

Coconut genome size determined by Aow cytometry: Tall versus Dwarf types

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ABSTRACT. Coconuts (Cocos nucifera L.) are tropical palm trees that are classized into Tall and Dwarf types based on height, and both types are diploid (2n = 2x = 32 chromosomes). The reproduction mode is autogamous for Dwarf types and allogamous for Tall types. One hypothesis for the origin of the Dwarf coconut suggests that it is a Tall variant that resulted from either mutation or inbreeding, and differences in genome size between the two types would support this hypothesis. In this study, we estimated the genome sizes of 14 coconut accessions (eight Tall and six Dwarf types) using Aow cytometry. Nuclei were extracted from leaf discs and stained with propidium iodide, and Pisum sativum (2C = 9.07 pg DNA) was used as an internal standard. Histograms with good resolution and low coef¿cients of variation (2.5 to 3.2%) were obtained. The 2C DNA content ranged from 5.72 to 5.48 pg for Tall accessions and from 5.58 to 5.52 pg for Dwarf accessions. The mean genome sizes for Tall and Dwarf specimens were 5.59 and 5.55 pg, respectively. Among all accessions, Rennel Island Tall had the highest mean DNA content (5.72 pg), whereas West African Tall had the lowest (5.48 pg). The mean coconut genome size (2C = 5.57 pg,

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corresponding to 2723.73 Mbp/haploid set) was classized as small. Only small differences in genome size existed among the coconut accessions, suggesting that the Dwarf type did not evolve from the Tall type.

Key words: Cocos nucifera L.; DNA content; C-value; Coconut accessions; Palm tree

INTRODUCTION

The coconut (Cocos nucifera L.) is a diploid palm species with 2n = 2x = 32 chromosomes. This seed-propagated, slow-growing tree occurs naturally throughout the tropics between 20°N and 20°S latitude (Batugal et al., 2009), and it is considered the most economically important tropical species (Persley, 1992). Coconuts comprise two main types: Tall and Dwarf (Purseglove, 1975). Tall coconuts are characterized by heights up to 30 m, and they are predominantly allogamous with medium- to large-sized fruits. The variety can produce 60 to 80 fruits tree⁻¹·year⁻¹ that are harvested 11 to 12 months after Àowering. Moreover, the Tall coconut ecotypes are named according to the geographic region or country in which they evolved.

Dwarf coconuts are autogamous, and Àowering is initiated after an average of 3 years. The plants live approximately 30 to 40 years, and the mean yield is 150 to 200 fruits-tree⁻¹-year⁻¹. The trees are smaller than Tall coconut trees, reaching a height of 10 to 12 m, and the fruits are smaller. Dwarf coconuts are classized into three groups based on fruit color: green, yellow, and red (Harries, 1978).

The origin and dispersal history of coconuts is still controversial (Clement et al., 2013; Baudouin et al., 2014; Harries and Clement, 2014). Several hypotheses have been developed to explain the origin of Dwarf coconuts, and one hypothesis suggests that the Dwarf type is a variant of the Tall coconut that resulted from either mutation or inbreeding (Swaminathan and Nambiar, 1961). Several authors have observed only minor differences between the karyotypes of the two coconut types, but the results do not support the idea that the Dwarf type evolved from the Tall type (Raveendranath and Ninan, 1973; Sisunandar and Adkins, 2007).

The estimation of genome size based on DNA content is important for genomic characterization. Moreover, this estimation is not only important for genetic improvement (Leus et al., 2009), cell biology, systematic biology, and taxonomy research (Loureiro, 2007), but also for evolutionary studies (Baack et al., 2005). To date, few studies have estimated the DNA content in coconuts via Àow cytometry, and to our knowledge, none have compared the genome sizes between Dwarf and Tall types. In this study, Àow cytometry was used to estimate the genome size of 14 coconut genotypes (eight Tall and six Dwarf types) to determine whether there are genome size differences that support the theory that the Dwarf type evolved from the Tall type.

MATERIAL AND METHODS

Plant material

Leaf samples from 14 coconut accessions (six Dwarf and eight Tall accessions) were used to analyze DNA content. These accessions are maintained in the Active Germplasm Bank of Coconut, a living collection grown at Embrapa Tabuleiros Costeiros, Aracajú, Sergipe, Brazil. The analyses were performed at the Laboratory of Plant Breeding, Universidade Estadual do Norte Fluminense Darcy Ribeiro, located in Campos dos Goytacazes, Rio de Janeiro, Brazil.

Flow cytometry analysis

Leaf samples were collected from adult plants, packed, transported to the laboratory, and stored in a refrigerator for up to 48 h. To prepare samples for cytometer readings, leaf discs (approximately 0.5 cm²) of each accession were used to extract nuclei (Galbraith et al., 1983; Otto, 1990). The leaf samples were placed in 500 μ L extraction buffer (CyStain UV Precise P, Partec GmbH, Münster, Germany) on Petri dishes, and they were chopped with a blade for 90 s at 25°C to suspend nuclei.

The samples were ¿Itered through 50-mm membranes, and 2 mL staining solution (CyStain UV Precise P, Partec GmbH) containing propidium iodide and RNase was added to each tube. The tubes were then incubated at 4°C for 30 min. The same procedure was performed with Pisum sativum leaÀets (2C = 9.07 pg), which had been de¿ned as the best internal standard at a preliminary stage. Samples containing the suspended nuclei of each coconut accession and the internal standard were prepared together for further analysis.

After adjusting the equipment, the samples were analyzed with a Àow cytometer (Partec PA II; Partec GmbH). Five readings (replicates) per accession were prepared, and 10,000 nuclei were counted per analysis. Data were collected and mean peak analyses were performed using the Flowmax software (Partec GmbH).

Genome size was calculated (Doležel and Bartos, 2005), and the data were tested by ANOVA. The mean values of the accessions were grouped, based on the Scott and Knott test results at 5% probability, using the Genes statistical package (Cruz, 2006). The genome size in megabase pairs (Mbp) was estimated using the conversion factor 1 pg = 978 Mbp (Doležel et al., 2003).

RESULTS

The DNA content differed signi¿cantly among the coconut accessions (P < 0.01). Of the 14 accessions studied, the highest DNA content was observed in Rennel Island Tall (5.72 pg/2C or 2.86 pg/C), and the smallest value was found in West African Tall (5.48 pg/2C or 2.74 pg/C). The coconut accessions formed four groups based on their 2C DNA content (Table 1). High-resolution histograms (Figures 1 and 2) were obtained for all samples, and the coef¿cients of variation (CVs) ranged from 2.5 to 3.2% (Table 1).

Coconut accessions	Symbol	Genome size		
		pg/2C	CV (%)	Mbp/haploid set
Rennel Island Tall	RIT	5.72 a	3.1	2797.08
Polynesian Tall	PT	5.68 b	2.8	2777.52
Rotuma Tall	RT	5.66 b	3.2	2767.74
Praia do Forte Tall	PFT	5.60 c	3.2	2738.40
Vanuatu Tall	VT	5.54 c	2.7	2709.06
Tonga Tall	TT	5.54 c	3.0	2650.38
Malayan Tall	MT	5.50 d	2.9	2689.50
West African Tall	WAT	5.48 d	2.9	2679.72
Mean Tall	-	5.59	-	2733.51
Gramame Red Dwarf	GRD	5.58 c	2.7	2728.62
Gramame Yellow Dwarf	GYD	5.56 c	2.8	2718.84
Malayan Yellow Dwarf	MYD	5.56 c	2.5	2718.84
Malayan Red Dwarf	MRD	5.56 c	2.8	2718.84
Cameroon Red Dwarf	CRD	5.54 c	3.0	2709.06
Jiqui Green Dwarf	JGD	5.52 d	2.5	2699.28
Mean Dwarf	-	5.55	-	2713.95
Overall mean	-	5.57	-	2723.73

Table 1. Mean genome size of coconut accessions estimated in picograms (pg), in megabase pairs (Mbp), and the coef¿cient of variation (CV).

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Figure 1. Flow cytometry histograms of Tall accessions. The ¿rst peak represents the Tall accession, and the second peak represents the internal standard (Pisum sativum L.). A. PFT; B. MT; C. WAT; D. PT; E. RIT; F. RT; G. TT; H. VT.

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Figure 2. Flow cytometry histograms of Dwarf accessions. The ¿rst peak represents the Dwarf accession, and the second peak represents the internal standard (Pisum sativum L.). A. MRD; B. JGD; C. GYD; D. GRD; E. MYD; F. RCD.

The mean DNA content for Tall accessions was 5.59 pg/2C, corresponding to 5.47 Gb/2C and 2733.51 Mbp/1C. Among the Dwarf accessions, Red Dwarf from Gramame, Brazil, had the highest DNA content (5.58 pg/2C), and Green Dwarf from Jiqui, Brazil, had the lowest (5.52 pg/2C). The 2C DNA content showed greater variation among the Tall group than among the Dwarf group (Table 1). The Dwarf accessions had low intra-group variation with non-signi¿cant differences, with the exception of Green Dwarf from Jequi, Brazil.

DISCUSSION

Few studies have determined DNA content via Àow cytometry in Arecaceae species. In our study, the mean 2C DNA content for coconuts was 5.59 and 5.55 pg for Tall and Dwarf

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accessions, respectively. One of the ¿rst studies of DNA content in palm trees estimated a 2C value of 7.10 pg for coconuts using Feulgen densitometry (Röser et al., 1997). DNA content is generally estimated based on nuclei extracted from fresh, young leaves (Doležel and Bartos, 2005). Coconut germplasm is maintained in vitro or in vivo, and in the few coconut studies using cytometry, nuclei were extracted from plants grown in vitro. In this study, nuclei were extracted from the mature leaves of trees that were stored in a coconut germplasm collection. Nuclear suspension quality is judged based on analyses of histograms depicting the relative DNA content. The histogram should contain minimal amounts of background debris, and the variation should be low (Doležel and Bartos, 2005). The variation is usually expressed as the CV, and CV values in our study varied from 2.5 to 3.0%. Moreover, high-quality analyses exhibited CV values of 1.0 to 2.0%, and routine analyses showed CV values of approximately 3.0% (Marie and Brown, 1993). However, CV values below 5.0% were considered satisfactory (Galbraith et al., 2002), and the authors also stated that in some recalcitrant species, such as coconuts, the recommended CV values are dif¿cult to obtain.

The mean 2C DNA content of the Tall group was 5.59 pg, which corresponded to 5467.02 Mbp (Doležel et al., 2003), and this was higher than the mean of the Dwarf accessions (5.55 pg). In Lathyrus, 2C DNA values were higher for outbreeding perennials and lower for inbreeding annuals (Rees and Hazarika, 1967). In this study, we observed that there was greater variation in 2C DNA values among allogamous and perennial Tall types than among autogamous Dwarf types.

The mean 2C DNA content for the Dwarf group was 5.55 pg, which corresponded to 5427.9 Mbp (Doležel et al., 2003). A DNA content value of 5.60 pg/2C was estimated for Malayan Yellow Dwarf, which was similar to the value found in this study (2C = 5.55 pg) (Sandoval et al. 2003) even though the authors used Hordeum vulgare (2C = 11.12 pg) and Petunia hybrida (2C = 2.85 pg) as internal reference standards. The internal reference standard should ideally have a similar, but not overlapping, genome size compared to the species under study (Doležel and Bartos, 2005). Arabidopsis thaliana, Solanum esculentum, and P. sativum were indicated as the most appropriate internal reference standards (Praça-Fontes et al., 2011).

The Dwarf accessions showed little variation of 2C DNA content, indicating that this coconut group is more homogenous than the Tall group. Genetic divergence was investigated among Dwarf genotypes, and the results indicated that Green Dwarf is most similar to Cameroon Red Dwarf and most distant from Malayan Yellow Dwarf (Cambuí et al., 2007). In this study, the DNA content of Green Dwarf differed from that of all other Yellow or Red Dwarfs. A study based on RAPD analyses showed that Red Dwarfs are genetically distinct from the Green and Yellow Dwarfs (Daher et al., 2002). Moreover, the Red, Yellow, and Green Dwarf varieties differ mainly in their fruit color, which is determined by two loci (R and G). The genotypes of Yellow, Red, and Green Dwarfs are rrgg, R-gg, and rrG-, respectively (Bourdeix, 1988). Thus, large differences in DNA content among the Red, Green, and Yellow Dwarf varieties were not expected.

The mean 2C DNA content of coconut species was 5.57 pg/2C, corresponding to 5447.46 Mbp (Doležel et al., 2003). The value calculated in this study was slightly different from values reported for other Arecaceae species (Bharathan et al., 1994; Rival et al., 1997). The ¿rst study of Arecaceae was carried out using Feulgen densitometry to investigate 83 species that represented 53 genera, and the 2C DNA content ranged from 1.90 pg in a diploid species (Phytelephas aequarorialis Spruce) to 78.20 pg in a species with a high polyploidy level (38x) (Voanioala gerardii J. Dransf.) (Röser et al., 1997). Recent studies using Àow cytometry have reported 2C values of 17.50 pg for Trithrinax campestris, which is considered high for palm trees (Gaiero et al., 2012), and 5.80 pg for Acrocomia aculeata (Abreu et al., 2011), which is comparable to the results of

this study. The 2C content was reported for Elaeis species, and it ranged from 3.64 to 4.10 pg (Madon et al., 2008). However, values higher than 8.00 pg (8.70 to 9.41 pg) were reported for three Euterpe species (Oliveira, 2011), and the DNA content ranged between 6.46 and 6.96 pg in ¿ve Oenocarpus species (Oliveira, 2012).

Based on the results, one might conclude that the coconut genome is small (5.57 pg/2C and 2723.73 Mbp/per haploid set). Leitch et al. (1998) de¿ned species with C-values and genome sizes \leq 1.40 and \leq 3.50 pg as having very small and small genomes, respectively. Similarly, species with C-values and genome sizes \geq 14.0 and \geq 35.0 pg were de¿ned as having large and very large genomes, respectively. A preliminary study estimated the coconut genome size at 2.6 Gb using the HiSeq technique (Alsaihati et al., 2014).

C-values often decrease during evolution, but more research is required to identify the molecular events that are involved (Bennett et al., 2000). Arecaceae have chromosome numbers ranging from 2n = 26 to 2n = 36 (Swaminathan and Nambiar, 1961; Sisunandar and Adkins, 2007). In palms, karyotype evolution has involved a reduction in the number of chromosomes from the ancestral value (2n = 36) via disploidy or aneuploidy (Röser, 1994), and this is evident in coconut varieties, which are now diploid (2n = 2x = 32).

Although the Tall and Dwarf coconut types differed in terms of their height and mode of reproduction, there were very small differences in their genome sizes (5.59 pg for Tall types and 5.55 pg for Dwarf types on average). Therefore, these results do not support the theory that Dwarf types evolved from Tall types, because the genome size is a quantitative species characteristic that is relatively consistent among individuals within a population (Greilhuber, 1998). This is the ¿rst report regarding the genome sizes of Tall and Dwarf coconut accessions, and this information may be useful for research on breeding, germplasm characterization, and genome sequencing.

ConÀicts of interest

The authors declare no conÀict of interest.

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