

Phosphorylated and Cross-Linked Wheat Starches in the Presence of Polyethylene Oxide and Their Application in Biocomposite Films

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Wheat starch is modified by phosphorylation and cross-linking. Starches are evaluated for phosphorus content, amylose content, swelling power, solubility, pasting properties, crystallinity, and morphology. Films are made with phosphorylated and cross-linked starches, and with addition of 10% (w/w) polyethylene oxide (PEO). The films are evaluated for morphology, water vapor permeability, mechanical properties, crystallinity, roughness, and angle of contact with water. Phosphorylated starch shows higher phosphorus content, amylose content, swelling power, and solubility than cross-linked starch and native starch did. The starch film containing PEO presents discontinuous morphology, is more crystalline and rugged, and less hydrophilic than starch films without PEO. The biocomposite proves promising for usage in flexible packaging.

of starch with polyethylene oxide (PEO) to produce biocomposite films is interesting because PEO is biodegradable, biocompatible, and forms hydrogen bonds with oxygen, which allows intermolecular interactions with the components of the polymer matrix. Furthermore, the presence of PEO improves the poor mechanical properties of films produced with starch and provides a semi-crystalline structure to the final material, which is an attractive property of films.^[3] Furthermore, the interaction between starch and PEO is influenced by the origin and proportion of starch and by the PEO semi-crystalline structure.^[3]

Modified starches, such as phosphorylated and cross-linked starches, have been suggested for use in films due to their excellent mechanical and water barrier properties compared to native starch films. Phosphorylation is a chemical modification in which phosphate groups are introduced into starch. The negatively charged phosphate groups cause repulsion among the starch chains and, consequently, increase their hydration. Thus, phosphorylation decreases the starch gelatinization temperature and increases retrogradation. Other changes in starch properties include increase in swelling power, granule solubilization, paste clarity, and starch gel viscosity.^[4] These properties are advantageous in the development of biodegradable films, as well as in improving film characteristics.

Cross-linking is a chemical modification obtained using reagents such as trimetaphosphate, sodium tripolyphosphate, epichlorohydrin, phosphoryl chloride, and a mixture of adipic acid, acetic anhydride, and vinyl chloride, all capable of forming intra- and intermolecular bonds between the hydroxyl groups of starch.^[5] Biodegradable starch films prepared from cross-linked starches provide improved mechanical properties, abrasion/cut through, resistance to stress cracking, high-temperature mechanical properties, overload characteristics, and decreased inflexibility.^[6] However, the degree of improvement depends on the type and content of cross-linking agent. Besides, the structure and molecular weight of the starch also affects the intermolecular interactions between molecules.

1. Introduction

Wheat starch can be used to produce biodegradable films because of its low cost, and renewable and biodegradable properties. However, use of natural starch films is limited due to their poor mechanical properties and low water barrier. To improve these properties, the association of natural and synthetic polymers for biocomposite formation has been studied. Previous studies showed that composite films made from starch and polyvinyl alcohol (PVA)^[1] and polyvinyl chloride (PVC)^[2] exhibit good processing characteristics, and improved mechanical and water barrier properties. The mixture

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DOI: 10.1002/star.201700192

Jagadish and Raj^[7] prepared films from blends of starch and PEO, and reported that these films presented interesting characteristics for application in food packaging. These authors observed that films with higher PEO/starch ratios presented higher tensile strength and lower water-vapor transmission rate. However, the behavior of films made with modified starch and PEO was not studied. The use of starches modified by phosphorylation or cross-linking may facilitate the interaction between starch and PEO. Therefore, the objective of this study was to develop and characterize biocomposite films using phosphorylated and cross-linked wheat starches and PEO in order to obtain materials with improved properties.

2. Experimental Section

2.1. Materials

Wheat (*Triticum aestivum*) grains of BRS Parrudo cultivar were provided by Embrapa-Trigo (State of Rio Grande do Sul, Brazil). All the chemical reagents used were of analytical grade.

2.2. Isolation of Starch

Wheat starch was extracted according to the procedure suggested by Knight and Olson.^[8] Briefly, a mixture of wheat flour and water was prepared in a ratio of 2:1 (w/v). After the formed mass was washed with water until complete removal of the starch, the starch-water mixture was filtered through a 200-mesh sieve. The filtrate was centrifuged at 3500 rpm for 20 min. After centrifugation, the dark layer on top of the precipitated material was removed and the remainder of the decantate was resuspended in distilled water and centrifuged again. The process was repeated three times. The starch was dried at 40 °C for 16 h to approximately 9% moisture content and stored at 17 ± 2 °C. The wheat starch presented approximately 89.5% purity (0.48% protein, 0.50% lipids, and 0.18% ash). The extraction rate of the wheat starch was approximately 50%.

2.3. Preparation of Phosphorylated Starch

Phosphorylation was performed according to the method described by Paschall.^[9] Briefly, 167 mL of 7.54% (w/v) sodium tripolyphosphate in water was adjusted to pH 5.5 using 1 mol L⁻¹ HCl solution. This solution was mixed with 100 g (dry basis) of starch and stirred in a magnetic stirrer for 20 min. Then, the mixture was filtered using a vacuum pump (SOLAB, SL-61, Brazil), and the sediment was dried in a forced-air oven for 48 h at 45 °C. The dried sample was ground in an analytical mill and transferred to an oven with forced-air circulation at 65 °C for 90 min. Then, the solid was transferred to a stationary oven at 155 °C for 40 min. After cooling, 300 mL of 50% ethanol was added, and the samples centrifuged at 2200 rpm for 5 min. The supernatant was discarded and the resultant material was dried in a forced-air oven at 45 °C for 24 h. After phosphorylation, the samples were dialyzed to remove unbound phosphorous salts. A suspension of 10% (w/v) phosphorylated starch was

placed into a 30-cm long cellulose dialysis membrane (12 000–16 000 molecular weight cut-off, 25 Å porosity) and immersed in distilled water (changed twice a day) under cooling over a period of 3 days. After dialysis, the starch was dried in a forced-air oven at 45 °C for 24 h. The degree of substitution (DS) was calculated according to Eq. (1).

$$DS = \frac{163 P}{(3100 - 124P)} \quad (1)$$

where P is the phosphorus content.

2.4. Preparation of Cross-Linked Starch

Starch crosslinking was carried out according to the method suggested by Woo and Seib.^[10] Briefly, a mixture of 100 g of starch, 2 g of sodium trimetaphosphate, and 2 g sodium sulphate in 200 mL of distilled water was adjusted to pH 11 using 0.5 mol L⁻¹ NaOH and, then, heated at 35 °C while stirring (IKA, RW20, Germany) for 16 h. After this period, the mixture was washed with excess distilled water and filtered using a vacuum pump (SOLAB, SL-61, Brazil) before further drying at 40 °C in an oven with air circulation for 24 h. After modification, the samples were dialyzed to remove unbound phosphate salts. Precisely, a suspension of 10% (w/v) cross-linked starch was placed into a 30-cm long cellulose dialysis membrane and immersed in distilled water (changed twice a day) under cooling over a period of 3 days. After dialysis, the starch was dried in a forced-air oven at 45 °C for 24 h. The DS was calculated according to Eq. (2).

$$DS = \frac{324 P}{(3100 - 96P)} \quad (2)$$

Where P is the phosphorus content.

2.5. Characterization of the Modified Starches

The phosphorus content of the starches was determined according to the method suggested by Smith and Caruso.^[11] Briefly, 10 mL of 10% zinc acetate was uniformly added to 5 g of starch samples and completely evaporated on a heating plate. The samples were placed in a muffle at 550 °C for 2 h. Then, 0.75 mL of 29% nitric acid was added and evaporated, and the sample was placed in a muffle at 550 °C for 30 min. After the sides of the crucible used to dry the samples were washed with 2.5 mL of 29% nitric acid and 3.75 mL of water, the crucible was heated on a water bath for 10 min. The content of the crucible was filtered using four 2.5-mL portions of distilled water. Absorbance at 460 nm was read using a spectrophotometer after 10 min. A standard curve was created using 0.439 g of anhydrous monobasic potassium phosphate dissolved 1 L of distilled water.

Apparent amylose content in starches was determined using the colorimetric method based on iodine as described by McGrane, Cornell, and Rix,^[12] in which a standard amylose curve was created using pure potato amylose (Sigma-Aldrich).

The swelling power and solubility of the starches at 90 °C were determined following the method described by Leach,

McCowen, and Schoch.^[13] Briefly, 1.0 g of starch was mixed with 50 mL of distilled water in centrifuge tubes. The various suspensions were heated at 90 °C for 30 min. The solubility was expressed as the percentage of undissolved dried solid weight over the initial weight of the dry sample. The swelling power was calculated as the ratio of the weight of the wet sediment to the initial weight of the sample (deducting the amount of soluble starch).

The pasting properties of the starch were determined using a Rapid Visco Analyzer (RVA-4, Newport Scientific, Australia) and the Standard Analysis 1 profile. Viscosity was expressed in rapid visco units (RVU). Starch (3.0 g of 14 g/100 g wet basis) was weighed directly in a RVA canister and added 25 mL of distilled water. Parameters comprising pasting temperature, peak viscosity, breakdown, final viscosity, and setback were recorded.

X-ray diffractograms of the starches were obtained using an X-ray diffractometer (XRD-6000, Shimadzu, Brazil). The scanning region of the diffraction ranged between 5° and 45°, with a target voltage of 30 kV, a current of 30 mA, and a scan speed of 1° min⁻¹. The relative crystallinity (RC) of the starch granules was calculated as described by Rabek^[14] according to Eq. (3).

$$RC = \frac{Ac}{(Ac + Aa)} \times 100 \quad (3)$$

where Ac is the crystalline area and Aa is the amorphous area on the X-ray diffractograms.

2.6. Elaboration of Biocomposite Films

Biocomposite films were prepared using a casting technique. Preliminary tests were performed to define the PEO concentration in the films. For film preparation, a solution was prepared with 3% starch in 100 g of distilled water, 0.30 g glycerol/g dry starch, and 0 or 10 g of PEO/100 g dry starch. PEO control film was prepared using 3% PEO in 100 g of distilled water. The PEO was suspended in water with subsequent stirring in an Ultraturrax homogenizer (IKA, T18B, Werke, Germany) at 11,000 rpm for 5 min; then, starch and glycerol were added. The solution was heated at 90 °C for 30 min. Then, 20 g of each film solution was spread on 9-cm diameter acrylic plates and dried in an oven with air circulation at 35 °C for 16 h. The film samples were stored in a hermetic container at 16 °C and approximately 65% relative humidity (RH) using a saturated solution of ammonium nitrate (NH_4NO_3) for 7 days.

2.7. Characterization of Biocomposite Films

2.7.1. Morphology

The surface and cross-section morphology of the composites were visualized using a scanning electron microscope (JEOL, JSM-6610LV, New Jersey, USA) with accelerating voltage of 10 kV. For cross-sections, the samples were fractured under liquid nitrogen prior to visualization. Samples were then placed in a stub, coated with gold using a sputter Desk V (JEOL, New Jersey, USA), and examined at 300× magnification.

2.7.2. Water Vapor Permeability (WVP) of the Biocomposite Films

Water vapor permeability tests of the films were performed following the American Society for Testing and Materials (ASTM) E96-95 standard method.^[15] Each sample was placed and sealed onto the circular opening of a permeation cell containing anhydrous calcium chloride (0% RH). Then, the cells were conditioned in desiccators with a saturated sodium chloride solution (75% RH) at 25 °C until the samples reached steady-state conditions. Finally, the cell weight was measured after 48 h.

2.7.3. Thickness and Mechanical Properties

The thickness of films was determined using a micrometer to the nearest 0.001 mm, at 8 random positions around the film. Average deposition rates were used in the calculations. The tensile strength and percentage of elongation at the break-up point of the films were evaluated according to the tensile test based on the ASTM D-882-91 method^[15] using a Texture Analyzer (TA.XT plus, Stable Micro Systems).

2.7.4. X-Ray Diffraction

The film samples were cut into circular pieces, placed in a sample holder, and evaluated using an X-ray diffractometer at a scan range of 5–60° (2θ), 30 kV voltage, and 30 mA current.

2.7.5. Atomic Force Microscopy

The surface topography of the films was obtained using an atomic force microscope (Agilent Technologies 5500 Equipment, Chandler, AZ, USA) equipped with a commercial, non-contact silicon tip PPP-NCL. The cantilever oscillation had a curvature in the nanometre order and resonance frequency of 154 kHz. The topography images were analyzed using the Pico Image Basic software. The quantitative parameter of surface roughness (Rq) was calculated using the topographic data extracted from the micrographs.

2.7.6. Biocomposite Films Contact Angle With Water

The contact angle of the films with water was determined using a goniometer (Labometric, LB-DX) at room temperature. The samples were adhered to a glass slide using a double-sided tape. A drop (3 μL) of water was dripped on the sample surface with a micropipette, and the drop image was captured using a digital camera. The contact angle values were calculated using the ImageJ software using a drop analysis plugin. The values represent the average of five repetition angles.

2.8. Statistical Analysis

All analytical determinations were performed in triplicate and the results were submitted to analysis of variance (ANOVA). The

mean values were compared by Tukey's test or *t*-test with 5% level of significance.

3. Results and Discussion

3.1. Phosphorus Content of Starches

The phosphorus content of native starch derives from phospholipids.^[15] Our phosphorylated starch showed higher phosphorus content than other starches did (Table 1), indicating that phosphate groups were successfully introduced in the starch chain. Stahl et al.,^[16] who studied the phosphorylation of corn and pinhão starches, reported lower phosphorus values (0.25%) than we did (0.57%). However, these authors performed a 7-day long dialysis process, which probably decreased the phosphorous content in their samples, whereas the dialysis carried out in this study was 3-day long.

According to the Food and Drug Administration, starches modified for food use must have a residual phosphorus content of 0.5% for phosphorylated starch and 0.04% for cross-linked starch (Table 1).^[17] Therefore, phosphorylated corn and pinhão starches could not be used in the food industry. However, they could be employed in the paper industry for sealing pores, and in the textile industry for sizing yarns, as well as in other fields such as production of biodegradable films for flexible packaging.

Deetae et al.^[6] evaluated rice starch cross-linked with 1% sodium trimetaphosphate at 120 °C, with reaction times ranging from 0 to 120 min. These authors reported that the phosphorus content of the starches ranged from 0.007 to 0.030% according to increasing reaction time.

Liu et al.^[18] reported that variations in the phosphorus content of starches can be explained by differences in the distribution of the phosphate groups between amylose and amylopectin. In addition, reaction conditions and starch origin also influence the introduction of phosphate groups in the starch molecule. Starch

contains two types of hydroxyl groups, primary (6-OH) and secondary (2-OH and 3-OH). These hydroxyl groups react with multifunctional reagents resulting in cross-linked starches. Higher phosphorus content of starch suggests a greater interaction among the polymer chains, which causes the structure to become increasingly rigid.

3.2. Amylose Content, Swelling Power, and Solubility of Starches

Interestingly, the phosphorylation process increased the amylose content (Table 1), which could be attributed to the depolymerisation of the external amylopectin chains that can form complexes with iodine. The ability of amylose to produce films has been long known and it is attributed to the ability of its linear chains to interact with other molecules through hydrogen bonds to a higher extent than the branched amylopectin chains. Moreover, amylopectin films are frail because of the high degree of entanglement caused by extensive branching and short average chain length.^[19]

Phosphorylation increased the swelling power and solubility of starch (Table 1). Sthal et al.^[16] reported on the increase of the swelling power of corn and pinhão starches phosphorylated with sodium tripolyphosphate. These authors attributed this result to

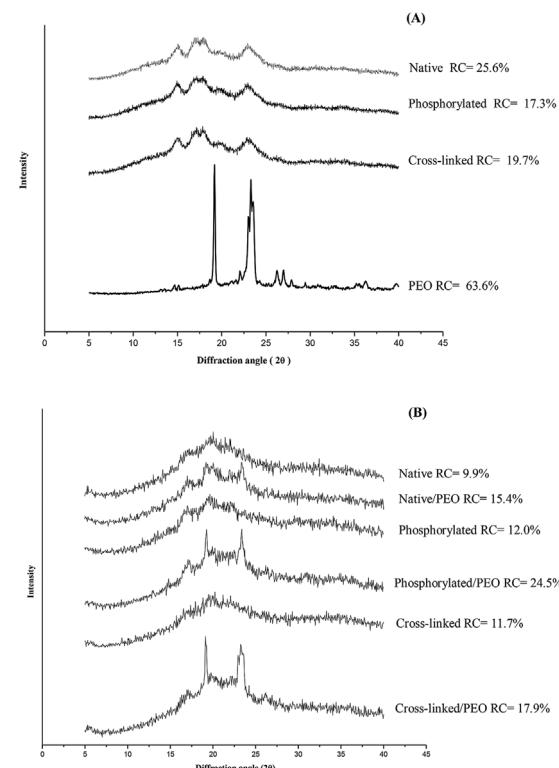


Figure 1. X-ray diffraction patterns and relative crystallinity of native, phosphorylated, and cross-linked wheat starches and polyethylene oxide (PEO) (A), and of films made of native starch, native starch and PEO, phosphorylated starch, phosphorylated starch and PEO, cross-linked starch, and cross-linked starch and PEO (B).

Table 1. Phosphorus content, amylose, swelling power, solubility, and pasting properties of the native and modified starches.

Parameters ^a	Starches		
	Native	Phosphorylated	Cross-linked
Phosphorus content (%)	0.06 ^c	0.57 ^a	0.09 ^b
Degree of substitution	–	0.030 ^a	0.009 ^b
Apparent amylose content (%)	24.75 ^b	27.65 ^a	13.02 ^c
Swelling power (g/g)	12.12 ^b	14.34 ^a	6.66 ^c
Solubility (%)	6.39 ^b	10.10 ^a	2.72 ^c
Pasting temperature (°C)	79.47 ^c	55.63 ^b	88.05 ^a
Peak viscosity (RVU)	235.37 ^a	184.5 ^b	11.41 ^c
Breakdown (RVU)	54.79 ^a	28.86 ^b	3.08 ^c
Final viscosity (RVU)	271.87 ^a	218.25 ^b	12.37 ^c
Setback (RVU)	89.54 ^a	62.08 ^b	4.41 ^c

Values with different letters in the same row are significantly different ($p < 0.05$) by a Tukey test.

the presence of negatively charged phosphate groups in the starch, which reduce the intermolecular binding forces favoring water absorption. However, reduced swelling power of cross-linked starches was observed compared to the native counterparts.

Cross-linking confers the granules a rigid macromolecular structure, which strengthen hydrogen bonds, slows water absorption speed, and increases the resistance of swollen granules to break-up. The increased solubility of phosphorylated starches can be attributed to the introduction of negatively charged phosphate groups in the starch chain, which reduces the strength of bonds in the starch structure.^[20]

As observed in other studies,^[21] modification by cross-linking decreases the solubility of starch in water (Table 1) because cross-links strengthen the granular structure.

3.3. Pasting Properties of Starches

Phosphorylated and cross-linked starches showed reduced peak viscosity, breakdown, final viscosity, and retrogradation (Table 1) compared to native starch. Starches modified by cross-linking showed higher retrogradation than native and phosphorylated starches. Setback, that is, high-viscosity gelled starch, has a great importance in the application of starch paste. During cooling, straight-chain solubilized amylose starch molecules begin to realign and form a stable gel structure. This phenomenon is called retrogradation.^[22] The low amylose content in cross-linked starch compared to other starches may be responsible for their low retrogradation.

Cross-linking and phosphorylation confer high and low pasting temperature, respectively, compared to that of native starch. The decrease in the phosphorylated starch pasting

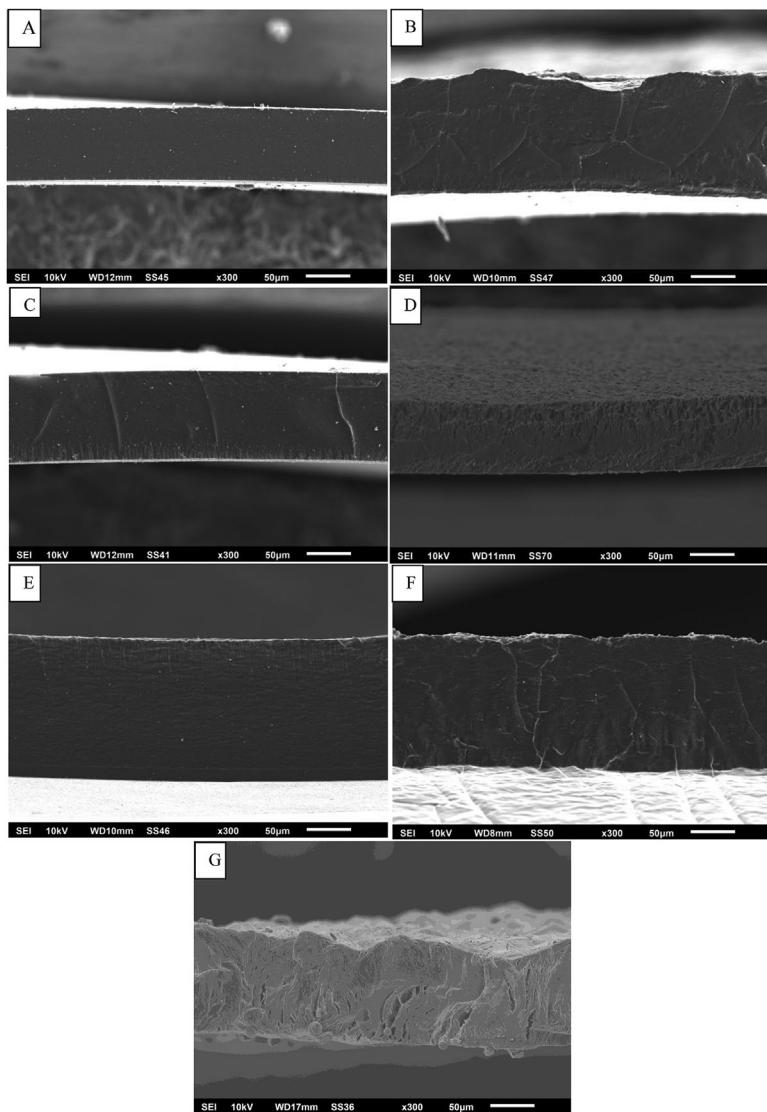


Figure 2. Scanning electron microscopy of the cross section of films made of native starch (A), native starch and polyethylene oxide (PEO) (B), phosphorylated starch (C), phosphorylated starch and PEO (D), cross-linked starch (E), cross-linked starch and PEO (F), and PEO film (G).

temperature is attributed to the incorporation of negatively charged functional groups (i.e., phosphates) in the starch structure, which reduces the attractive forces in the amorphous region, thereby weakening the intramolecular hydrogen bonding bead and forming binders at low temperatures.^[5]

The increased pasting temperature of starch after the introduction of cross-links is likely due to the reduced mobility of the amorphous chains in the starch granules resulting from intermolecular bonds. In fact, cross-links enhance the integrity of the starch granules and retard the breakdown of hydrogen bonds during the gelatinization process, thus leading to low viscosity.

3.4. X-Ray Diffraction of Starches

The X-rays diffraction patterns of native starches showed peaks at 15°, 17°, 18°, and 23° (2θ) (Figure 1a), characteristic of cereal starch type A. The chemically modified starches did not show changes in the diffraction patterns (Figure 1). The phosphorylated and cross-linked starches showed reduced crystallinity compared to the native starch (Figure 1a). Błaszcak et al.^[23] reported that the phosphorylated waxy cornstarch showed noticeable reduction in crystallinity compared to the native starch. These authors suggested that functional groups were monosubstituted by phosphate mainly in the amorphous regions of the waxy maize structure. Kittipongpatana and Kittipongpatana^[24] reported that cross-linking by sodium trimetaphosphate did not alter the pattern of the elephant foot starch X-ray because modifications occurred in the amorphous region of the starch granule.

3.5. Morphology of Films

The micrographs of the cross section of the biocomposite films of native and modified starches, with and without the addition of PEO, are shown in Figure 2. The phosphorylated starch film obtained showed cracks (Figure 2C), whereas cross-linked starch films were homogeneous (Figure 2E).

Table 2. Water vapor permeability, thickness, mechanical properties, and contact angle of films.

Biocomposite ^a						
Starch	PEO (%)	WVP (g.mm/kPa.day.m ²)	Thickness (mm)	Tensile strength (MPa)	Elongation (%)	Contact angle (°)
Native	0	2.4 ± 0.1 ^{b ns}	0.131 ± 0.01 ^a	1.2 ± 0.2 ^b	53.7 ± 1.3 ^b	23.8 ± 0.6 ^c
	10	2.6 ± 0.1 ^c	0.120 ± 0.01 ^b	1.6 ± 0.1 ^c	113.4 ± 0.8 ^b	69.3 ± 0.1 ^a
Phosphorylated	0	3.2 ± 0.5 ^{b ns}	0.105 ± 0.02 ^b	6.5 ± 0.2 ^{a ns}	74.8 ± 4.5 ^{a ns}	25.35 ± 0.6 ^b
	10	2.6 ± 0.1 ^c	0.116 ± 0.02 ^b	6.5 ± 0.2 ^a	77.1 ± 2.8 ^c	57.7 ± 0.1 ^c
Crosslinked	0	5.4 ± 0.4 ^a	0.131 ± 0.010 ^a	1.2 ± 0.1 ^b	43.8 ± 2.9 ^c	27.7 ± 0.2 ^a
	10	4.2 ± 0.1 ^a	0.170 ± 0.02 ^a	2.9 ± 0.3 ^b	146.0 ± 0.7 ^a	66.4 ± 0.3 ^b
Film of PEO ^b		6.0 ± 0.3	0.124 ± 0.010	2.8 ± 0.1	165.1 ± 4.2	40.7 ± 0.7

The results are mean ± standard deviation of three determinations.

Film prepared with only 3 g of PEO.

Values with different letters in the same column represent significant difference by Tukey test ($p \leq 0.05$).

^{ns}Significant and insignificant, respectively, by *t*-test ($p \leq 0.05$), between films with 0 and 10% PEO. PEO: polyethylene oxide.

The addition of PEO in the films caused discontinuous cross sections, regardless of the type of starch (Figure 2). Pereira et al.^[25] found that blends of PEO and cassava starch were irregular, which was attributed to the characteristic spherulitic structure of PEO. The discontinuities of the biocomposite films affect their properties.

3.6. WVP of the Films

The WVP of biocomposite films is shown in Table 2. WVP is an essential parameter for the evaluation of possible biocomposite films applications. The addition of PEO in the cross-linked starch film decreased the WVP (Table 2), which could be related to the high crystallinity of this starch (Figure 1). As previously reported in a study on composite films of chitosan and cellulose, low WVP provides high retention capacity, low water absorption, and adsorption capacity.^[26] Alterations in the starch chains following cross-linking reduced water retention compared to native starches, which contributed to reducing the WVP of the film.

The films made from native and phosphate starches, with and without PEO, showed low WVP values, demonstrating that the presence of PEO does not affect this parameter (Table 2). According to Su and Shen,^[27] since PEO itself has the ability to retain water, it contributes to the retention of water absorbed by biocomposite films made of native and phosphorylated starches, which are weaker than cross-linked starches.

3.7. Thickness and Mechanical Properties of Films

The films made of phosphorylated and cross-linked starches showed increased thickness upon addition of PEO. The phosphorylated starch film without PEO showed higher tensile strength than other films, which was unaltered by the addition of PEO. The higher tensile strength of the phosphorylated starch film could be attributed to its high amylose content compared to that of other starches (Table 1). In fact, most starches with high amylose content confer an elevated tensile strength to the films due to retrogradation.

Addition of PEO increased the tensile strength of the cross-linked starch film, which could be attributed to starch and PEO molecules having a large number of hydroxyl groups in their structures that remain associated by intermolecular hydrogen bonds. Das et al.^[28] studied biocomposite films made of cross-linked starch and PVA. These authors reported that the cross-linking agents used to modify the native starch reacted with the OH groups present in starch and PVA, and formed both linkages contributing to the increased tensile strength of the biocomposites, as well as to reducing water absorption. Moreover, according to Das et al.,^[28] cross-linking between PVA and starch molecules contributed to the high tensile strength of the cross-linked starch films (Table 2).

The control film containing only PEO showed higher elongation than that of starch films containing PEO (Table 2). Mao et al.^[29] reported that the addition of PVA increased the tensile strength and elongation of composite films made of

cornstarch. Moreover, these authors reported that the addition of PVA to the starch prevented the development of surface cracks, indicating a good compatibility of starch and PVA. In fact, both the starch and PVA are polar substances harboring hydroxyl groups (-OH), which form intramolecular and intermolecular hydrogen bonds that improve the integrity of starch-PVA mixtures.

Pereira et al.^[3] reported that the interaction between starch and PEO in various blends depends on their proportion and, especially, on the type of starch. When the amount of starch in the mixture is low, intra-chain interactions form preferentially. When the amount of starch is increased, intermolecular hydrogen bonds form between the hydroxyl groups of the starch and the PEO oxygen atoms. Therefore, PEO proved capable of forming strong interactions with starches, forming biocomposite films with improved mechanical properties compared to the films with no PEO addition.

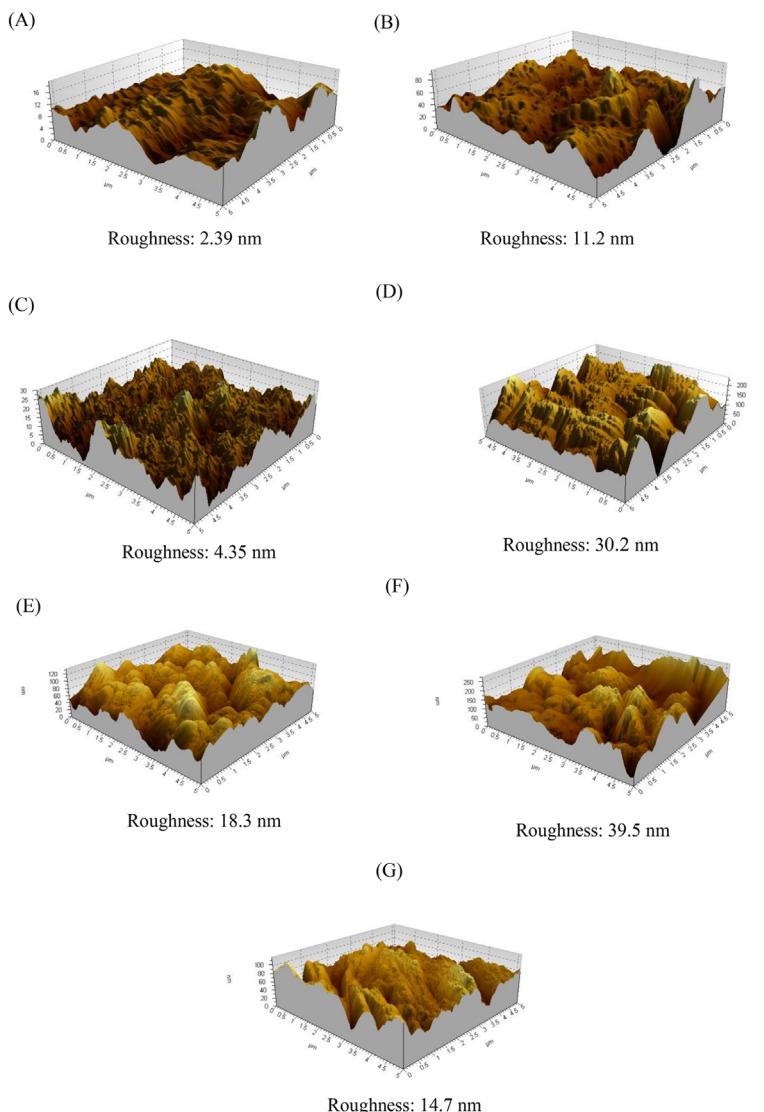


Figure 3. Atomic force microscopy of films made of native starch (A), native starch and polyethylene oxide (PEO) (B), phosphorylated starch (C), phosphorylated starch and PEO (D), cross-linked starch (E), cross-linked starch and PEO (F), and PEO film (G).

3.8. X-Ray Diffraction of Films

The X-ray diffraction and relative crystallinity of the native, phosphorylated, and cross-linked starches films, with and without PEO, are shown in Figure 1b. The peaks at 19.2° and 23.3° are more intense in the films with PEO compared to the films without PEO. According to Pereira et al.,^[3] the peak around 19° is related to the crystallographic plane (120), whereas the peak around 23° may correspond to the planes: (032), (132), (112), (212), (004), and (124). Furthermore, as shown in Figure 1a, pure PEO has peaks from 16° to 30° , which are characteristic of crystalline PEO structures.^[3]

The addition of PEO increased the relative crystallinity of the films due to its high crystallinity (Figure 1a). Moreover, the high crystallinity of PEO contributed to the decrease of the WVP values and to the increase of the tensile strength of cross-linked starch films (Table 2).

3.9. Atomic Force Microscopy of Films

The 3D topography of the films is shown in Figure 3. The images indicate structures with irregularities represented by alternating

dark and light regions. Roughness values vary for all films, with films with PEO having higher roughness values (Figure 3). Furthermore, the films containing PEO had a rougher surface (Figure 2) than films without PEO did, confirming that the PEO favors the formation of rougher biocomposites. Kaczmarek et al.^[30] reported that a film made of pectin and PEO had a high roughness (54.3 nm) due to the presence of spherulites of different sizes forming layers or “smooth hills” on the polymer surface.

3.10. Contact Angle of the Films

The films with added PEO (Figure 4B, D, and F) showed higher contact angle than the films without PEO did (Figure 4A, C, and E, and Table 2). Since higher contact angle indicates lower surface hydrophilicity, the surface of the film with PEO is supposedly less hydrophilic. Jayasekara et al.^[31] reported that starch and PVA blends were hydrophobic due to the presence of impurities on their surfaces or due to the three-dimensional helical structure of the polymer matrix.

The films with rough surfaces (Figures 2 and 3) also showed high contact angle (Figure 4). According to Jayasekara et al.,^[31]

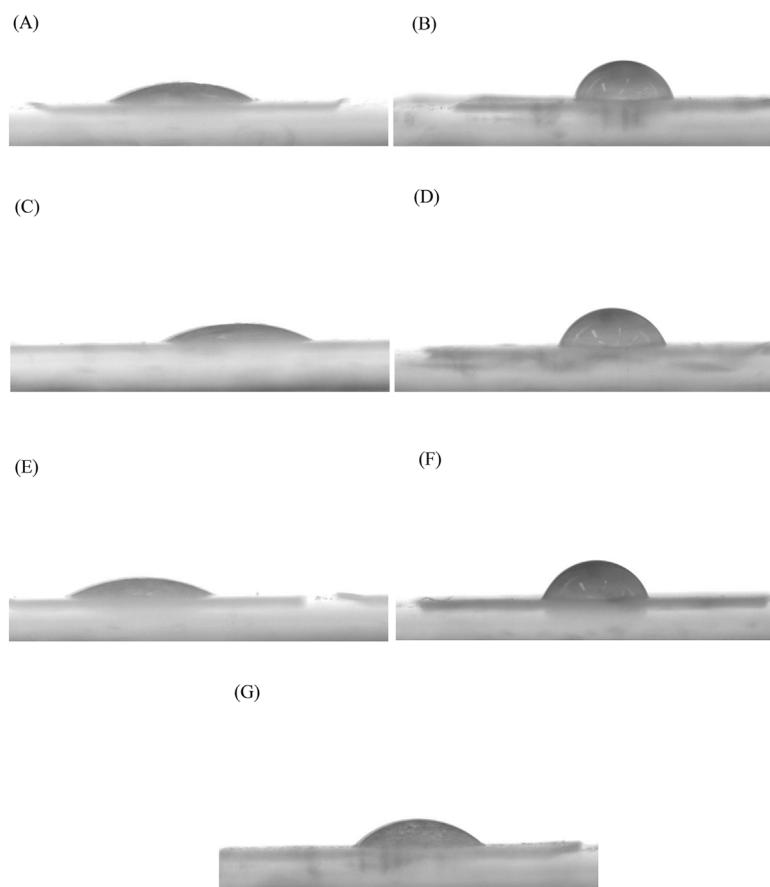


Figure 4. Contact angle of films made of native starch (A), native starch and polyethylene oxide (PEO) (B), phosphorylated starch (C), phosphorylated starch and PEO (D), cross-linked starch (E), cross-linked starch and PEO (F), and PEO film (G).

roughness is another factor that may be related to the increase of the contact angle. In fact, the contact angle of films increases with the surface roughness. Phosphorylated and cross-linked starches were produced to reduce the hydrophilicity of biocomposite films. As evident highlighted in Figure 4G and Table 2, PEO is less hydrophilic than starch, which is reflected by the contact angle value of pure PEO films. Therefore, the incorporation of PEO also contributed to the reduction of the hydrophilicity of biocomposite films.

4. Conclusions

Phosphorylation and cross-linking confer different characteristics to wheat starch. The phosphorylated starch has higher phosphorus content, amylose content, swelling power, and solubility than the cross-linked starch does. The granules of cross-linked starch are resistant to high temperatures due to inter- and intermolecular bonds. In addition, phosphorylated and cross-linked starches show decreased peak viscosity at their breakdown point, final viscosity, retrogradation, and relative crystallinity compared to native starch. Starch film containing PEO presented discontinuous morphology, was more crystalline, more rugged, and less hydrophilic than starch films without PEO did. Therefore, the films with and without PEO possess properties for different industrial applications, especially for flexible packaging. However, future studies are needed to evaluate all the potential applications.

Acknowledgements

We would like to thank CAPES, Embrapa, CNPq, FAPERGS, SCT-RS, CEME-SUL and Pólo de Inovação Tecnológica em Alimentos da Região Sul.

Conflict of Interest

The authors declare no conflict of interest.

Keywords

biocomposites, cross-linking, phosphorylation, polyethylene oxide, wheat starch

Received: July 23, 2017

Revised: December 6, 2017

Published online:

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