Maturation curves of sweet sorghum genotypes
Curvas de maturação de genótipos de sorgo sacarino

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ABSTRACT
Sweet sorghum \(Sorghum bicolor\) stands out as a complementary crop to sugarcane \(Saccharum\) spp. for the production of ethanol, since it has juicy stems with directly fermentable sugars. Due to this fact, there is a need for the analysis of sweet sorghum properties in order to meet the agro-industry demand. This work aimed to develop and study the maturation curves of seven sweet sorghum cultivars in ten harvest dates. The results showed a significant difference between cultivars and harvest dates for all parameters analysed (p≤0.01). Regarding the sugar content, the cultivars BRS508, XBWS80147 and CMSX629 showed the highest means for the total reducing sugars (TRS) and recoverable sugar (RS). In the production of ethanol per tonne of biomass (EP), the cultivars BRS508 and CMSXS629 presented the best results.

Index terms: Ethanol; reducing sugars; period of industrial use.

INTRODUCTION
The growing demand for energy and the intensive exploration of fossil fuels reserves bring us to a reflection about the need for diversification of the worldwide energy matrix. According to Tomaz and Assis (2013), uncertainty about the availability of non-renewable resources in the future and geopolitical tension in oil producing regions is creating a growing interest in biofuels, due to the fact that these fuels can supply part of the demand for energy in a long term period. Furthermore, as a result of public awareness and knowledge about climate change, there is a need for finding energy sources that do not cause negative impacts on the environment and have lower production cost (Zegada-Lizarazu; Monti, 2012).

Sugarcane \(Saccharum\) spp. is a crop well developed for the production of biofuel. According to a survey by the National Supply Company of Brazil-Conab, in the 2014/2015 season, the cultivated area corresponded to approximately 9.098 million hectares. The total production of sugarcane was estimated in 659.1 million tonnes with an average yield of 74.769 t. ha\(^{-1}\) (Conab, 2014). Despite the fact that sugarcane exhibit high yields, the crop has an offseason period that causes inactivity in the ethanol plants. Additionally, the availability of water tends to become a major constraint to agricultural production in the next years (Ryan; Spencer, 2001), thereby the cultivation of sugarcane may become difficult in some regions, since this crop has a high water requirement (Dayakar et al., 2004). In this scenario, sweet sorghum
Sorghum bicolor (L.) Moench would become a logical option to complement the production system.

Sweet sorghum is characterized by high photosynthetic efficiency and energy production capacity similar to sugarcane with satisfactory production of fresh mass and juicy stems with high levels of fermentable sugars (May et al., 2012a). Due to this production of sugars, sweet sorghum is considered an energy crop of particular importance for the production of ethanol (Gnanoumou; Dauriat; Wyman, 2005; Worley; Vaughan; Cundiff, 1992).

Compared to other energy crops, sorghum shows a wider adaptability to different environments and soil conditions. When compared to maize, sorghum is more tolerant to drought and it is more efficient in water and nutrients usage. In comparison to sugarcane, it has a shorter growth cycle and it is propagated by seeds which facilitates planting operations (Guiying, 2000; Prasad, 2007).

One of the obstacles to this crop is the fact that sugarcane has established dominance over the production chain of sugar and ethanol, receiving the majority of the investments. However, sweet sorghum is rapidly expanding and has a great potential for further growth. In countries like China, India, United States, Brazil, Iran, Italy and Spain, this crop is considered a promising feedstock for ethanol production, and many research projects have been developed with it in these countries (Almodares; Hadi; Ahmadpour, 2008; Channappagoudar et al., 2007; Wortmann et al., 2010).

Even though this plant has a great adaptability, we must remember that in order to exploit the full potential of the crop, it is necessary to provide suitable conditions for its development. According to May et al. (2012a), contrary to what is long believed, in order to obtain satisfactory yields, sweet sorghum needs development conditions similar to other crops, as the adequate soil preparation, fertilization and control of insects, diseases and weeds.

In Brazil, Embrapa (Brazilian Corporation of Agricultural Research) began a program for the development of sweet sorghum cultivars in the 1970s. Initially, 50 genotypes of the United States Department of Agriculture (USDA), Africa and India, were introduced and had its agronomic traits assessed. However, with the poor success of Proálcool, the national ethanol program, and the national policy directed to large ethanol plants, the focus of research on sweet sorghum was redirected to the production of fodder (Parrella, 2011).

Currently, sweet sorghum is being included in the ethanol industry with the proposal of providing feedstock additionally to sugarcane, with the main objective of anticipating the harvest, being processed in the mills in March and April, reducing the need for sugarcane in a period of low quality of its juice. Therefore, sorghum seeds should be sown preferably in November or December of the previous year, since the cycle of the species is from 120 to 150 days (May et al., 2012b).

Unlike sugarcane, sweet sorghum starts the accumulation of sugars in the stem in its reproductive stage, mainly on the physiological maturity of the grains (Teixeira et al. 1999). Due to this difference, the evaluation of the sugar accumulation profile in the stem is an important factor in determining the potential use of each cultivar. This assessment can be done through the study of the period of industrial use (PIU) required for the harvest planning and processing of the material, which should be the longest possible, with a minimum threshold of 30 days.

The PIU comprehends the period in which the cultivar can stay on the field maintaining productivity and quality at optimal levels, according to the minimum standards established to ensure the viability of the crop until it is harvested and processed by the ethanol plant. In this context, this study aimed to establish the maturation curves of seven cultivars of sweet sorghum, by analyzing the sugar accumulation profile in the stem over time, in order to determine the PIU and the ethanol yield of each genotype throughout the evaluation period.

**MATERIAL AND METHODS**

During the season of 2013-2014, comparative trials of sweet sorghum cultivars were conducted in an experimental area of Embrapa Miho e Sorgo in Sete Lagoas – Minas Gerais, 19° 28’ S, 44° 15’08’’ W and altitude 732 m. The climate in the region is Aw (Köppen), typical of savannah, with dry winter and average temperature of the coldest month above 18 °C. The soil of the area is classified as ferralsols whose characteristics are shown in Table 1. In the seeding operation, 400 kg ha\(^{-1}\) of 8-28-16 (NPK) were applied, and 200 kg ha\(^{-1}\) of urea were used in topdressing 20 days after sowing. After 15 days of emergence, roughing was performed in order to keep eight plants per linear meter, totaling 40 plants per rows of five meters.

The weed control was performed with Atrazine, at a dose of 1.5 kg of active ingredient per hectare and it was complemented by manual weeding. Supplementary irrigation was applied during crop establishment to avoid water deficit. The genotypes used in this work were BRS508 and BRS511 (commercial varieties of Embrapa), XBWS80147 and XBWS80007 (Commercial Hybrids of Monsanto), two experimental genotypes of Embrapa (CMSXS647, CMSXS629) and the Sugargraze (commercial Hybrid of Advanta).
The seven genotypes were evaluated in an experiment designed in randomized block with three replications and the sowing performed on 15 of October 2013. The experimental plot consisted of three rows of 5 m long with 0.70 m of spacing with a population of 114,000 plants ha⁻¹. Only the central row was considered to perform the evaluation. The plots were harvested sequentially starting from the flowering stage, 93 days after sowing, considering that the genotypes had similar flowering patterns. In total, ten harvests were performed with an interval of seven days, following the schedule shown in Table 2. During the harvests, the evaluations were focused on the fresh mass production (FMP) in t ha⁻¹, the percentage of total reducing sugars (TRS) in % of juice, the recoverable sugars (RS) in kg of sugar per tonne of biomass and the ethanol production in liters per tonne of biomass (EP).

The fresh mass production was calculated in the plot and transformed into tonnes per hectare. The plants were harvested at 5.0 cm from the soil surface. After removing the panicles, the stalks with leaves were weighed using a digital scale. Samples of eight stalks without leaves were collected in each plot and then fragmented and homogenized in appropriate equipment. Subsamples of 500 grams of the material were pressed using a hydraulic press (245 kg cm⁻² for 60 seconds) for the juice extraction. The juice extracted from the stalks was analysed in the saccharimeter for determination of sucrose and Redutec® for determination of reducing sugars. For the calculation of the variables analyzed in this work, the following formulas adapted from Conselho dos Produtores de Cana-De-Açúcar e Álcool do Estado de São Paulo-Consecana (2006), were used:

- Total reducing sugars extracted from the juice (TRS) – (% of reducing sugars in the juice):

\[
\text{TRS} = \% \text{ reducing sugars} + \frac{\text{Pol}}{0.95}
\]

*Pol* = Apparent content of sucrose

- Recoverable sugar (RS) – (kg of recoverable sugar per tonne of biomass):

\[
10 \times \text{PC} = \text{Pol per tonne of biomass}
\]

\[
1.05263 = \text{stoichiometric coefficient of conversion of sucrose into reducing sugars}
\]

**Table 1**: Soil analysis of the experimental area – EMBRAPA Milho e Sorgo CNPMS – Sete Lagoas/MG.

<table>
<thead>
<tr>
<th>Sample</th>
<th>pH</th>
<th>M. O. (dag/kg)</th>
<th>P (mg/dm³)</th>
<th>K (mg/dm³)</th>
<th>Ca (mg/dm³)</th>
<th>Mg (mg/dm³)</th>
<th>Al (mg/dm³)</th>
<th>H+Al (cmolc/dm³)</th>
<th>SB (cmolc/dm³)</th>
<th>CTC (%cmolc/dm³)</th>
<th>C (cmolc/dm³)</th>
<th>V (%)</th>
<th>Sat. Al (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>00-20cm</td>
<td>6</td>
<td>5.3</td>
<td>15.0</td>
<td>246.7</td>
<td>9.1</td>
<td>0.8</td>
<td>0.04</td>
<td>7.4</td>
<td>10.6</td>
<td>17.9</td>
<td>3.0</td>
<td>59</td>
<td>0.38</td>
</tr>
<tr>
<td>20-40cm</td>
<td>6.1</td>
<td>4.6</td>
<td>14.2</td>
<td>206.9</td>
<td>8.9</td>
<td>0.8</td>
<td>0.05</td>
<td>7.4</td>
<td>10.3</td>
<td>17.7</td>
<td>2.7</td>
<td>58</td>
<td>0.48</td>
</tr>
</tbody>
</table>

PH in distilled water; Extractor of P, K and micronutrients, Mehlich-1; M.O - Organic Matter; Cation-exchange capacity - S+H+Al; V – Percent Base Saturation – V=100 S/T.

**Table 2**: Harvest schedule of the experimental plots.

<table>
<thead>
<tr>
<th>Operation</th>
<th>Date</th>
<th>Days of Plant Development</th>
</tr>
</thead>
<tbody>
<tr>
<td>Seeding</td>
<td>October 15, 2013</td>
<td>0</td>
</tr>
<tr>
<td>1st Harvest</td>
<td>January 16, 2014</td>
<td>93</td>
</tr>
<tr>
<td>2nd Harvest</td>
<td>January 23, 2014</td>
<td>100</td>
</tr>
<tr>
<td>3rd Harvest</td>
<td>January 30, 2014</td>
<td>107</td>
</tr>
<tr>
<td>4th Harvest</td>
<td>February 06, 2014</td>
<td>114</td>
</tr>
<tr>
<td>5th Harvest</td>
<td>February 13, 2014</td>
<td>121</td>
</tr>
<tr>
<td>6th Harvest</td>
<td>February 20, 2014</td>
<td>128</td>
</tr>
<tr>
<td>7th Harvest</td>
<td>February 27, 2014</td>
<td>135</td>
</tr>
<tr>
<td>8th Harvest</td>
<td>March 06, 2014</td>
<td>142</td>
</tr>
<tr>
<td>9th Harvest</td>
<td>March 13, 2014</td>
<td>149</td>
</tr>
<tr>
<td>10th Harvest</td>
<td>March 20, 2014</td>
<td>156</td>
</tr>
</tbody>
</table>
0.905 = coefficient of recovery for an industrial loss of 9.5% 
10 x RST = reducing sugars per tonne of biomass

- Hydrated Ethanol Production Estimation (EP) – 
  (liters of ethanol per tonne of biomass): 

  \[ EP = TRS \times 10 \times 0.6475^* \]

  \* Considering a fermentation process of 85% efficiency.

The collected data were subjected to a combined analysis of variance in factorial design, with seven genotypes x ten harvests, using the software Genes® (Cruz, 2006). In addition, regression curves were generated for each genotype to evaluate the sequence of physical and chemical changes that occurred during the maturation, as well to monitor the formation of sugars in the stalks of the plant in order to find the best moment for harvesting. To build the curves, it was used the fresh mass production (FMP), total reducing sugars (TRS), recoverable sugars (RS) and an estimated production of ethanol in liters per tonne of biomass (EP).

The parameters used for the determination of the period of industrial use (PIU) used in this work were the minimum levels of sugars, corresponding to 12.5% for total reducing sugars (TRS) and 80 kg t⁻¹ for recoverable sugars (RS) (May et al., 2012a; Schaffert, 2011).

**RESULTS AND DISCUSSION**

**Statistical analysis**

The analysis of variance for fresh mass production (FMP), total reducing sugars (TRS), recoverable sugars (RS) and ethanol production (EP) is shown in Table 3. A significant difference was identified for cultivars and harvest day for all characteristics (p≤0.01). This fact demonstrates the genetic differences between cultivars and the influence of the harvest time over the potential yields, indicating the different physiological development profile of the plants during the evaluations. On the interaction between cultivars and harvest day, no significant differences were observed for FMP, which shows that changes in the performance ranking or in the magnitude of the differences between of cultivars did not occur during the evaluations of this characteristic.

On the other hand, in this same source of variation, there was a significant difference at 5% probability for the TRS, RS and EP. This fact shows that the cultivars had different responses to the interaction between their genotype and the moment of the harvest. It further indicates that the cultivars have distinct profiles of formation and concentration of non-structural carbohydrates in the stalk. The coefficient of variation of the data, for all characteristics (FMP - 13.58%, TRS - 9.79%, RS – 9.18% and EP - 9.79%) were satisfactorily low, showing that the experiment had acceptable errors for the study of these characteristics.

**Fresh mass production (FMP)**

The fresh mass production is a key factor for the production of ethanol and there is a strong association between fresh mass production and the ethanol yield per hectare because the sugar-rich juice is extracted entirely from this biomass (Murray et al., 2008; Ritter et al., 2008). Therefore, the higher the production of fresh mass, the greater will be the quantity of juice and sugar per hectare and consequently a higher volume of ethanol.

<table>
<thead>
<tr>
<th>Source of Variation</th>
<th>DF</th>
<th>FMP</th>
<th>TRS</th>
<th>RS</th>
<th>EP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Block/Harv. day</td>
<td>20</td>
<td>164.46</td>
<td>1.56</td>
<td>106.52</td>
<td>47.27</td>
</tr>
<tr>
<td>Cultivars (C)</td>
<td>6</td>
<td>2627.98**</td>
<td>51.14**</td>
<td>2510.17**</td>
<td>1549.23**</td>
</tr>
<tr>
<td>Harv. day (H)</td>
<td>9</td>
<td>1581.67**</td>
<td>92.32**</td>
<td>4744.89**</td>
<td>2796.22**</td>
</tr>
<tr>
<td>C x H</td>
<td>54</td>
<td>90.92 NS</td>
<td>1.64*</td>
<td>90.39*</td>
<td>49.84*</td>
</tr>
<tr>
<td>Residual</td>
<td>120</td>
<td>92.72</td>
<td>1.05</td>
<td>55.51</td>
<td>31.88</td>
</tr>
<tr>
<td>Mean</td>
<td>70.91</td>
<td>10.47</td>
<td>81.16</td>
<td>57.63</td>
<td></td>
</tr>
<tr>
<td>CV(%)</td>
<td>13.58</td>
<td>9.79</td>
<td>9.18</td>
<td>9.79</td>
<td></td>
</tr>
</tbody>
</table>

*, ** significant on F-test at 5% and 1% probability, respectively.
The regression curves corresponding to the fresh mass production (FMP) of the genotypes (Figure 1) show initial values between about 70 and 100 t ha\(^{-1}\), with emphasis on CMSXS647, XBWS80147 and XBWS80007 with total average yield of 88.74 t ha\(^{-1}\), 75.18 t ha\(^{-1}\) and 70.51 t ha\(^{-1}\), respectively. The other genotypes showed an average yield between 60 t ha\(^{-1}\) and 70 t ha\(^{-1}\). These results show the potential for biomass production of sweet sorghum compared to sugarcane which has an average yield of 74.769 t ha\(^{-1}\) (Conab, 2014).

In a study conducted by Albuquerque et al. (2012) where two cultivars (BRS506 and BRS507) were compared in different row spacing and densities in the state of Minas Gerais in Brazil, it was possible to reach yields between 36.44 t ha\(^{-1}\) and 45.75 t ha\(^{-1}\) of fresh mass, values lower than those indicated in the present work. In contrast, May et al. (2012b) also evaluating the potential of sweet sorghum in different settings for the production of ethanol, reached higher values, with yields of 80.61 t ha\(^{-1}\) in the central region of Minas Gerais.

In the analysis of the curves in Figure 1, it is possible to note a remarkable performance of the experimental line CMSXS647, which had the highest production in all harvests. Despite the fact that this genotype had the lowest coefficient of determination, it is clear that this material has a great potential for further use in sweet sorghum breeding programs.

The high yield of fresh biomass is an important indicator of the potential of sweet sorghum, but it alone does not provide the necessary requirements for the qualification of cultivars when the objective is the ethanol production. Therefore, it is necessary to evaluate the sugar accumulation curves during the plant development.

**Total reducing sugars (TRS)**

The reducing sugars are monosaccharides able to be oxidized in the presence of oxidizing agents in alkaline solutions, since they have free carbonyl and ketone group (Silva, 2003). In this study, the reducing sugars considered were glucose and fructose.

The total reducing sugars are the result of the sum of reducing sugars themselves, contained in the sorghum juice, and the glucose and fructose originated from the hydrolysis of sucrose. They are represented by all sugars in the material in the form of inverted sugar and make up all the sugar (sucrose + glucose + fructose) in the juice, expressed in percentage (Fernandes, 2011). The reducing sugars represent a quality parameter of the feedstock widely used by the sugar and ethanol industry. Usually the cultivars that have high content of sugars and low fibers are more susceptible to physical damage and attack of pests and microorganisms.

**Figure 1:** Curves of Fresh Mass Production (FMP) in t ha\(^{-1}\) of seven cultivars of sweet sorghum in ten harvest dates.
The Figure 2 shows the performance of the cultivars regarding the accumulation of total reducing sugars (TRS). The cultivar BRS508 had the highest means for total reducing sugars in all harvests, with values higher than the established minimum starting from 28 days after flowering and keeping the high content until the end of the evaluations, totaling 35 days.

The cultivar XBSW80147 reached a sugar level above the established minimum after 42 days and maintained it for 21 days. The cultivar CMSXS629 also reached the minimum value of TRS after 42 days, keeping it for 14 days and decreasing to a value lower than the established minimum in the last evaluation. The other cultivars did not reach the minimum threshold for TRS. This fact may be related to the occurrence of sugarcane borer *Diatrea saccharalis* in some of the plots and the consequent red rot in the stalk caused by the fungus *Colletotrichum falcatum* and *Fusarium moniliforme*.

**Recoverable sugars (RS)**

The recoverable sugars correspond to the amount of total sugars extracted from the juice (Fernandes, 2011; Consecana, 2006). Currently, the payment to growers is based in kilograms of total recoverable sugars (RS), as established by the Conselho dos Produtores de Cana-de-Açúcar e Álcool do Estado de São Paulo-Consecana. Among the advantages of this payment method, there is the quality of the feedstock because the grower seeking better earnings, deliver a better quality material, not only focusing on yield per hectare (Consecana, 2006).

As can be seen in Figure 3, the maturation curves of recoverable sugars resemble the TRS. Analysing these curves, it is possible to evaluate the period of industrial use (PIU), which comprises the cultivating period for which the RS is at or above 80 kg t⁻¹. It is recommended a PIU of at least 30 days to ensure a good planning and processing of the feedstock. The RS in the cultivars BRS508 and CMSXS629 were above the established minimum starting from the third evaluation (14 DAF), totaling approximately 50 days of PIU, with and average value of 98.87 kg t⁻¹ and 86.11 kg t⁻¹ and the highest value of 122.78 kg t⁻¹ and 104.49 kg t⁻¹ at 42 DAF, respectively. The cultivars XBSW80147 and XBWS80007 reached the minimum threshold at 21 and 28 days after flowering, totaling approximately 42 and 35 days of PIU, with average production of 80.38 kg t⁻¹ and 75.89 kg t⁻¹ and highest of 99.07 kg t⁻¹ and 93.70 kg t⁻¹ at 42 DAF, respectively.

The cultivars BRS511 and Sugargraze reached the minimum value after 35 days, maintaining a PIU of 28 days.

![Figure 2](image.png)  
*Figure 2*: Curves of total reducing sugars (TRS), in % of extracted juice of seven cultivars of sweet sorghum in ten harvest dates.
and an average yield of 77.60 kg t\(^{-1}\) and 73.96 kg t\(^{-1}\) and highest value of 94.71 kg t\(^{-1}\) and 86.00 kg t\(^{-1}\), respectively. The cultivars CMSXS647 reached the minimum 56 days after flowering, thereby having the shortest PIU (7 days) and the lowest average (72.28 kg t\(^{-1}\)). These results show a longer PIU than those found in a study conducted by Fernandes et al. (2014) where the BRS 508 maintained sugar concentration greater than 80 kg t\(^{-1}\) (140 g L\(^{-1}\)) for 42 days and reached a maximum value of 92.57 kg t\(^{-1}\) (162 g L\(^{-1}\)).

In this same study, BRS 511 showed a better performance, with the sugar levels peaking at 109 kg t\(^{-1}\) (191 g L\(^{-1}\)) and a PIU of 44 days. In general, after 42 days of flowering, the sugar concentrations decreased for all genotypes studied.

**Ethanol production (EP)**

The major free sugars in the juice of sweet sorghum, as explained before, are glucose, fructose and sucrose. It is noteworthy that sweet sorghum has a higher amount of reducing sugars compared to sugarcane. However, there is no difference in the total quantities of soluble sugars and the technological parameters and analytical methods for quality control of sweet sorghum may be the same as the ones used for the sugarcane (Gomes; Rodrigues; Oliveira, 2011).

The levels of sugar found in sweet sorghum are between 15-21\%, values very close to those found in sugarcane. As sweet sorghum has a higher concentration of reducing sugars (glucose and fructose), one should be more careful during the production process of ethanol, because there may be a greater risk of contamination, taking into account that contaminating bacteria in the material prioritize reducing sugars during the fermentation process (Machado, 2011).

In the Figure 4 there are the curves for the production of ethanol in liters per tonne. As in TRS and RS curves, it is possible to see that there is an increase in production over time. According to (May et al., 2012a; Schaffert, 2011) the optimal values for ethanol production for sweet sorghum cultivars in Brazil should be above 60 L t\(^{-1}\).

The genotype CMSXS629 reached 63.24 L t\(^{-1}\) at 28 DAF with a peak production of 75.5 L t\(^{-1}\) at 42 DAF maintaining a satisfactory production for 35 days. Other important results were the XBWS80147 and XBWS80007 that maintained a satisfactory production for a period of 28 days, reaching 66.23 L t\(^{-1}\), 60.67 L t\(^{-1}\) at 35 DAF, respectively. The cultivars BRS511 reached the minimum threshold at 42 DAF with 65.29 L t\(^{-1}\), maintaining the production for 21 days. The sugargraze reached 61.9 L t\(^{-1}\) at

**Figure 3:** Curves of recoverable sugars (RS), in kg t\(^{-1}\) of seven cultivars of sweet sorghum in ten harvest dates.
49 DAF and maintained an average yield near this value until the end of the evaluations, totaling 14 days. The cultivar CMSXS647 reached the minimum limit only at 56 DAF, with 69.97 L t\(^{-1}\).

These results show the potential of sweet sorghum for the production of ethanol and are higher than the amounts reported by Emygdio et al. (2011) studying the cultivar BR506, where it reached an average of 55 liters of ethanol per tonne of stalks. Similarly, Zhang et al. (2010) assessing the ethanol potential of sweet sorghum in China by ArcGIS methods obtained an average ethanol yield of approximately 62.5 L t\(^{-1}\).

The adequate knowledge of the industrialization potential of each genotype is the most important factor in determining the amount and the planting moment of each of them. This planning is necessary in order to schedule the harvest and provide enough feedstock for a longer time for the full operation of the ethanol plant. In the Figure 5, it is presented the period of industrial use (PIU) for all genotypes, based on the 80 kg t\(^{-1}\) threshold for recoverable sugars (RS) established by (May et al., 2012a; Schaffert, 2011). In this figure, it is possible to analyse the profiles of fresh mass production, accumulation of sugars and ethanol production of all cultivars and the relationship between the variables in each case. The adjustment of the data to the regression measured by the coefficient of determination was satisfactory for all genotypes analyzed.

**Figure 4:** Curves of ethanol production (EP), in liters per tonne of seven cultivars of sweet sorghum in ten harvest dates.
Figure 5: Maturation Curves of seven cultivars of sweet sorghum in ten harvest dates. PIU corresponding to RS above 80 kg t⁻¹. FMP (kilograms per tonne of biomass), TRS - Total reducing sugars (%), RS - Total recoverable sugars (kilograms per tonne of biomass) and EP - ethanol production (liters per tonne of biomass).
CONCLUSIONS

Analysing the maturation curves, it was found that there was an increase in the concentrations of total sugar in the juice and consequently a higher ethanol yield throughout the evaluations. It is worth mentioning that the cultivar BRS508 and the CMSXS629 showed a satisfactory performance and productivity with values similar to those obtained for sugarcane under the same operating conditions. These facts demonstrate the high potential of using sweet sorghum for the production of ethanol, and the capacity of these cultivars to sustain good yields for a long period of time may provide greater certainty in the harvest planning and usage of the feedstock in existing industrial facilities.

Despite the fact that the curves were variable for each genotype, it was possible to observe a trend in the analysis of the FMP, TRS, RS and EP, where the time of harvesting with the highest yield is found between 21 and 42 days after flowering.

ACKNOWLEDGEMENTS

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