

processes. These morphological characters and those of the males matched those reported

in the original description of Aphelenchoides besseyi by Christie (1942) and a revision by

specimens from cotton were used for phylogenetic analysis to verify genetic congruence between the sequences of this population of *A. besseyi* from cotton and those published for

sequences were obtained for the near full length 225 of the ribosomal small subunit (SSU) and the expansion segment 226 of the large subunit (D2-D3 LSU) according to the protocol of de Jesus et al. (2016). The newly obtained sequences (MH187564 and MH187565) were assembled using BioEdit version 7.2.5 (http://www.mbio.ncsu.edu

Fortuner (1970) and Franklin and Siddigi (1972). Additional individual A. besseyi

other populations of this nematode in Brazil. The genomic DNA extraction and the

Your ORCID ID ensures you get credit for your work throughout your career.



Regulation,

and Evolution

of MTI and ETI

Molecular Plant-Microbe Interactions /bioedit/page2.html). Guide sequences to phylogenetic analysis were from Meyer et al. (2017). Sequence data were aligned using Mega 7.0 (ClustalW algorithm). The ambiguously aligned regions from SSU and LSU alignments were removed to optimize the phylogenetic analysis. For phylogenetic reconstruction, the best fitting model of sequence evolution was determined for each data set separately using the Akaike information criterion in Mega 7.0. The model GTR+G+I (general time reversible + gamma distributed with invariant sites) was selected for both SSU and LSU. Therefore, phylogenetic trees were constructed applying the maximum likelihood method with 100 bootstrap replications. Phylogenetic analysis indicated that the sequence of the cotton population of A. besseyi grouped with those of a population infesting soybean (Favoreto and Meyer 2017; Meyer et al. 2017) and other populations included in the group rice by de Jesus et al. (2016). The ability of A. besseyi to infect cotton was confirmed in two greenhouse experiments. For these experiments, specimens of A. besseyi obtained from field-infected cotton plants were reared on fungal cultures of Fusarium sp. growing in potato dextrose agar in Petri dishes. The reared nematodes were injected at the stem base of individual healthy cotton cultivar TMG 47B2RF seedlings growing in pots at a density of 6,000 specimens per seedling. The same field symptoms were observed in the inoculated seedlings, and a high number of nematodes were recovered. These results fulfill the modified Koch's postulates. The final nematode densities in the two experiments were 970 and 2,244 specimens per gram of fresh tissue. Field observations in Brazil also indicated that soybeans grown after or before cotton in the same nematode-infested area were damaged by the nematode as well (similar symptoms on soybean aerial tissues with population levels of 70 specimens/g). A. besseyi therefore poses a considerable threat to cotton-soybean cropping systems in Mato Grosso State in Brazil.

References:	Section:	Choose
Christie, J. R. 1942. Proc. Helminthol. Soc. Wash. 9:82. [Google Scholar]		

de Jesus, D. S., et al. 2016. Nematology 18:337. https://doi.org/10.1163 /15685411-00002962 [Crossref] [ISI] [Google Scholar]
Favoreto, L., and Meyer, M. C. 2017. Nematropica 47:ABST 13. [Google Scholar]
Fortuner, R. 1970. J. Helminthol. 44:141. https://doi.org/10.1017 /S0022149X00021702 [Crossref] [Google Scholar]
Franklin, M. T., and Siddiqi, M. R. 1972. C. I. H. Descript. Pl.-paras. Nematodes, Set. 1, No. 4, 3p. [Google Scholar]
Meyer, M. C., et al. 2017. Trop. Plant Pathol. 42:403. https://doi.org/10.1007 /s40858-017-0167-z [Crossref] [ISI] [Google Scholar]

Funding: Funding was provided by IMA-MT and Faped.

## Citation |

Journals Home | Books Home | APS Home | IS-MPMI Home | Contact Us | Permissions | Privacy | Copyright The American Phytopathological Society