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Antioxidants and sensory properties of the infusions of wild passiflora from Brazilian savannah: potential as functional beverages

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Abstract

BACKGROUND: The study of biodiversity for species recovery and sustainable use has encouraged research with plants from Brazilian savannah. We aimed to characterize chemical and sensory properties of infusions of passifloras, due to their potential as functional beverages. Infusions and hydroalcoholic extracts of four species of wild passifloras, three varieties of *Passiflora edulis* and a commercial passiflora tea were evaluated for total phenolics (TPs), total flavonoids (TFs), condensed tannins (CTs), and antioxidant activity (DPPH and FRAP). Free-choice Profile and acceptance, compared with green tea, were performed for sensory characterization.

RESULTS: In general, infusions had higher levels of TPs and CTs than hydroalcoholic extracts, which in turn had higher levels of TFs. Infusion of *P. nitida* showed higher amounts of TPs and antioxidant activity. Acceptance of passiflora infusions was similar or higher than that of green tea, except for *P. alata*. *P. setacea* presented a sensory profile similar to other commercial teas and higher acceptance by a group of consumers.

CONCLUSION: Passiflora infusions showed different degrees of suitability as acceptable functional beverage. Identification of phenolics and other bitter compounds is needed to understand the intense bitterness of *P. alata*, as it did not present the highest contents of TPs, CTs and TFs. © 2014 Society of Chemical Industry

Supporting information may be found in the online version of this article.

Keywords: passiflora; infusions; phenolic compounds; antioxidants; sensory analysis

INTRODUCTION

Studies for the recovery of fruits from Brazilian savannah are aligned to projects such as Biodiversity for Food and Nutrition (BFN), internationally coordinated by Bioversity International and implemented by the United Nations Program for the Environment (UNEP) and the United Nations Food and Agriculture Organization (FAO), approved by the Global Environment Fund (GEF).¹ These projects aim to promote the conservation and sustainable use of biodiversity in programs that contribute to the improvement of food security and human nutrition, by investigating food and nutritional properties of species related to agricultural biodiversity and by rescuing the cultural values played by many of those species. In this context, food materials rich in antioxidants and with records of popular use as functional foods have become objects of numerous studies for their chemical and nutritional characterizations.²

The Brazilian Agricultural Research Corporation (Embrapa), unit Cerrado, is focused on research on Brazilian savannah species and has the largest passiflora collection worldwide, with over 150 accessions, including species and varieties with functional and medicinal potential. Some commercial and wild species of passiflora are already part of the global ethno-pharmacological repertoire, which recommended leaves, flowers, roots and fruits to combat many different diseases, especially those of the nervous system.^{3,4}

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Some recent studies have evaluated the levels of phenolic compounds, antioxidant activity and some health promoting effects of passiflora pulp, especially the species *P. edulis* and *P. alata*.^{5–8} Other studies have evaluated these properties in extracts of leaves of passiflora.^{9–12} However, no studies evaluating the contents of phenolic compounds in combination with the sensory properties of the products of passiflora leaves are currently reported. It is well known that polyphenolics, apart from presenting valuable biological properties, impart a high sensory activity to foods. Flavonoid phenols have been indicated as the compounds that are mainly responsible for the bitter taste and the tactile sensation of astringency in beverages such as tea, cider, and red wine.¹³

For commercial exploration and aiming to foster consumption of passiflora species by their functional properties, further studies on sensory characteristics of leaf infusions and their acceptance are needed. Sensory analysis generates information that cannot be obtained instrumentally.¹⁴ Descriptive studies involving chemical and sensory characteristics with the acceptance of the products are extremely useful in generating information that drives breeding and plant cultivation systems with a focus on quality optimization and increased consumption. A recent study¹⁵ demonstrated that the growth system (organic or conventional, with or without shading) has an effect on the sensory quality of nectars made with the pulp of *P. edulis*.

Therefore, we aimed to determine antioxidant properties, sensory profile and acceptance of infusions made with leaves of different passiflora acessions from Brazilian savannah, evaluating their potential as a functional beverage.

EXPERIMENTAL

The leaves of passiflora were obtained from Embrapa Cerrado. Four wild species (*P. alata, P. tenuifila, P. nitida* and *P. setacea*) and three hybrid passionfruit varieties (*P. edulis* cv. BRS Ouro Vermelho, *P. edulis* cv. BRS Gigante Amarelo and *P. edulis* cv. BRS Sol do Cerrado) were studied. As a reference, a sample of commercial *Passiflora* spp. (useful part, stem and leaf; Santosflora Laboratory, São Paulo, SP, Brazil), bought in a popular pharmacy, was included in the experiment. Commercial green tea dried leaves were used as a sample in the acceptance test.

Preparation of infusions

Infusions were prepared at a ratio of 5 g of dry leaf per 1 L of boiling water. The concentration was previously established by a consumer focus group, where infusions with 3 g L⁻¹, 5 g L⁻¹ or 7 g L⁻¹ were tested. The leaves were added after water boiling temperature was achieved. The extraction lasted for 10 min. The infusions were completed for 1 L with hot water.

Preparation of hydroalcoholic extracts

Preparation of the extracts was performed according to Rudnicki *et al.*⁵ Five grams of dried leaves were mixed with 100 mL of aqueous ethanol [40% (v/v)]. The extraction was carried out with an Ultraturrax homogenizer (102E) at moderate speed for 3 min and refluxed in a glycerine bath (80 °C) for 30 min. The liquid was filtered under vacuum and rota-evaporated at 60 °C for about 5 min and the volume was completed with methanol to 50 mL. The sample was stored at -80 °C.

Infusions and hydroalcoholic extracts (HAs), three replications of each treatment, were analysed for total phenolics (TPs), condensed tannins (CTs), total flavonoids (TFs) and antioxidant activity by the

2,2'-diphenyl-1-picrylhydrazyl (DPPH) and ferric reducing antioxidant power (FRAP) assays. Analyses were carried out in triplicates.

Total phenolics

TPs were quantified using a modified Folin–Ciocalteu colorimetric method.¹⁶ A 0.2 mL aliquot of the water diluted extracts was added to a 15 mL tube and 0.2 mL of 1:10 Folin–Ciocalteu reagent: water solution was added to the mixture. The tube was allowed to stand at room temperature for 1 min. Then, 2 mL of 7.5% (w/v) Na₂CO₃ were added to the mixture. After 2 h at room temperature, absorbance was measured at 765 nm versus a blank. The results were expressed as mg of gallic acid equivalent (GAE) g⁻¹. For the hydroalcoholic extracts, dilution was carried out between 5 and 50 µL of extract in 1 mL, and for infusions, between 60 and 500 µL in 1 mL.

Condensed tannins

CTs were quantified using the vanillin method.¹⁷ Test tubes were covered with foil and added 5 mL of vanillin reagent (4 g of pure vanillin diluted to 56 mL in HCl, analytical purity, 37% w/w, and 83 mL of methanol). The reactants were preheated in a water bath at 30 °C for 30 min. We added 1 mL of HA extract or infusion, and 5 mL of 72% methanol in duplicate of the 'sample blank' and 1 mL in the 'vanillin blank'. The reaction was kept in the water bath for 20 min. Finally, the absorbance was measured at 510 nm. Results were expressed as mg of catechin equivalent (CE) g⁻¹.

Total flavonoids

TFs were determined according to Pereira *et al.*¹⁸ An aliquot of 5 mL was used for infusion analysis and volumes ranging from 100 to 400 μ L for HA extract analysis. Each sample was mixed with 500 μ L of methanol solution of aluminium chloride and completed to the volume of 10 mL with methanolic solution of acetic acid. The solution rested for 30 min, protected from light, and absorbance was read at 425 nm. Results were expressed in mg of quercetin equivalent (QE) g⁻¹.

Determination of the antioxidant activity by the DPPH assay

Antioxidant activity was determined by the DPPH radicalscavenging method according to Rufino *et al.*¹⁹ Aliquots of 0.1 mL of the previously diluted extracts were mixed to 3.9 mL of 0.06 mmol L⁻¹ of DPPH (initial absorbance of 0.756). The three dilutions applied consisted of volumes from 5 μ L to 350 μ L for HA extracts and from 30 μ L to 900 μ L of infusions, completed to 0.1 mL with methanol. The solutions were incubated at 25 °C for 25 min. Absorbance was recorded at 517 nm using methanol as blank. Total antioxidant activity was expressed as EC₅₀ (g g⁻¹ DPPH).

Determination of the antioxidant activity by the FRAP assay

Antioxidant activity by FRAP assay was determined according to Rufino *et al.*²⁰ A 90 mL aliquot of each aqueous extract dilution (from 12.5 mg mL⁻¹ to 500 mg mL⁻¹ for HA extracts and from 12.5 mg mL⁻¹ to 500 mg mL⁻¹ for infusions) was mixed with 270 mL of distilled water and 2.7 mL of FRAP reagent. Tubes were vortexed and incubated at 37 °C for 30 min. Absorbance was determined at 595 nm using FRAP reagent as blank and 500–2000 mmol L⁻¹ ferrous sulfate solutions substituting extracts as control. Results were expressed as µmol ferrous sulfate g⁻¹.

Chemical data were compared by ANOVA and when significant differences were identified we applied the Fisher test (P < 0.05).

Sensory evaluation

Infusions at a concentration of 5 g L^{-1} were established by the focus group qualitative method,¹⁴ with 10 tea consumers, who also chose, as the best method of consumption, sugar-free iced infusions, due to the healthy characteristic that is expected for a functional beverage.

Descriptive profile

Sensory profile was assessed by the Free-choice Profile method.²¹ The method was chosen to eliminate training, calibration and validation phases, which involve a large number of sessions for sensory evaluation and require a large number of samples. Seasonality and low availability act as limitation factors, once wild varieties and species are not commercially available, existing only in experimental fields.

Authorization for research involving human subjects, including written informed consent of panel participants, was previously obtained from the Ethical Commission of the Faculty of Health Sciences (CEP FS/UnB Number191/11).

Recruitment of the panelists

Assessors were recruited from students, staff and professors of the Catholic University of Brasilia (UCB). For the selection of panelists, a sequence of 12 triangular tests was applied comparing two infusions of *P. edulis*. BRS Sol do Cerrado sweetened with a difference of 0.8% in relation to sucrose content. This difference was previously verified by one triangular test (n = 30 panelists), and concluded as significant (P < 0.05). Panelists who correctly marked at least 60% of the tests were approved.²²

Lexicon development and individual sensory scale definition

Kelly's Repertory Grid Method.²³ was used to develop the list of descriptive terms to be employed by panelists for sample evaluation. All the combinations of pairs of samples were presented to panelists, individually, and they were asked to find as many differences and similarities among infusions as possible, in two sessions of evaluation. Once the list of terms had been developed, each term was located on a 9 cm line scale marked with the anchors 1 = 'null or very slight' to 9 = 'very intense'. In this way, individual score sheets were defined.

Sample evaluation

All samples were evaluated in a sequential monadic way, in two sessions. All samples were coded with three-digit numbers and presented at 9 ± 1 °C, in complete randomized design, with two replications for each treatment. Data were analyzed by generalized procrustes analysis (GPA), with Euclidean transformations of data by rotation, translation and self-scaling, followed by principal component analysis (PCA) of the consensus configuration (P < 0.05).

Acceptance

Students, professors and staff of UCB (n = 100), consumers of teas, at least twice a month, were selected. Samples were analyzed monadically in a randomized complete bock design,¹⁴ with a nine-point hedonic scale. Cluster analysis was run for a careful assessment of consumer preference. The dissimilarity coefficient used was Euclidean distance and to perform clustering, the Ward method, and automatic truncation. After clustering, means were subjected to ANOVA and to the Fisher test (P < 0.05). Green tea was also evaluated for a reference of acceptance. Statistical analyses

were performed using XLSTAT 2011 program (Addinsoft, Paris, France).

RESULTS AND DISCUSSION Chemical characterization

Infusions presented higher contents of TP than HA extracts, except for P. nitida (Table 1). Higher water extraction of TPs was also found by Bastos et al. working with yerba mate (Ilex paraguariensis) and green tea (Camelia sinensis) extracts.²⁴ Wild passifloras presented equal or higher amounts of TPs than the commercial species of Passiflora edulis. For infusions and HA extracts, the descending order of TP content was: P. nitida > P. setacea > P. tenuifila > P. edulis 'Gigante Amarelo', *P. edulis* 'Sol do Cerrado', *P. alata* \geq *P. edulis* 'Ouro Vermelho' > Passiflora ssp. Silva et al.¹⁰ found a content of 8.3 mg GAE g⁻¹ of TPs in the infusion of leaves of *Passiflora edulis* grown in Campinas, São Paulo, Brazil, closer to the lowest content found in this work, for Passiflora ssp. Colomeu et al.9 found lower values for aqueous (9.5 mg GAE g^{-1}) and methanolic (4.9 mg GAE g^{-1}) extracts of Passiflora alata Curtis, also grown in Campinas. It is interesting to notice that one serving (200 mL) of P. nitida infusion showed 43.60 \pm 0.50 mg of TPs, a level comparable with 40 mL of Moscatel wine studied by Silva et al.,²⁵ indicating the potential of passiflora infusions as non-alcoholic functional beverages.

Unlike TPs, the extraction of TFs was higher in HA extracts, except for *P. edulis* 'Sol do Cerrado', *P. edulis* 'Ouro Vermelho' and *Passiflora* ssp. For HA extracts, wild passiflora exceeded the levels found in the extracts of *P. edulis* and *Passiflora* ssp. The descending order was *P. nitida*, *P. alata*, *P. tenuifila*, *P. setacea* > *P. edulis* 'Gigante Amarelo' \geq *P. edulis* Ouro Vermelho \geq *P. edulis* 'Sol do Cerrado' > *Passiflora* ssp. We found low discrimination among the passiflora infusions for flavonoids, except for the lower content observed for *Passiflora* ssp. The concentrations of TFs of passiflora infusions were close to those found for four brands of green tea and three brands of black tea in the study by Pereira *et al.*¹⁸

CT contents were also higher in the infusions and in wild passifloras, presenting the following descending order: *P. nitida* > *P. alata* ≥ *P. tenuifila* ≥ *P. setacea* ≥ *P. edulis* 'Gigante Amarelo' ≥ *P. edulis* 'Sol do Cerrado' ≥ *Passiflora* ssp. ≥ *P. edulis* 'Ouro Vermelho'. Infusion of *P. nitida* showed higher contents of CT (17.81 ± 0.61 mg CE g⁻¹), compared with green tea, black tea and white tea, which presented, respectively, 12.52, 6.63 and 1.43 mg CE g⁻¹, in the study by Jacques *et al.*²⁶ The remaining infusions showed much lower values, between 0.2 and 1.5 mg CE g⁻¹.

The antioxidant capacity of passiflora extracts was mainly associated with the results of TPs and CTs and was higher for most infusions. The order of antioxidant activity for both extracts and for both methods was the same, with changes in significant differences among accessions. Analyzing infusions, the decreasing order of antioxidant activity by the FRAP assay was *P. nitida* > *P. tenuifila* > *P. setacea* ≥ *P. edulis* 'Gigante Amarelo' ≥ *P. edulis* 'Sol do Cerrado', *P. alata*, *P. edulis* 'Ouro Vermelho' ≥ *Passiflora* spp.

The structural diversity of phenolic compounds interfere in their physico-chemical behavior (solubility, partition coefficient, ionization constant), which may explain the different results found for antioxidant variables in infusions and HA extracts.²⁷

Our results suggest a higher antioxidant activity of passiflora infusions than that of berries, well known as rich in antioxidants. Souza *et al.*²⁸ determined antioxidant activity of Brazilian blackberry, red raspberry, strawberry, blueberry and sweet cherry fruits and found values between 2140 g g⁻¹ DPPH (blackberry) and 7775 g g⁻¹ DPPH (blueberry). In our work, passiflora infusions

Table 1. Total ph	enolics (TP), total fl	avonoids (TF), co	ndensed tannins	(mg g ⁻¹) and ar	ntioxidant activ	ities (AA) of infu	Table 1. Total phenolics (TP), total flavonoids (TF), condensed tannins (mg g ⁻¹) and antioxidant activities (AA) of infusions and hydroalcoholic (HA) extracts of passiflora leaves	olic (HA) extracts of p	assiflora leaves	
	Total pl	Total phenolics	Flavonoids	spior	Condense	Condensed tannins	AA-DPPH	Hdd	AA-	AA-FRAP
Passiflora	Infusion	HA extract	Infusion	HA extract	Infusion	HA extract	Infusion	HA extract	Infusion	HA extract
P. nitida	$43.60^{Aa} \pm 0.50$	$43.60^{Aa} \pm 0.50$ $46.05^{Aa} \pm 2.76$ $1.69^{Bab} \pm 0.11$	1.69 ^{Bab} ± 0.11	$3.07^{Aa} \pm 0.10$	$3.07^{Aa}\pm0.10\ 17.81^{Aa}\pm0.61\ 12.68^{Ba}\pm0.38$	$12.68^{Ba} \pm 0.38$	$472.53^{Ba} \pm 61.00$	$262.84^{Aa} \pm 22.17$	$262.84^{Aa} \pm 22.17$ 589.65 ^{Aa} ± 14.39	$558.00^{Aa} \pm 44.06$
P. alata	17.87 ^{Ad} ± 0.31	$17.87^{Ad} \pm 0.31$ 13.72 ^{Bde} ± 1.25 1.59 ^{Babc} ± 0.00	$1.59^{\text{Babc}} \pm 0.00$	$2.93^{Aab} \pm 0.15$ $1.47^{Ab} \pm 0.01$ $0.63^{Bc} \pm 0.00$	$1.47^{Ab} \pm 0.01$	$0.63^{Bc} \pm 0.00$	$1531.17^{Ac} \pm 159.06$	$2179.02^{Bf} \pm 71.56$	$2179.02^{Bf} \pm 71.56$ 178.33 ^{Ae} ± 12.25 123.70 ^{Bdef} ± 14.85	123.70 ^{Bdef} ± 14.85
P. tenuifila	23.66 ^{Ac} ± 0.89	$23.66^{Ac} \pm 0.89$ 18.82 ^{Bc} ± 2.73 1.58 ^{Babc} ± 0.23	$1.58^{\text{Babc}} \pm 0.23$	$2.93^{Aab} \pm 0.19$ 1.36 ^{Abc} ± 0.14 0.65 ^{Bc} ± 0.04	$1.36^{Abc} \pm 0.14$	$0.65^{Bc} \pm 0.04$	$652.15^{Aa} \pm 11.26$	783.74 ^{Bb} ± 22.83	$783.74^{Bb} \pm 22.83$ $352.20^{Ab} \pm 11.14$	$262.10^{Bb} \pm 13.87$
P. setacea	27.78 ^{Ab} ± 1.62	$27.78^{Ab} \pm 1.62$ 22.17 ^{Bb} ± 2.39 1.45 ^{Bbc} ± 0.02	$1.45^{\text{Bbc}} \pm 0.02$	$3.08^{Aa} \pm 0.09$	$1.08^{Abc} \pm 0.04$	$0.55^{Bcd} \pm 0.05$	$3.08^{Aa} \pm 0.09$ $1.08^{Abc} \pm 0.04$ $0.55^{Bcd} \pm 0.05$ $1055.08^{Ab} \pm 178.49$	$1121.80^{Ac} \pm 78.78$ $239.89^{Ac} \pm 10.03$	$239.89^{Ac} \pm 10.03$	$207.54^{Bc} \pm 9.49$
<i>P. edulis</i> 'G. Amarelo' 18.35 ^{Ad} \pm 0.46 13.93 ^{Bd} \pm 0.41 1.51 ^{Babc} \pm 0.13	10° 18.35 ^{Ad} ± 0.46	$13.93^{Bd} \pm 0.41$	$1.51^{\text{Babc}} \pm 0.13$	$2.61^{Abc} \pm 0.20$	$1.29^{Abc} \pm 0.11$	$0.37^{\text{Bde}} \pm 0.01$	$2.61^{Abc} \pm 0.20$ 1.29 ^{Abc} ± 0.11 0.37 ^{Bde} ± 0.01 1203.59 ^{Abc} ± 70.28	$1365.01^{Bd} \pm 19.44$ 221.89 ^{Acd} ± 8.29	$221.89^{Acd} \pm 8.29$	$142.19^{Bd} \pm 12.62$
P. edulis 'S. do	$18.76^{Ad} \pm 0.47$	$18.76^{Ad} \pm 0.47$ 14.68 ^{Bd} ± 1.23	$1.77^{Aa} \pm 0.01$	$2.28^{Ac} \pm 0.25$	$1.08^{Abc} \pm 0.16$	$0.30^{Be} \pm 0.01$	$2.28^{Ac} \pm 0.25$ 1.08 ^{Abc} ± 0.16 0.30 ^{Be} ± 0.01 1230.65 ^{Abc} ± 92.90	$1131.62^{Ac} \pm 15.21$	$1131.62^{Ac} \pm 15.21 213.08^{Ad} \pm 19.64 126.36^{Bde} \pm 9.85$	$126.36^{Bde} \pm 9.85$
Cerrado'										
<i>P. edulis</i> 'O. Vermelho' 15.75 ^{Ae} \pm 0.76 10.72 ^{Be} \pm 0.45 1.36 ^{Ac} \pm 0.20	io' 15.75 ^{Ae} ± 0.76	$10.72^{Be} \pm 0.45$	$1.36^{Ac} \pm 0.20$	$2.38^{Ac} \pm 0.08$	$1.05^{Ac} \pm 0.16$	$0.21^{Be} \pm 0.04$	$2.38^{Ac} \pm 0.08$ $1.05^{Ac} \pm 0.16$ $0.21^{Be} \pm 0.04$ $2219.87^{Ad} \pm 91.34$ $2697.53^{Bg} \pm 132.51$ $136.12^{Af} \pm 8.40$	$2697.53^{Bg} \pm 132.51$	$136.12^{Af} \pm 8.40$	$103.66^{\text{Bef}} \pm 4.41$
Passiflora ssp. (commercial)	11.71 ^{Af} ± 0.33	6.59 ^{Bf} ± 1.13	0.59 ^{Ad} ± 0.03	0.57 ^{Ad} ± 0.01	1.26 ^{Abc} ± 0.13	$0.57^{Ad} \pm 0.01 1.26^{Abc} \pm 0.13 0.99^{Bb} \pm 0.02$	2758.28 ^{Be} ± 468.80	$2758.28^{Be}\pm468.80 1876.49^{Ae}\pm152.70 130.22^{Af}\pm14.35$	130.22 ^{Af} ± 14.35	94.11 ^{Bf} ± 1.64
Total phenolics were determined as gallic acid equivalents; flavonoids as quercetin equivalents; condensed tanni AA-FRAP as µmol ferrous sulfate equivalent g^{-1} . Means followed by different small letters in a column differ according to the Fisher test ($P \leq 0.05$). Means followed by different capital letters in a row, for each type of analysis, differ according to the <i>t</i> test ($P \leq 0.05$). AA, antioxidant activity; DPPH, 2,2'-diphenyl-1-picrylhydrazyl; FRAP, ferric reducing antioxidant power.	re determined as g errous sulfate equiv different small lett different capital le' ivity; DPPH, 2,2'-di	gallic acid equiva valent g ⁻¹ . ers in a column d tters in a row, for	lents; flavonoids liffer according tc each type of ana /drazyl; FRAP, ferr	ds as quercetin equivalents to the Fisher test ($P \leq 0.05$) nalysis, differ according to the erric reducing antioxidant p	uivalents; cond $P \le 0.05$). ding to the <i>t</i> te oxidant power.	densed tannins st (P ≤ 0.05).	Total phenolics were determined as gallic acid equivalents; flavonoids as quercetin equivalents; condensed tannins as catechin equivalents; AA-DPPH was determined as the EC ₅₀ , g g ⁻¹ DPPH; and AA-FRAP as µmol ferrous sulfate equivalent g ⁻¹ . Means followed by different small letters in a column differ according to the Fisher test ($P \le 0.05$). Means followed by different capital letters in a row, for each type of analysis, differ according to the t test ($P \le 0.05$). AA, antioxidant activity; DPPH, 2.2'-diphenyl-1-picrylhydrazyl; FRAP, ferric reducing antioxidant power.	ts; AA-DPPH was det	ermined as the EG	0, g g ⁻¹ DPPH; and

presented values varying from $472.53 \pm 61.00 \text{ g g}^{-1}$ DPPH to $2219.87 \pm 92.90 \text{ g g}^{-1}$ DPPH, indicating higher antioxidant capacities. Regarding the RAP assay, the antioxidant activity found for *P. nitida* infusion (589.65 ± 14.39 µmol ferrous sulfate g⁻¹) was about 17-fold the antioxidant activity of strawberries, a well-known source of phenolic compounds.²⁹

The main groups of polyphenols are defined according to their carbon skeleton. From a sensory point of view, larger molecules tend to be less bitter and more astringent.¹³ Besides their antioxidant capacity, phenolic compounds have attracted considerable interest because of their influence on sensory characteristics such as color, bitterness and astringency and in formation of certain flavors. Only in studies with wines are such assessments more exploited.³⁰

Sensory profile

According to selection criteria and evaluation of the residual variance of each panelist from GPA, we selected nine assessors. Attributes raised by the panel are shown in Table 2, as well as their descriptions and identifications of panelists.

In relation to appearance, all panelists indicated the same attributes, which were yellow color and translucency. Regarding odor, only sweet odor was mentioned by all panelists. Earthy odor, odor of green fruit, artificial odor of passion fruit and odor of flowers were cited by one assessor each. Bitterness was the flavor attribute cited by all panelists. Sweetness was cited by six panelists. Flavor of gilo (*Solanum gilo*), flavor of green fruit and flavor of the infusion of passion fruit leaf had one mention each.

According to the overlap of each attribute (Fig. 1), high consensus was observed for bitterness, yellow color and translucency. Sweet odor and odor of honey also showed a quite significant consensus and it is still possible to see a positive association between these attributes. Lack of consensus was visible for sweetness, which may indicate that other attributes are confounding panelists for their quantification, possibly bitterness.

PCA (Fig. 2a and b) of the consensus configuration presented 75.45% of explanation for D1 (55.27%) and D2 (20.18), and showed the formation of three distinct groups of samples. Sufficiently far from the other samples, *P. alata* appeared in the first quadrant, positively correlated with the attributes bitterness, flavor of green tea, flavor of gilo and, as a contradiction between tastes and odors, the odor of honey and sweet odor, as indicated by the proximity of the sample to those attributes.

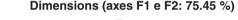
P. setacea, also separated from the others, was strongly characterized by the attributes odor and flavor of black tea, odor of yerba mate tea and yellow color, indicating a higher sensory similarity with other commercial teas consumed on a large scale.

The remaining samples were very close together, which suggests similar sensory profiles. For these samples, the attributes that stood out were: sweetness, odor of green leaf, earthy odor, odor of flowers and translucency.

According to chemical analysis, the infusion with higher amounts of TP and condensed tannins was *P. nitida* (Table 1). However, that infusion was not superior for bitterness nor astringency, whose closest attribute, among the reported ones, was 'flavor of green fruit'. We believe that the phenolic profile of *P. alata* or the presence of other bitter compounds may provide explanation for their sensory differentiation from other infusions, with rather sharp bitterness (Fig. 2), although this infusion did not show a high amount of TP (Table 1).

For astringency, the molecular weight seems to be important for its perception and the interactions of tannins with salivary proteins

Attribute	Description	Panelists
Appearance		
Yellow color	Intensity of yellow color	A, B, C, D, E, F, G, H,
Translucent	Easy passage of light by the liquid	A, B, C, D, E, F, G, H,
Odor		
Sweet odor	Refers to sweet odor	A, B, C, D, E, F, G, H,
Artificial odor of passion fruit	Characteristic odor of unnatural passion fruit	A
Earthy odor	Refers to characteristic wet earthy odor	G
Odor of leaf	Characteristic odor of leaf	B, D, F, I
Odor of black tea	Characteristic odor of black tea	A, D, E, F, G
Odor of honey	Characteristic odor of natural honey	A, B, G
Odor of green fruit	Fruit flavor that have not matured	G
Odor of yerba mate tea	Characteristic odor of yerba mate tea	C, I
Odor of flowers	Odor of citric flowers	E
Odor of dry leaf of passion fruit	Odor of sun-dried passion fruit leaf	C, D, I
Flavor		
Bitterness	Refers to bitter taste	A, B, C, D, E, F, G, H,
Sweetness	Refers to sweet taste	A, C, D, F, G, I
Flavor of gilo	Bitter taste that remind flavor of gilo	F
Flavor of green fruit	Flavor of something astringent	F
Flavor of green tea	Flavor that reminds taste of green tea	B, C, D, H
Flavor of black tea	Characteristic flavor of black tea	B, C, D, G, H
Flavor of yerba mate tea		
Flavor of tea of passion fruit	Characteristic flavor of yerba mate tea C, F Characteristic flavor of tea of passion fruit homemade I	



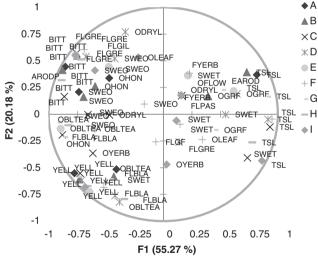


Figure 1. Overlapping of individual and consensus configurations (panelists A–I). Attributes are: YELL, yellow color; TSL, translucency; SWEO, sweet odor; ARODP, artificial odor of passion fruit; EAROD, earthy odor; OLEAF, odor of leaf; OBLTEA, odor of black tea; OHON, odor of noney; OGRF, odor of green fruit; OYERB, odor of yerba mate tea; OFLOW, odor of flower; ODRYL, odor of dry leaf of passion fruit; BITT, bitterness; SWET, sweetness; FLGIL, flavor of gilo; FLGF, flavor of green fruit; FLGRE, flavor of green tea; FLBLA, flavor of black tea; FYERB, flavor of yerba mate tea; FLPAS, flavor of tea of passion fruit.

result in the perception of astringency.³¹ In the method of vanillin to determine condensed tannins, both the leuco-anthocyanidins (catechins) and pro-anthocyanidins (condensed tannins) react with vanillin in the presence of HCI to produce a red condensation product, which is detected spectrophotometrically.¹⁷ Despite

the high concentration of tannins in *P. nitida*, the degree of condensation cannot be known by this method. The high value of condensed tannins could be related to the presence of catechins and/or tannins with low degree of polymerization, which does not affect strongly astringency. In fact, the 'flavor of green fruit' attribute is located near the origin (Fig. 2), indicating the low importance of this characteristic in discriminating the samples.

Acceptance

The results of acceptance were subjected to cluster analysis resulting in two groups of consumers, with 60 and 40 panelists (Table 3). In general, passiflora infusions were more accepted than green tea, except for the infusion of *P. alata*, the least accepted among all samples.

From the segmentation of consumers, it was observed that the means of acceptance of the Cluster 1 were significantly higher than those of Cluster 2. For Cluster 1, the acceptance of *P. setacea* was statistically higher than that of *Passiflora* spp. and green tea. The other samples showed intermediate acceptances. For Cluster 2, there was higher acceptance for *P. nitida, Passiflora* spp., and *P. edulis* of the three cultivars. The acceptance of *P. nitida* was significantly higher than that of *P. setacea* and *P. tenuifila*, which were in turn more acceptable than green tea and *P. alata*.

Despite the option of the focus group by the sensory analysis of unsweetened iced teas, the lack of sugar in the infusions may be one strong cause for not so high acceptance means, as we observed. Commercial iced teas are sweetened with sugar or sweeteners. People who consume unsweetened teas are a small and specific group of consumers. Further studies could be recommended with sweetened infusions. Many discussions regarding health, however, are the backdrop for the development of this product, since sugar sweetened teas have a high glycemic

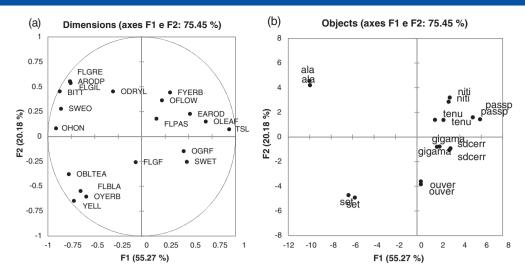


Figure 2. Principal component analysis (PCA) of the data generated by free profile factorial plan (D1 × D2). (a) Configuration of the observations; (b) configuration of the variables. The attributes are as in Fig. 1. The variables are: *P. nitida* (niti), *P. alata* (ala); *P. tenuifila* (tenu); *P. setacea* (set); *P. edulis* 'Gigante Amarelo' (gigama), *P. edulis* 'Sol do Cerrado (sdcerr); *P. edulis* 'Ouro Vermelho (ouver); *Passiflora* ssp. (passp).

Table 3. Acceptance of passiflora infusions				
Class	Cluster 1 (<i>n</i> = 40)	Cluster 2 (<i>n</i> = 60)	Total	
P. nitida P. alata P. tenuifila P. setacea P. edulis 'G. Amarelo'	$\begin{array}{c} 5.875^{aAB}\pm1.65\\ 2.80^{aC}\pm2.02\\ 5.95^{aAB}\pm1.65\\ 6.23^{aA}\pm1.37\\ 5.80^{aAB}\pm1.50\end{array}$	$\begin{array}{c} 4.50^{\text{bA}}\pm1.95\\ 1.80^{\text{bD}}\pm1.38\\ 3.57^{\text{bC}}\pm2.03\\ 3.77^{\text{bBC}}\pm1.97\\ 3.98^{\text{bABC}}\pm1.80\end{array}$	$5.05^{A} \pm 1.95$ $2.20^{C} \pm 1.72$ $4.52^{A} \pm 2.21$ $4.75^{A} \pm 2.12$ $4.71^{A} \pm 1.89$	
P. edulis 'S. do cerrado' P. edulis 'O. vermelho'	$6.05^{aAB} \pm 1.52$ $5.98^{aAB} \pm 1.51$	$4.25^{bAB} \pm 1.64$ $4.03^{bABC} \pm 1.79$	$4.97^{A} \pm 1.81$ $4.81^{A} \pm 1.93$	
<i>Passiflora</i> ssp. Green tea	$5.35^{aB} \pm 2.00$ $5.45^{aB} \pm 1.83$	3.97 ^{bAB} C ± 2.20 2.17 ^{bD} ± 1.32	$4.52^{A} \pm 2.20$ $3.48^{B} \pm 2.22$	

Means followed by the same lowercase letters in the same row (just between cluster 1 and cluster 2) do not differ according to the *t* test ($P \le 0.05$).

Means followed by the same uppercase letters in the same column (within each cluster) do not differ according to the Fisher test ($P \le 0.05$).

index.³² The addition of synthetic sweeteners could be contradictory to consumers that seek to use those teas as functional beverages. Furthermore, studies indicate that the application of the natural stevia-based sweetener implies intense bitter aftertaste, which could increase the rejection of the product.³³ Studies show that one mechanism for increasing the liking of food is the provision of information reporting the health benefits.^{34–36} Cox *et al.*³⁴ found that health information influenced positively acceptance responses of Brussels sprouts. Similarly, Sabbe *et al.*³⁶ observed a positive effect of health information about acai (*Euterpe oleracea*) juices on hedonic and sensory measures, as well as on health and nutrition-related attribute perceptions and purchase intention.

CONCLUSIONS

The infusion of *P. nitida* stood out from a chemical point of view, presenting relevant amounts of phenolics and high antioxidant

activity. It was also preferred by one group of consumers. From a sensory point of view, infusion of P. alata was less accepted. Iced infusions of passiflora were equally or more accepted than iced green tea, except for P. alata in Cluster 1, suggesting that these drinks have sensory potential to be consumed as a functional beverage, like green tea. Bitterness was the attribute most positively associated with *P. alata* and may be possibly the driver of dislike of these iced infusions. P. setacea, with higher acceptance for Cluster 1, already stood out for greater sensory similarity to other commercial teas consumed on a large scale. Further studies are suggested for the identification of phenolic and other bitter compounds present in infusions of passiflora from Brazilian savannah, in order to assign specific structures, probably in higher concentrations in P. alata, and responsible for its higher bitterness, since this sample showed no highlight in terms of TP, condensed tannins and total flavonoids. It was concluded that passiflora infusions showed different degrees of suitability as functional beverage and have potential for the development of other form of consumption, beyond the passiflora fruits. Consumption and increased acceptance may be stimulated by disseminating information about their levels of antioxidants and the discovery of new properties, as researches to evaluate existing claims from popular use are carried out.

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SUPPORTING INFORMATION

Supporting information may be found in the online version of this article.

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