289-292, DOI: <u>10.1080/10412905.2015.1037020</u>

To link to this article: https://doi.org/10.1080/10412905.2015.1037020



Published online: 27 Apr 2015.

-	_
ſ	
L	Ø,
	_

Submit your article to this journal 🖸

Article views: 91



View Crossmark data 🗹

Citing articles: 4 View citing articles 🖸



Scents from Brazilian Cerrado: *Psidium myrsinites* DC. (Myrtaceae) leaves and inflorescences essential oil[†]

Fernando C.M. de Medeiros^a, Cláudio H.S. Del Menezzi^a, Humberto R. Bizzo^b and Roberto F. Vieira^c*

^aDepartamento de Engenharia Florestal, Universidade de Brasília (UnB), Brasília, DF, Brasil; ^bEmbrapa Agroindústria de Alimentos, Rio de Janeiro, RJ, Brasil; ^cEmbrapa Recursos Genéticos e Biotecnologia, Brasília, DF, Brasil

(Received 22 September 2014; accepted 28 March 2015)

The chemical profile of the essential oil obtained from leaves and inflorescences of *Psidium myrsinites* DC. (Myrtaceae) growing wildly in the Brazilian Cerrado was analyzed by GC/FID and CG/MS. The essential oil had a slimy and translucent appearance. The oil yield was 0.1% and the main components were oxygenated sesquiterpenes (68.2%) and non-oxygenated sesquiterpenes (18.3%). Caryophyllene oxide (26.1%), humulene epoxide II (8.8%), β -caryophyllene (7.4%) and α -caryophyllene (5.4%) were the major compounds. It is worth to notice that the most abundant substances have very close structures. The oil yield and composition was very similar to other species belonging to the same genus.

Keywords: Psidium myrsinites; Myrtaceae; essential oil; Brazilian Cerrado; sesquiterpenes; caryophyllene oxide

Introduction

The Brazilian savannah, called Cerrado, is one of the most important biomes subject to actions of conservation because of their high degree of endemism and huge land degradation, being one of the *hot spots* of biodiversity (1).

The existence of many micro-environments provides an intricate mosaic of ecosystems and allows a great number of endemic species in Cerrado (2). It has been estimated that there is more than 12,000 species of native flora from Brazilian Cerrado (3). Forty-four percentage of plant life occurring in this biome is endemic, making the Brazilian Cerrado the most biodiverse tropical savannah in the planet (4).

Among so many species, it is noteworthy that the genus *Psidium* L. is one of the most ordinary in the Cerrado. There are thirty-one different native *Psidium* species in this biome, meanwhile in the whole country there are sixty-one species from which forty-one are endemic in Brazil (5).

Species from the genus *Psidium* could be used in the production of woods and in the traditional medicine, however, their main characteristic is the presence of fruits, better known as araçás (6). Furthermore, the chemical constituents of the fruits show high content of phenolic compounds, usually responsible for the elevated antioxidant activity and other biological properties already observed (7–9). In the Brazilian Cerrado, it has already been studied the nutritional potential of the araçás, being rich in fibers and minerals (10).

The aim of this work was to analyze the essential oil extracted by hydrodistillation from the leaves and inflorescences of a *Psidium* species: *Psidium myrsinites* DC. (Myrtaceae).

Experimental

Plant material

Leaves and inflorescences of five individuals were randomly collected in June 2013, between 9 and 11 AM. The collection was performed in Cerrado microenvironment known as *stricto sensu* inside Fazenda Água Limpa (FAL), a protected area from the University of Brasília (UnB). The specimens were small trees which rose on red yellow latosol soil at 1,096 m above sea level.

Right after collection, the plant material was weighted and dried in a forced-air drier at 38°C until constant weight. A voucher specimen (CEN 84482) was deposited at the Embrapa Genetic Resources and Biotechnology herbarium, and identified by a taxonomist.

Extraction

The essential oil was extracted by hydrodistillation of the dried material (353.93 g) in a modified Clevengertype apparatus. Entire leaves and inflorescences were

^{*}Corresponding author. Email: roberto.vieira@embrapa.br [†]Part 4 in the series Scents from Brazilian Cerrado.

carefully separated from stems and distilled without crushing. The essential oil was collected, dried with anhydrous sodium sulfate and stored in the freezer for later analysis.

Essential oil analysis

The essential oil was analyzed in an Agilent 7890A gas chromatograph fitted with a flame ionization detector (GC-FID) and equipped with a 5%-phenyl-95%-methylpolysilixane (HP-5MS, 30 m × 032 mm × 0.25 μ m) fused silica capillary column. The oven temperature was programmed from 60°C to 240°C at 3°C/min, the injector was kept at 250°C and the detector was kept at 280°C. Hydrogen was used as carrier gas at 1.5 mL/min. The oil was diluted at 1% in dichloromethane (V/V) and 1 μ L of the solution was injected in split mode (1:20). The quantification

of the constituents was done by normalized relative area of the peaks from a mean of three injections considering all factor responses as 1.

The mass spectra analyses were made in an Agilent 5973N gas chromatograph operating in the same conditions than above. The carrier gas was helium (1.0 mL/min) and the mass detector was operated in electronic ionization mode (70 eV) at 3.12 scans/s with a mass range from 40 to 450 u.

The linear retention indices (LRI) were obtained by the injection of a homologous serious of n-alkanes (C₉-C₂₈) in the same conditions and column as above. The indices were calculated by the Van Den Dool and Kratz equation (11).

The identification of the compounds was made by comparison of their mass spectra with the NIST MS database and the linear retention indices found in the literature (12, 13).

Table 1. Chemical constituents of the essential oil from leaves and flowers of P. myrsinites.

Peak #	Constituents	LRI^{a}_{exp}	LRI^b_{lit}	Concentration (%) ^c
1	tricyclene	921	919	0.6
2	α-pinene	932	932	0.8
3	β-pinene	974	975	0.3
4	myrcene	988	990	5.4
5	<i>p</i> -cymene	1020	1023	0.2
6	limonene	1024	1026	1.5
7	linalool	1095	1099	2.5
8	β-pinene oxide	1154	1154	0.5
9	4-terpineol	1174	1174	0.2
10	a-terpineol	1186	1188	0.3
11	α-copaene	1374	1371	0.4
12	β-caryophyllene	1417	1413	7.4
13	aromadendrene	1439	1432	1.0
14	α-caryophyllene	1452	1447	5.4
15	allo-aromadendrene	1458	1454	0.2
16	γ-muurolene	1478	1471	0.7
17	β-selinene	1489	1479	1.3
18	(E)-methyl isoeugenol	1491	1481	0.7
19	α-selinene	1498	1488	1.0
20	γ-cadinene	1513	1508	0.4
21	caryophyllenyl alcohol	1570	1562	0.4
22	caryophyllene oxide	1582	1577	26.1
23	globulol	1590	1584	4.3
24	humulene epoxide II	1608	1602	8.8
25	caria-4(12),8(13)-dien-5-ol	1639	1629	4.7
26	α-muurolol	1644	1635	2.5
27	β-eudesmol	1649	1644	1.5
28	a-cadinol	1652	1647	1.7
	Total identified			80.9
	Monoterpenes			9.0
	Oxygenated monoterpenes			4.0
	Sesquiterpenes			18.3
	Oxygenated sesquiterpenes			68.2
	Others			0.7

Notes: ^aOn a HP5-MS column, ^bAdams (2007), ^cAverage of 3 injections.

Results and discussion

Leaves of *P. myrsinites* presented 48.4% humidity. The essential oil had a slimy and translucent appearance and the yield was 0.1% (w/w, dry weight).

The chemical profile of the essential oil and the total ion chromatogram (TIC) are presented in Table 1 and Figure 1, respectively. The chromatogram contained forty-four peaks, but only twenty-eight of them were identified corresponding to 80.9% of the total essential oil composition. It is believed that the large amount of sesquiterpenes (86.5% of the oil) has disrupted greatly the identification process. In these conditions, the main components in the oil of P. mvrsinites were both oxygenated and non-oxygenated sesquiterpenes such as caryophyllene oxide (26.1%), humulene epoxide II (8.8%), β -caryophyllene (7.4%) and α -caryophyllene (5.4%). Castelo et al. (2012) previously reported for P. myrsinites the presence caryophyllene oxide, β -caryophyllene, β -guaiane α -humulene and viridiflorol as the major constituents, results quite similar with those observed in the present work. However, the essential oil composition was not completely elucidated and authors did not describe quantitatively each compound, since no quantification by GC-FID was performed in their study (14). Therefore, this is the first study to report the qualitative and quantitative composition of the essential oil of P. myrsinites.

Although the essential oil yield was not very high, it is possible to note that the amount of oil obtained is consistent with the genus production. For example, the oil from four species from the Amazon rainforest (*Psidium acutangulum* DC., *P. striatulum* DC., *P. guineense* Sw. and *P. guajava* L.) showed yields between 0.1% and 0.8% (15). In the Cerrado, the species *Psidium myrsinoides* O. Berg, a close related species of *P. myrsinites* presented an yield of 0.4% (16).

Psidium myrsinoides presented an essential oil very similar to that obtained from *P. myrsinites* in which β -caryophyllene and caryophyllene oxide were the main compounds (16).

The guava tree (*Psidium guajava*), native from Central America, is the best known species of this genus. Many articles on its essential oil were published. β -caryophyllene is the most common substance, however, there are others like α -pinene, 1,8-cineol and limonene in significant amounts (17). It needs to be considered that guava is a cultivated and domesticated species, and some changes in the methabolic pathways could have happened during this process.

In the Amazon some essential oil from *Psidium* were previously described. The chemical composition of the oils from *P. acutangulum* DC., *P. striatulum* Mart. ex DC., *and P. guineense* and *P. guajava* indicated, as main components, α -pinene, 1,8-cineole

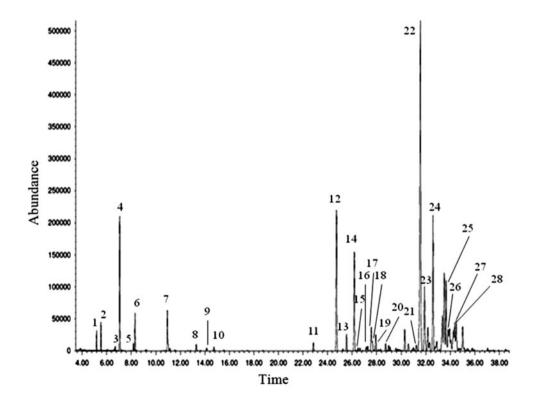


Figure 1. Total ion chromatogram of the essential oil from P. myrsinites.

(*P. acutangulum, P. guajava*); β -caryophyllene, α -selinene (*P. striatulum*); β -bisabolol, and limonene (*P. guineense*) (15).

The essential oils from *Psidium pohlianum* Berg and *Psidium guyanensis* Pers., both from the biome Caatinga, a Brazilian semiarid vegetation, were reported (18). It was observed a very close oil composition between these two species with 1,8-cineole being the main constituent in both plants, followed by the occurrence of α -pinene, β -eudesmol and γ -eudesmol.

In the Atlantic rainforest, it was found out α -thujene, 1,8-cineole and β -caryophyllene as major compounds in the oil from *Psidium cattleianum* Sabine (19).

The genus *Psidium* is largely distributed in all Brazilian territory and presented a predominance of mono and sesquiterpenes, with great variability, as reported above. The species of *Psidium* observed in the Cerrado biome showed a predominance of sesquiterpenes, mostly caryophyllene derivatives, which could be associated to the environmental conditions. However, there are still many other *Psidium* species to be chemically exploited to have a better understanding of this genus.

Acknowledgements

The authors are grateful to CAPES, CNPq, FAPERJ, UnB and EMBRAPA for financial support. The authors also would like to thank Sebastião for guidance in field works and Prof. Carolyn Proença (UnB) for the specimen identification.

Disclosure statement

No potential conflict of interest was reported by the authors.

References

- R.C. Forzza, J.F.A. Baumgratz, C.E.M. Bicudo, D.A.L. Canhos, A.A. Carvalho, M.A.N. Coelho, A.F. Costa, D.P. Costa, M.G. Hopkins, E.M. Leitman, I.G. Lohmann, E.N. Lughadha, L.C. Maia, G. Martinelli, M. Menezes, M.P. Morim, A.L. Peixoto, J.R. Pirani, J. Prado, L.P. Queiroz, S. Ssouza, V.C. Souza, J.R. Stenmann, L.S. Sylvestre, B.M.T. Walter and D.C. Zappi, *New brazilian floristic list highlights conservation challenges*. Bioscience, **62**, 1, 39–45 (2012).
- L.M.D.S. Aguiar, R.B. Machado and J. Marinho-filho, A diversidade biológica do Cerrado. In: Cerrado: ecologia e caracterização. Edits., L.M.S. Aguiar and A.J.A. Camargo, pp. 17–40, Embrapa Informação e Tecnológica, Brasília, DF (2004).
- R.C. Mendonça, J.M. Felpili, B.M.T. Walter, M.C. Silva junior, T.S. Filgueiras, P.E. Nogueira and C.W. Fagg, *Flora vascular do bioma Cerrado: Checklist com 12.356 espécies*. In: *Cerrado: ecologia e flora*, vol. 2. Edits., S.M. Sano, S.P. Almeida and J.F. Ribeiro, pp. 423–1279, Embrapa Informação Tecnológica, Brasília, DF (2008).
- C.A. Klink and R.B. Machado, Conservation of the Brazilian Cerrado. Conserv. Biol., 19, 707–713 (2005).

- M. Sobral, C. Proença, M. Souza, F. Mazine and E. Lucas, Myrtaceae. In: Lista de Espécies da Flora do Brasil. Jardim Botânico do Rio de Janeiro. http://floradobrasil.jbrj.gov. br/jabot/floradobrasil/FB10853> (27 February 2015).
- R.C. Frazon, L.Z.D.O. Campos, C.E.B. Proença and J.C. Sousa-silva, *Documentos 226 - Araçás do gênero Psidium: principais espécies, ocorrência, descrição e* usos., Embrapa Cerrados, Planaltina, DF (2009).
- A.L. Medina, L.I. Reckziegel, F.C. Chaves, M. Salvador, R.C. Zmbiazi, W.P. Dilva, L. Nora and C.V. Rombaldi, Araçá (Psidium cattleianum Sabine) fruit extracts with antioxidant and antimicrobial activities and antiproliferative effect on human cancer cells. Food Chem., 128, 916–922 (2011).
- G. Flores, K. Dastmalchi, S. Wu, K. Whalen, A.J. Dabo, K.A. Reynertson, R.F. Foronjy, J.M. D'Armiento and E.J. Kennelly, *Phenolic-rich extracts from the Costa Rican guava (Psidium friedchsthalianum) pulp with antioxidant and anti-inflammatory activity. Potential for COPD therapy.* Food Chem., **141**, 899–895 (2013).
- K.P. Mccook-Russell, M.G. Nair, P.C. Facey and C.S. Bowen-Forbes, *Nutritional and nutraceutical comparison* of Jamaican Psidium cattleianum (strawberry guava) and Psidium guajava (common guava) fruits. Food Chem., 134, 1069–1073 (2012).
- C. Damiani, E.V.B. Vilas-boas, E.R. Asquieri, M.E. Lage, R.A. Oliveira, F.A. Silva, E.M. Pinto, L.J. Rodrigues, E.P. Silva and N.R.R. Paula, *Characterization of fruits* from the savanna: Araça (Psidium guinnesis Sw.) and Marolo (Annona crassiflora Mart.). Ciência e Tecnologia de Alimentos, **31**, 723–729 (2011).
- H. Van Den Dool and P.D. Kratz, A generalization of the retention index system including linear temperature programmed gas-liquid partition chromatrography. J. Chromatrogr., 11, 463–471 (1963).
- R.P. Adams, *Identification of Essential Oil Components* by Gas Chromatography/Mass Spectrometry, 4th ed. Allured Publ. Corp, Carol Stream, IL (2007).
- NIST (National Institute of Standards and Technology), *PC Version of the NIST/EPA/NIH Mass Spectral Database*. U.S, Department of Commerce, Gaithersburg, MD (2008).
- A.A.V.M. Castelo, C.H.S.D. Menezzi and I.S. Resck, Seasonal variation in the yield and the chemical composition of essential oils from two brazilian native arbustive species. J. Applied Sci., 12, 753–760 (2012).
- J.D.D. Silva, A.I.R. Luz, M.H.L.D. Silva, E.H.A. Andrade, M.D.G.B. Zoghbi and J.G.S. Maia, *Essential* oils of the leaves and stems of four Psidium spp. Flavour Frag. J., 18, 240–243 (2003).
- M.O. Freitas, S.M.D. Morais and E.R. Silveira, *Volatile constituents of Psidium myrsinoides* O. Berg. J. Essen. Oil Res., 14, 364–365 (2002).
- M.E.A. Stefanello, A.C.R.F. Pascoal and M.J. Salvador, Essential oils from netropical Myrtaceae: chemical diversity and biological properties. Chem. Biodivers, 8, 73–94 (2011).
- M. Andrade-Neto, J.W.D. Alencar, A.N. Cunha, E.R. Silveira and T.G. Batista, *Volatile constituents of Psidium* pohlianum Berg and Psidium guyanensis Pers. J. Essen. Oil Res., 6, 299–300 (1994).
- F.A. Marques, E.P. Wendler, B.H.L.N.S. Maia, J.V. Coffani-nunes, J. Campana and P.G. Guerreiro, *Volatile* oil of Psidium cattleianum Sabine from the Brazilian Atlantic Forest. J. Essen. Oil Res., 20, 519–520 (2008).