

Special Issue Article

Cite this article: Avila LA, Marchesan E, Camargo ER, Merotto A Jr., Ulguim AR, Noldin JA, Andres A, Mariot CHP, Agostinetto D, Dornelles SHB, Markus C (2021). Eighteen years of Clearfield™ rice in Brazil: what have we learned? *Weed Sci.* doi: [10.1017/wsc.2021.49](https://doi.org/10.1017/wsc.2021.49)

Received: 4 November 2020

Revised: 13 June 2021

Accepted: 23 July 2021

Associate Editor:

Nilda Roma-Burgos, University of Arkansas












Keywords:

Integrated weed management; red rice; resistance to imidazolinone herbicides; sustainable weed management; weed resistance

Author for correspondence:

Luis Antonio de Avila, Departamento de Fitossanidade, Universidade Federal de Pelotas, Campus Universitário s/n, CP 354, 96.160-000, Pelotas, RS, Brazil. (Email: laavilabr@gmail.com)

Eighteen years of Clearfield™ rice in Brazil: what have we learned?

Luis Antonio de Avila¹ , Enio Marchesan² , Edinalvo R. Camargo³ , Aldo Merotto Jr.⁴ , André da Rosa Ulguim⁵ , José Alberto Noldin⁶ , André Andres⁷ , Carlos H. P. Mariot⁸ , Dirceu Agostinetto¹ , Sylvio H. B. Dornelles⁹  and Catarine Markus¹⁰ 

¹Professor, Crop Protection Department, Federal University of Pelotas (UFPEL), Pelotas, Brazil; ²Professor, Department of Crop Science, Federal University of Santa Maria (UFSM), Santa Maria, Brazil; ³Associate Professor, Crop Protection Department, Federal University of Pelotas (UFPEL), Pelotas, Brazil; ⁴Associate Professor, Crop Science Department, Federal University of Rio Grande do Sul (UFRGS), Porto Alegre, Brazil; ⁵Associate Professor, Crop Protection Department, Federal University of Santa Maria (UFSM), Santa Maria, Brazil; ⁶Senior Researcher, Epagri (Institution for Agricultural Research and Extension of Santa Catarina State), Itajaí Experimental Station, Itajaí, Brazil; ⁷Senior Researcher, Brazilian Agricultural Research Corporation (Embrapa), Pelotas, Brazil; ⁸Researcher-Technical Consultant, Rio Grande do Sul Rice Institute (IRGA), Cachoeirinha, Brazil; ⁹Associate Professor, Department of Biology, Federal University of Santa Maria (UFSM), Santa Maria, Brazil and ¹⁰Assistant Professor, Crop Science Department, Federal University of Rio Grande do Sul (UFRGS), Porto Alegre, Brazil

Abstract

Clearfield™ (CL) rice (*Oryza sativa* L.) is a weedy rice (*Oryza* spp.; synonym = red rice) control tool that has been used in Brazil since 2003. This system includes the use of an imidazolinone (IMI)-tolerant cultivar and the application of IMI herbicides. In this review article, Brazilian weed scientists evaluate the challenges and lessons learned over 18 yr of CL use. CL system benefits include selective weedy rice control, better crop establishment during the most advantageous period of the year, and more efficient fertilizer use. In Rio Grande do Sul state, the CL system, in conjunction with other improvements, has contributed to rice grain yield gains from 5,500 kg ha⁻¹ before 2002 to around 8,400 kg ha⁻¹ currently. In contrast, the main problem that has arisen over this period is the rapid evolution of IMI-resistant weedy rice, caused by gene flow from CL rice cultivars. The off-label use (rate and continuous use) of IMI herbicides has contributed to the evolution of resistance in *Echinochloa* spp. and other weeds. IMI herbicide carryover has also affected susceptible crops grown after CL rice. Crop rotation with soybean [*Glycine max* (L.) Merr.] is increasing, ensuring system sustainability. The importance of minimum tillage has also become apparent. Such cultivation includes applying nonselective herbicides before sowing or just before crop emergence (at the spiking stage to eliminate as much weedy rice as possible and other weeds at an early growth stage). It also includes the use of certified seeds free of weedy rice, following label instructions for IMI herbicides, applying the herbicide PRE followed by POST, and complementary weedy rice management practices, such as roguing of surviving weedy rice plants.

Introduction

The Clearfield™ (CL) rice production system is currently the most successful and effective system for weedy rice control. The adoption of the system globally was reviewed by Sudianto et al. (2014). In the current review, Brazilian weed scientists describe the challenges and lessons learned from 18 yr of CL rice production in Brazil. This review also provides critical insights on improving weedy rice management in CL rice and in emerging technologies.

Rice (*Oryza sativa* L.) is grown throughout agricultural areas in Brazil, and its cultivation is divided into two basic systems. The first system, adopted in the north and central regions, involves cultivation in upland rainfed areas; the second consists of lowland rice paddy cultivation, predominantly in the south. The primary rice-producing states in Brazil are Rio Grande do Sul (RS) and Santa Catarina (SC) in the south, and Tocantins (TO) in the north (CONAB 2020) (Figure 1). The southern states of RS and SC are the leading rice producers with a combined area of 1.15 million ha cultivated annually and producing more than 70% of Brazilian rice. Rice production in these states is primarily in flooded systems, and CL rice is planted in 70% to 80% of the total area. In RS, where rice production records began in 1921, rice was produced on 80,000 ha with an average yield of 2,190 kg ha⁻¹. The area and rice grain yield increased with time, reaching 400,000 ha and yielding 3,700 kg ha⁻¹ in the 1970s, and the area increased further to 1 million ha with an average yield of 8,400 kg ha⁻¹ in the 2019/20 growing season (IRGA 2020). Historically, in RS and SC, rice has been planted without crop rotation, which increased weed infestations, mainly weedy rice, grasses from



Figure 1. Rice production in Brazil in the 2019/20 growing season. The circles represent rice production in 558 Brazilian rice production microregions, with the largest circles representing the highest production. Abbreviations inside the map correspond to the states: AC, Acre; AL, Alagoas; AM, Amazonas; AP, Amapá; BA, Bahia; CE, Ceará; DF, Distrito Federal; ES, Espírito Santo; GO, Goiás; MA, Maranhão; MG, Minas Gerais; MS, Mato Grosso do Sul; MT, Mato Grosso; PA, Pará; PB, Paraíba; PE, Pernambuco; PI, Piauí; PR, Paraná; RJ, Rio de Janeiro; RN, Rio Grande do Norte; RO, Rondônia; RR, Roraima; RS, Rio Grande do Sul; SC, Santa Catarina; SE, Sergipe; SP, São Paulo; TO, Tocantins. Map courtesy of Alcido Elenor Wander (Embrapa Rice and Beans).

the *Echinochloa* genus (*Echinochloa* spp.), and sedges (*Cyperus* spp.). Weedy rice is a significant pest in rice crops in Brazil and worldwide, impacting rice yield and quality (Ziska et al. 2015). Weedy rice in southern Brazil is predominately of the *indica* and *indica-aus* type (Piveta et al. 2021; Qiu et al. 2020). In Brazil, the *indica*-type cultivars replaced the *japonica* type in the 1970s (Merotto et al. 2016). The origin of the *aus*-type background found in Brazilian weedy rice is not known.

Weedy rice management in Brazil can be divided in two periods: the period before the launching of CL technology, which we call the “coexistence era,” and the period after the launching of CL for weedy rice control, which we call the “selective control era.” Before the CL launch, there were no options for selective weedy rice control (Ziska et al. 2015). During the coexistence era, many tools were used, with limited effectiveness, to reduce weedy rice infestations and to mitigate their effect on rice yield (Andres et al. 2012; Avila et al. 2000b). Historically, weedy rice infestations were commonly managed with fallow periods and late sowing to provide intense soil preparation to reduce the seedbank. This management strategy was ineffective in controlling weedy rice and on using natural resources and inputs, resulting in low rice yield. Indeed, before the introduction of CL in Brazil, many areas could not be planted with rice due to severe weedy rice infestation (Agostinetto et al. 2001). Alternative cultivation practices such as direct sowing, minimum tillage, or water seeding became popular, with water seeding proving to be the most effective due to the presence of the water early in the growing season (Avila et al. 2000b). Other alternatives included soil preparation in the summer to reduce the seedbank, the use of winter crops, summer crop rotation, fallow periods, targeted application of nonselective herbicides on the remaining weedy rice plants (Andres et al. 2012; Avila et al. 2000b; Marchesan et al. 2003), and the use of

certified seeds free of weedy rice (SOSBAI 2018). Safeners applied to cultivated rice seeds had been proposed elsewhere for selective weedy rice control with clomazone based on pot experiments under greenhouse conditions (Busi et al. 2017). This technology was extensively evaluated in Brazil with the safener dietholate and the herbicide clomazone and resulted in reduced efficacy on weedy rice. The differences between field and pot experiments can be attributed to weedy rice diversity in the field and field plants being more robust than those grown in pots.

The CL rice system was made commercially available in Brazil in 2003, when the Rio Grande do Sul Rice Institute (IRGA) launched the cultivar ‘IRGA 422CL’ under a legal agreement with BASF SA (Lopes et al. 2003). The stewardship program proposed by BASF (2020) included: (1) use of certified seeds, (2) application of labeled herbicides, (3) adoption of label recommendations concerning herbicide application rates and timing, (4) establishment of permanent flooding soon after the POST herbicide application, (5) roguing the weedy rice escapes, (6) planting CL varieties for no more than two consecutive growing seasons, and (7) crop rotation. These stewardship protocols aimed to ensure that the system would provide a sustainable solution for weedy rice control.

The CL rice system brought enormous benefits to rice production in Brazil, as well as some important challenges. Understanding what we have learned from the CL system is important for maintaining the effectiveness of the system and better managing emerging weedy rice management technology. This review aims to give a brief overview of the challenges and benefits of the CL rice production system for weedy rice management in southern Brazil and to summarize what we have learned by managing this technology over the past 18 yr. The review is divided into cultivars and crop breeding, the contributions of CL to rice production in Brazil, the challenges of CL system in Brazil, and a synthesis of what we have learned.

Cultivars and Crop Breeding

Over the 18-yr history of the CL system in Brazil, public (IRGA, Embrapa, and Epagri) and private (BASF, Ricetec, Oryza, Agronorte, and Metropolitana) companies have launched and registered in Brazil 33 cultivars and hybrids with different resistance levels to imidazolinone (IMI) herbicides (MAPA 2020) (Table 1). These cultivars were initially developed through induced mutations of the *acetolactate synthase* (*ALS*) gene (Croughan et al. 1996). The first CL cultivar developed and commercialized in Brazil was IRGA 422CL (Lopes et al., 2003), obtained from the mutant line 93AS3510 (Croughan et al. 1996) as the resistance gene donor. In Brazil, the so-called first-generation cultivars originated from the 93AS3510 line, which has a single point mutation in the *ALS* gene causing the G654E substitution (Roso et al. 2010b). Following the launch of the IRGA 422CL cultivar, Ricetec launched the ‘Tuno CL’ hybrid in 2003 and Epagri launched the ‘SCS 115 CL’ cultivar in 2007. The Tuno CL hybrid carries the S653D mutation derived from the PCW16 line (Roso et al. 2010b). SCS 115 CL is a first-generation cultivar developed for the pre-germinated system (Schicocchet et al. 2007). Embrapa’s first CL cultivar was the ‘BRS Sinuelo CL’, obtained from the mutant 93AS3510 (Magalhães et al. 2011).

BASF registered the ‘Puitá INTA CL’ cultivar in 2008 and the ‘Guri INTA CL’ cultivar in 2012, both of which are considered second-generation cultivars harboring the A122T mutation (Roso et al. 2010b). Also considered to be second-generation IMI-resistant varieties are those developed from the PCW16 line, which are associated with the S653D mutation (Roso et al. 2010b) used in most hybrid varieties.

Table 1. Rice cultivars and hybrids with tolerance to imidazolinone herbicides available in Brazil for the Clearfield™ (CL) production system presented in chronological order of registration at Brazil's Ministry of Agriculture, Livestock, and Supply (MAPA 2020).

Cultivar/hybrid	Registration year	CL generation ^a	Company
IRGA 422CL	2002	1st	IRGA
Tuno CL	2003	2nd	Ricetec
Sator CL	2006	2nd	Ricetec
SCS 115 CL	2006	1st	Epagri
Avaxi CL	2007	2nd	Ricetec
XP710 CL	2007	2nd	Ricetec
Apsa CL	2008	2nd	Ricetec
Puitá INTA-CL	2008	2nd	BASF
BRS Sinuelo CL	2009	1st	Embrapa
Ecco CL	2009	2nd	Ricetec
Inov CL	2009	2nd	Ricetec
IRGA 428	2011	2nd	IRGA
RT5310 CL	2011	2nd	Ricetec
SCS 117CL	2011	1st	Epagri
Guri INTA CL	2012	2nd	BASF
IRGAP H7RI (Prime CL)	2012	2nd	IRGA/ Metropolitana
IRGAP H9RI (QM 1010CL)	2012	2nd	IRGA/ Metropolitana
IRGA 424 RI	2013	2nd	IRGA
XP111 CL (Titan CL)	2013	2nd	Ricetec
XP112 CL	2013	2nd	Ricetec
SCS121 CL	2014	2nd	Epagri
XP102 CL (Lexus CL)	2014	2nd	Ricetec
BRS A501 CL	2015	2nd	Embrapa
BRS A701 CL	2015	2nd	Embrapa
BRS A702 CL	2015	2nd	Embrapa
BRSCIRAD AH703 CL	2016	2nd	Embrapa
Primoroso CL	2016	2nd	Oryza
IRGA 431 CL	2017	2nd	IRGA
ANa9005 CL	2018	2nd	Agro Norte
BRS Pampa CL	2019	2nd	Embrapa
Memby Porá INTA CL	2019	2nd	BASF
Risobaco CL	2020	2nd	Oryza
Inov Fullpage	2020	2nd	Ricetec
XP113 Fullpage	2020	2nd	Ricetec

^aCL generation: 1st, originated from Louisiana State University, of the 93AS3510 lineage (point mutation in the *ALS* gene that confers the G654E substitution in the *ALS* enzyme); 2nd, originated from LSU, of the PCW16 lineage (point mutation in the *ALS* gene that confers the S653D substitution in the *ALS* enzyme) or INTA (point mutation in the *ALS* gene that confers the A122T substitution in the *ALS* enzyme). Source: MAPA (2020).

The first-generation CL cultivars were replaced, because they had limited selectivity to IMI herbicides, resulting in severe crop injury. At present, the most widely used CL cultivars in Brazil are the second-generation 'IRGA 424 RI' from IRGA (RS) and 'SCS121 CL' from Epagri (SC). The hybrids launched in Brazil in the past 18 yr have occupied a small percentage of the total rice cultivated area. The list of all CL cultivars and hybrids registered in Brazil is shown in Table 1.

Contributions of the CL System to Rice Production in Brazil

Weedy Rice Control

Before the introduction of the CL system, selective weedy rice control was not possible; most paddy fields were highly infested with weedy rice, and some areas were abandoned (Merotto et al. 2016).

The CL rice system was the first and most successful selective weedy rice control in rice. The IMI herbicides, which are inhibitors of the acetolactate synthase (*ALS*) enzyme (EC 2.2.1.6, formerly EC 4.1.3.18), are used to control weedy rice in the CL system.

When the CL system was first introduced in Brazil, the manufacturer recommended a single POST application of the IMI herbicides imazethapyr + imazapic (Villa et al. 2006). This approach was not effective for weedy rice control, and thus the recommendation was modified to PRE followed by early POST application, which resulted in higher levels of weedy rice control (Marchesan et al. 2011b; Santos et al. 2007; Villa et al. 2006). With this new approach, the CL system revolutionized weedy rice management, attaining 99% control with two sequential applications.

Successful weedy rice control with this technology reduced the use of integrated weed management, and many producers use the CL system as their single tool for weedy rice management. On the other hand, producers who continued to use integrated weed management had more stable and sustainable weedy rice management. Although research on integrated weedy rice management has been less intense since the introduction of CL rice, several alternatives to weedy rice control methods are currently being adopted due to the increasing evolution of IMI-resistant weedy rice.

Adequate weedy rice control with the CL system depends on crop rotation, use of certified cultivated rice seeds, alternation between wet seeding and dry drill-seeded establishment systems, and roguing of weedy rice escapes. Different adjustments of these practices are being used in small and large farms to attain adequate crop and weed management of IMI-tolerant rice (Merotto et al. 2016). Other practices are also useful in suppressing weedy rice infestations and contribute to the reduction of its seedbank, such as adopting the pre-germinated system (Avila et al. 2000a), crop rotation, and planting earlier in the season in early September (Sartori et al. 2014).

An additional strategy is to prepare the land immediately after rice harvest to reduce the weedy rice seedbank. Laying rice straw on the soil surface has shown promise, especially with tools such as a "knife roller," which keeps the seeds (weeds and rice seeds) on the surface topsoil layer and stimulates their germination, reducing the weedy rice seedbank (Massoni et al. 2013). However, this off-season management was mentioned by only 2% of consultants in the RS rice areas (Ulguim et al. 2021). According to Noldin et al. (2006), producers should avoid incorporating weedy rice seeds into the soil, because seeds left on the surface are more likely to germinate and be controlled by natural agents or with herbicides.

Integration with livestock is an important strategy in rice cultivation. Fallow reduces the weedy rice seedbank, especially with the presence of animals and their trampling action. Cattle production in the fallow period can reduce the weedy rice seedbank by 85% per year (Marchesan et al. 2003). Marchesan and colleagues (2003) also reported that seeds located on the soil surface (0- to 1-cm deep) lose viability faster than those located at greater depths. However, it is essential to quarantine animals for at least 6 d between fields to avoid weed infestation in new areas, as viable weedy rice seeds can be found in the cattle feces until the 5th day after ingestion (Viero et al. 2018). Another strategy for weedy rice control is nonselective herbicide application, usually glyphosate, at the spiking stage of cultivated rice (*S*₃ stage) (Counce et al. 2000). This strategy involves applying the herbicide before complete rice emergence, while only part of the coleoptile is visible (SOSBAI 2018). The integrated weedy rice management strategy is

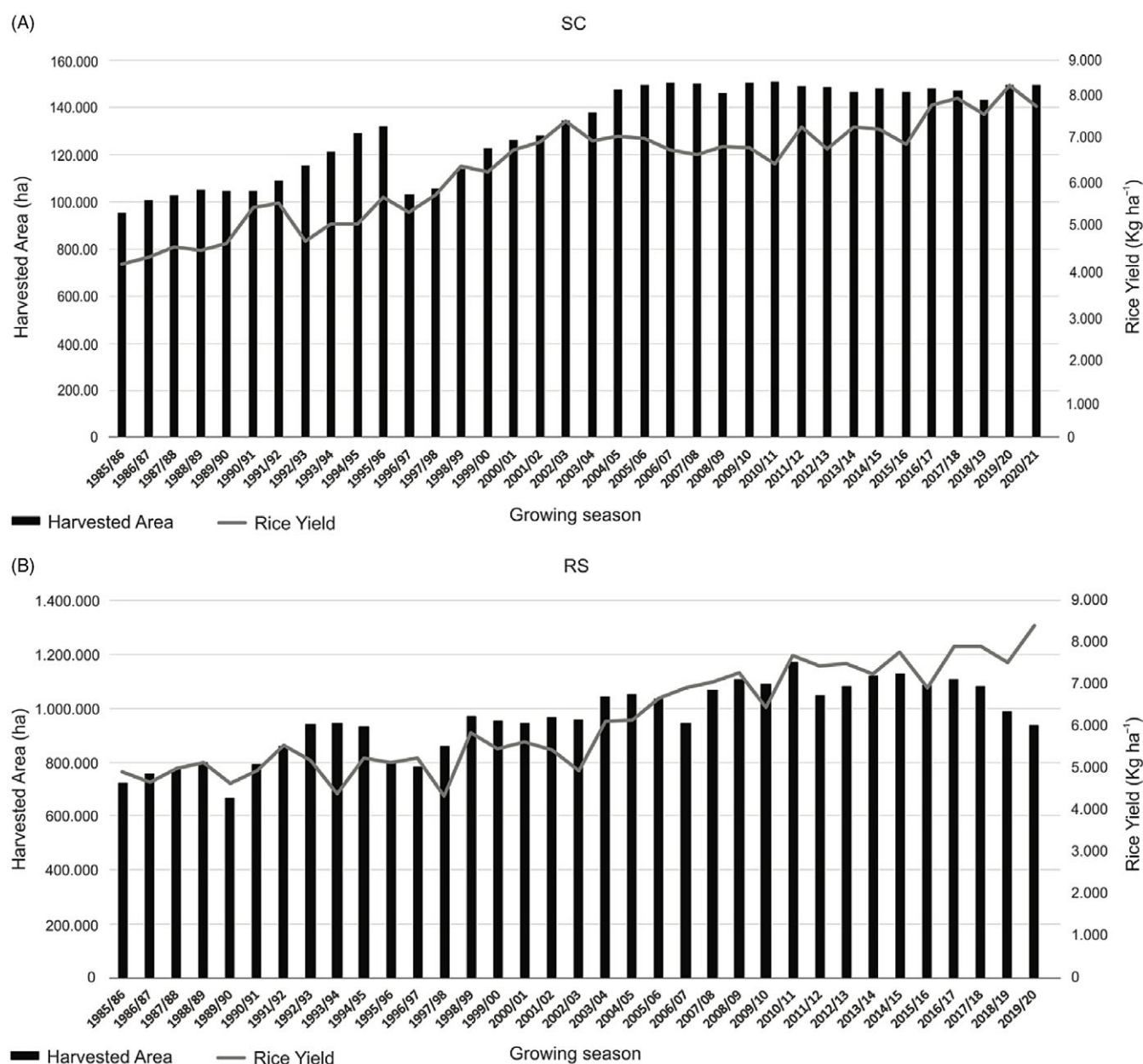


Figure 2. Evolution of rice yield and harvested area in the states of (A) Santa Catarina (SC) and (B) Rio Grande do Sul (RS) 18 yr before (1985–2003) and after (2003–2021) the launch of the Clearfield™ (CL) rice system. Data source: IRGA and EPAGRI.

important to extend the lifetime of the technology and to ensure efficient weedy rice management.

Control of Other Weeds

Besides affording selective weedy rice control, IMI herbicides are broad-spectrum (Alister and Kogan 2005), controlling a wide variety of species, including many broadleaf weed species and sedges (Krausz and Kapusta 1998). An exception is imazethapyr, which is less effective on sedges and is not effective on some broadleaf weeds, especially those of the Fabaceae family, which includes the *Aeschynomene* and *Sesbania* genera (Pellerin et al. 2003, 2004).

The application of IMI herbicides in CL rice in conjunction with other herbicides has provided a good weed control spectrum. Due to the CL system's success in managing weeds, one negative consequence was its use in areas where weedy rice was not present

or in fields where the weedy rice infestation levels were not high enough to justify its use. One undesirable effect on these areas is related with the carryover effect of IMI herbicides. Another consequence of the success of IMI system is that most rice farmers adopted the CL system for many years (see "Continuous Use of CL Rice").

Rice Grain Yield Increase

In RS, rice yields had increased rapidly overall after CL rice commercialization (Figure 2). Indeed, by the 2019/20 season, 85% of the RS rice production area (IRGA 2020) and more than 60% of the rice-growing area of SC were planted with CL cultivars (Avila et al. 2021). The yield increase promoted by the CL system was due to more effective weed control, especially of weedy rice, and the simplicity with which certain management practices can

Table 2. History of the number of weedy rice seeds allowed in commercial rice seeds in Rio Grande do Sul.

Period	Number of red rice seeds in commercial seeds ^a	Source
Through 1977	20 seeds 500 g ⁻¹	ST Peske (2021)
1978–1984	12 seeds 500 g ⁻¹	ST Peske, personal communication
1985–1988	5 seeds 500 g ⁻¹	ST Peske, personal communication
1989–1992	3 seeds 500 g ⁻¹	ST Peske, personal communication
1993–2003	2 seeds 500 g ⁻¹	ST Peske, personal communication
2004–2005	1 seed 500 g ⁻¹	ST Peske, personal communication
2006–2013	Foundation seed = 0 C ₁ = 0 C ₂ = 0 S ₁ = 1 seed 700 g ⁻¹ S ₂ = 2 seeds 700 g ⁻¹	MAPA (2005) ^b
Since 2013	Foundation seed = 0 C ₁ = 0 C ₂ = 0 S ₁ = 1 seed 700 g ⁻¹ S ₂ = 1 seed 700 g ⁻¹	MAPA (2013) ^b

^aFoundation seeds; C, certified seed; S, noncertified seed.

^bFederal law applied nationwide.

be implemented, such as sowing at an optimal time, so the flowering and grain-filling period will happen when maximum solar radiation availability occurs, and response to fertilization and other production factors (Merotto et al. 2016).

Additional yield improvements under the CL rice system were achieved through institutional programs, including Projeto 10 from IRGA, CFC from Fondo Latinoamericano para Arroz de Riego (FLAR), and Marca from Embrapa, which promoted crop management practices to improve yields and better management of weedy rice and other critical weed species such as barnyardgrass [*Echinochloa crus-galli* (L.) P. Beauv.] and rice flatsedge (*Cyperus iria* L.) (Gomes et al. 2004; Menezes et al. 2004). As a result, many rice farmers rapidly adopted the CL system, and average grain yield during the last 18 yr increased by more than 2,000 kg ha⁻¹, reaching 8,400 kg ha⁻¹ in RS (IRGA 2020; Merotto et al. 2016). Although grain yield increase due to the CL system is unquestionable, problems involving IMI herbicide resistance in grasses and sedges have increased as will be addressed in the “Weed Resistance to Herbicides” section.

Challenges for CL Rice Production in Brazil

Seed Production System

Rice seed quality is vital to the success of weedy rice control. The use of certified seeds free of weedy rice seeds is fundamental to the success of the CL rice production system. Historically, the legislation for rice seed production in Brazil allowed weedy rice grains in commercial seeds. In the 1970s, the seed law allowed up to 20 weedy rice grains in 500 g of rice seeds. Nowadays, zero weedy rice seeds are allowed per 500 g for certified seed, and 1 and 2 weedy rice seeds allowed per 500 g for S₁ and S₂ seeds, respectively (SOSBAI 2018). S₁ seeds are non-certified seeds, first generation, originated from foundation or certified seeds (C₂ or C₂); S₂ seeds are non-certified seeds, second generation, originated from foundation, certified (C₁ or C₂), or S₁ seeds. In the 1980s, a state-certified seed production program was initiated in SC that

expressly prohibited the presence of weedy rice seeds (Noldin et al. 1997). The certified seed production program was gradually expanded and subsequently adopted in RS.

The changes in rice seed quality with respect to weedy rice contamination are summarized in Table 2. The launch of the CL system in 2003 stimulated discussion concerning tougher legislation regarding rice seed quality as it relates to weedy rice contamination. In 2009, Brazilian legislation finally established zero tolerance for weedy rice in certified seed (MAPA 2013). However, many producers prefer to produce their seeds and even sell noncertified seed that contains weedy rice. Goulart et al. (2014) showed that the genetic diversity of weedy rice resulting from uncertified seeds is high due to seed migration between fields. Only approximately 60% of growers in RS and 76% of growers in SC use certified seed, despite a push by public and private institutions to increase its use (Martins et al. 2017).

Before introduction of the CL technology in RS, only 17% of seeds used by producers were free of weedy rice seeds (Marchesan et al. 2001). In addition to gene flow via pollen between CL rice and weedy rice, noncertified seed has contributed significantly to proliferation of IMI-resistant weedy rice and reduced effectiveness of the CL system in many production areas. Weedy rice plants have recently been found with characteristics similar to cultivated rice, with respect to plant size, vegetative cycle, and seed types (Piveta et al. 2021). The main difference between these weedy varieties and the cultivated rice plants is the reddish pericarp, which can only be detected in laboratory analysis. Other characteristics such as seed shattering and seed longevity are common characteristics of weedy rice. Recent data from seed certification studies in RS showed that 12% of seed lots were infested with weedy rice, even though no weedy rice had been identified during field inspections. This could reflect contemporary weedy rice populations that are more crop-like escaping detection. Ongoing technical discussions have focused on maintaining the production of quality seed free of weedy rice (JAN, personal observation).

Farmers' Training

One of the initiatives that boosted grain yield following the introduction of CL rice was Projeto 10, which consisted of several strategies to transfer the available technology to rice growers in RS via an intensive training program for farmers (Menezes et al. 2004). The aim was to minimize the cultivar yield gap. The project integrated management practices by implementing a technology transfer system that involved establishing demonstration plots in different rice-producing regions and discussing management practices with producers and technicians. Basically, the program was based on use of the CL rice system to control weedy rice and the optimization of other crop management practices.

Despite intense information campaigns by public and private entities on the correct use of CL rice, the technology was adopted by many producers who failed to follow the stewardship protocols; thus, many farmers were unable to control weedy rice adequately. There is a need to continue monitoring how rice growers use the CL technology, to educate growers, to provide technical support, and to understand the residual effect of IMI herbicides on rotational crops (Sudianto et al. 2014). Such a program must offer sustainable solutions for weedy rice management and a permanent and intensive educational program devoted to rice producers. One of the lessons learned from the use of the CL in Brazil is the need for consistent information and widespread training of extension agents and farmers on this production system.

Continuous Use of CL Rice

Since the launch of the CL system in Brazil, one recommendation has been the use of the system for no more than two consecutive growing seasons (BASF 2020; Eberhardt et al. 2007, 2015; Menezes 2003). However, in a recent survey (Avila et al. 2021), only 40% of RS producers and 15.2% of SC producers reported using CL cultivars for up to three consecutive years. In RS, 26.7% of producers have used the CL technology for more than 10 consecutive years. In SC, 78.8% of producers did not answer this question, as 48.5% of them do not use this technology.

The principal reasons for the continuous use of CL rice were the possibility of improving rice yields through the control of weedy rice (Marchesan et al. 2011b), the relatively low cost of the weed control program compared with conventional programs, and the convenience of using one herbicide to control virtually all the weeds in rice (Menezes 2003). Other reasons included the carry-over effect on non-CL cultivars (Avila et al. 2010b), risk of herbicide drift to adjacent non-CL rice fields (Dal Magro et al. 2006), and the fact that the seedbank of weedy rice could persist in the field for at least three seasons (Marchesan et al. 2011a). Therefore, although there are justifications for the widespread adoption of the CL system with cultivation in consecutive growing seasons, the consequences must be better understood to plan management practices that minimize its adverse effects.

Lack of Crop Rotation

Historically, crop rotation has not been adopted in the production of irrigated rice in southern Brazil. Before the launch of the CL rice system, farmers in RS used to keep part of the rice areas in fallow as pasture for cattle. The lack of crop rotation is partially due to the difficulty of managing dryland crops in lowland environments. The lowland soils are highly compacted due to their physical and chemical characteristics and the intensive soil preparation for rice production, with low total porosity and hydraulic conductivity (Drescher et al. 2017; Giacomeli et al. 2016), resulting in high mechanical resistance (Henry et al. 2018). Thus, the lowland soils are frequently subject to water scarcity or excess, low levels of organic matter and phosphorus, and high acidity, requiring a higher investment in fertilization (Vedelago et al. 2012).

However, the growing number of problems associated with the CL system, such as resistant weedy rice evolution, led researchers and producers to intensify the search for alternatives. The alternatives included soybean [*Glycine max* (L.) Merr.], which started very slowly, with only 11,000 ha in the 2009/10 growing season compared with approximately 1 million ha of cultivated rice (IRGA 2020). Widespread appreciation of the need for crop rotation and a greater understanding of the limitations of alternative crops in floodplain soils has led to greater adoption of soybean in rotation with rice. Lately, soybean crop rotation in the lowlands has reached 341,288 ha in RS (IRGA 2020). This area may be considered low compared to the total rice production area but is a remarkable improvement for the rice-based production system. Also, the use of soybean in crop rotation in farmers' fields is now established in up to 25% of the total rice production area (Ulguim et al. 2021), with the remainder being rice or fallow.

An important aspect in crop rotation in lowland areas is the use of cover crops adapted to the floodplain environment. The most important winter cover crop is ryegrass (*Lolium multiflorum* Lam.), which benefits rice and soybean cultivated in the summer (Moraes et al. 2009). The crop rotation associated with cover crops prevents the increase of weedy rice in the seedbank (Avila et al.

2000b; Ulguim et al. 2018). The cultivation of soybeans and the necessary management demonstrate how changing practices can make weed control tactics more effective. The benefits of crop rotation include the diversification of herbicide mechanisms of action (MOAs) used in other crops in addition to those used in rice. Growers should minimize the use of glyphosate in the soybean rotation to avoid the chances of increasing evolution of weed resistance to glyphosate, especially weedy rice.

Lack of Herbicide Rotation

As previously discussed, there has been widespread noncompliance with the recommendation that the CL system should not be used in the same field over more than two consecutive growing seasons. Furthermore, there has been minimal rotation of the herbicides used. In a recent survey covering the last nine growing seasons in RS, ALS herbicides were used for POST weed management in approximately 95% of the surveyed fields (Ulguim et al. 2019b). PRE or "spiking" application had greater diversity in terms of the herbicide MOA, with ALS herbicides used in 25% of cases (Ulguim et al. 2019b). It is noteworthy that the primary approach to eliminating resistant weeds in RS has been nonselective herbicides at the spiking stage (S_3) (Fruet et al. 2020).

In southern Brazil, 65% of rice farmers combined different herbicides, while 38% rotate herbicides with different MOAs to manage resistant weeds (Fruet et al. 2020). That study also showed that 52% of producers increased the rate of herbicide application on their farms. In another survey, 92% of producers indicated that they applied herbicides at higher levels than the labeled rate (Ulguim et al. 2019b). Increased herbicide application is a major concern, because even though producers may initially improve weed control, this practice increases the selection pressure for weed resistance (Agostinetto et al. 2011).

The continuous use of CL rice, the lack of diverse herbicide MOAs, and higher herbicide rates are factors that contribute to more frequent weed resistance to ALS-inhibiting herbicides in rice farms. In general, few growers adopt herbicide rotation to minimize the evolution and spread of weed resistance to herbicides (Kalsing et al. 2019; Norsworthy et al. 2013).

Gene Flow

Although cultivated rice and weedy rice are autogamous plants, cross-fertilization can occur. While the frequency is generally less than 1% (Gealy et al. 2003), it can sometimes reach as high as 52% (Langevin et al. 1990). In 2002, the first study in Brazil on conditions of gene flow between a breeding line of genetically modified cultivated rice and two weedy rice genotypes (one strawhull and one blackhull) showed values ranging from 0.02% to 0.48% (Noldin et al. 2002). Subsequently, a natural crossing rate of 0.065% was reported between cultivated rice and weedy rice (Villa et al. 2006). Due to the high levels of weedy rice control achieved in the CL system (>95%), many producers discontinued roguing (Eberhardt et al. 2007, 2015). Consequently, weedy rice resistance to IMI herbicides evolved rapidly in several rice-growing regions of RS due to gene flow (Menezes et al. 2009).

Studies of the ALS gene mutation associated with resistance in weedy rice revealed the predominance of the G654E mutation, which is harbored by the first-generation cultivar IRGA 422CL (Roso et al. 2010b). The S653D and A122T mutations were also detected in some outcrosses. The S653D and A122T mutations were present in hybrids and the Puitá INTA CL cultivar,

respectively; these mutations confer a higher level of resistance than the G654E mutation (Roso et al. 2010b).

The hybridization rate with different rice cultivars was evaluated in approximately 1 million seeds in a field experiment using the concentric circle method (Goulart et al. 2014). In this study, the occurrence of gene flow for weedy rice from the genotypes IRGA 422 CL (G653E), 'Sator CL' (S653D), Puitá INTA CL (A122T), and resistant weedy rice (G653E) was 0.024%, 0.027%, 0.019%, and 0.022%, respectively. The highest hybridization from Sator CL was likely due to its floral biology, which is more prone to pollen release. The details about the floral biology of the evaluated hybrid cultivar that favored hybridization is unknown. The data also demonstrate pollen flow from HR weedy rice to the natural weedy rice population. Therefore, once a few resistant weedy rice plants are present, resistance will spread quickly from cultivated rice and weedy rice via gene flow.

Weedy rice resistance to IMI herbicides can also arise from independent selection, as demonstrated in an exclusion paternity analysis conducted on IMI-resistant individuals of weedy rice (Goulart et al. 2012). Seed migration is another means of gene flow when seeds from resistant weedy rice plants are present in cultivated rice seed, which sometimes occurs in noncertified seed. The lessons learned from the evolution of resistant biotypes in the CL system, especially gene flow, should be considered in developing new technologies involving herbicide-resistant rice cultivars.

Weed Resistance to Herbicides

Weedy rice resistance evolution to IMI herbicides is the largest concern in CL rice. In Brazil, resistant weedy rice was detected 3 to 4 yr after adoption of the CL rice production system (Merotto et al. 2016). A long-term study conducted between 2006 and 2012 found resistant weedy rice in 56% to 100% of the analyzed suspected populations (Kalsing et al. 2019), indicating the high occurrence of IMI-resistant weedy rice in rice cultivation in southern Brazil. Gene flow was the primary origin of IMI-resistant weedy rice, but independent selection occurred in 1.1% of the weedy rice plants evaluated, a frequency similar to that in other weeds subject to herbicide selection pressures (Goulart et al. 2012). IMI-resistant weedy rice plants have increased production costs and reduced yield (Merotto et al. 2016).

Currently, infestations of the *Echinochloa* complex resistant to herbicides are as prevalent as weedy rice in Brazil. Biotypes of *E. crus-galli* resistant to quinclorac were first identified in 1998, with resistance becoming widespread even before the CL system was introduced (Heap 2021; Schaedler et al. 2008). Initially, the IMI herbicides used in CL system were highly effective against *Echinochloa* spp., including quinclorac-resistant populations. However, approximately 4 yr after the introduction of the CL system, IMI-resistant *E. crus-galli* populations were identified (Merotto et al. 2016). Between 2009 and 2011, a study evaluating 624 populations of *E. crus-galli* identified IMI resistance in 81% of the populations, with resistance broadly distributed across RS rice-growing regions (Matzenbacher et al. 2015). Resistance to quinclorac was also found in 19% of the evaluated populations, but all were susceptible to cyhalofop-butyl. Recently, *E. crus-galli* biotypes resistant to glyphosate were found in soybean fields in southern Brazil, signaling the risk of resistance evolution to this herbicide in rice (Heap 2021).

In *Echinochloa* spp., the mechanism of resistance to IMI is associated with the mutations W574L and S653N of the *ALS* gene and enhanced herbicide detoxification caused by the increased

expression of *cytP450* and *glutathione S-transferase* genes (Dalazen et al. 2018). However, the mechanisms of resistance to quinclorac in these biotypes remain unclear. These results indicate that *E. crus-galli* populations in Brazil have evolved complex multiple herbicide resistance that may result in biotypes that are resistant to herbicides with a variety of MOAs, as has already occurred in other rice-growing regions outside Brazil (Rouse et al. 2018). Resistance to the ACCase inhibitor cyhalofop-butyl is already present in SC (Eberhardt et al. 2016), indicating increasingly complex herbicide resistance in *E. crus-galli*. The occurrence of multiple resistance and the lack of complete *E. crus-galli* control in soybean rotations with rice demonstrate that *E. crus-galli* is as great a threat to rice cultivation as weedy rice.

Herbicide resistance among sedges has also occurred in areas under the CL rice system. Resistance initially evolved in small-flower umbrella sedge (*Cyperus difformis* L.), and more recently in *C. iria* (Heap 2021). Populations of *C. iria* in RS showed resistance to ALS-inhibiting herbicides with a resistance factor (RF) of 5.49 for ethoxysulfuron and 2.47 for pyrazosulfuron (Spatt et al. 2016). Resistance to ALS-inhibiting herbicides has also occurred in globe fringerush [*Fimbristylis miliaceae* (L.) Vahl], while California arrowhead (*Sagittaria montevidensis* Cham. & Schldtl.) has evolved multiple resistance to ALS inhibitors and bentazon, a photosystem II-inhibiting herbicide (Heap 2021).

One of the most important tools for mitigating the evolution of resistant genotypes in weeds is rotation of herbicide MOAs (Burgos et al. 2008; Ulguim et al. 2017). Good practices for reducing resistance evolution include rotating herbicide use spatially, temporally, or both (Hicks et al. 2018; Norsworthy et al. 2012). However, the rotation of herbicide MOAs for weedy rice control in rice can only be accomplished through crop rotation, as we presently lack alternatives for weedy rice control in rice other than the CL system, but as described earlier, there are several limitations for rotation in lowland areas, resulting in the high selection pressure of IMI weed resistance.

Although rice producers and extension agents understand the strategies needed to manage and mitigate resistance, farmers are reluctant to adopt them for economic reasons (Fruet et al. 2020; Norsworthy et al. 2013; Ulguim et al. 2019b). Researchers have developed protocols for the rapid diagnosis of weedy rice herbicide resistance at the seed, seedling, and tiller stages and using molecular markers to identify the three types of mutations in the *ALS* gene (Roso et al. 2010b, 2010a). However, these protocols have not become widely used, and the producers generally identify herbicide resistance as a problem once resistant biotypes cover a large area of their farm.

Over the years, we have observed different experiences with CL rice on the part of producers. Some farmers noticed problems with resistant weeds shortly after they began using CL system. In other cases, the technology has remained effective due to effective management slowing the evolution of herbicide-resistant weeds. Producers who effectively adopted appropriate control methods were able to obtain better performances in the long term. A valuable lesson is that we cannot underestimate the massive selection pressure that herbicides can exert on weed populations, resulting in quick evolution of complex resistance mechanisms.

Volunteer CL Rice

A low proportion of rice seeds shatter at harvest or fall from the combine. Shattered seeds that are not winterkilled or lost through predation emerge as volunteer rice plants in the following season.

Hybrid rice has a higher degree of seed shattering than inbred rice, resulting in a higher population of volunteer rice. These volunteers have low yield and grain quality, which affects the crop's productivity and quality (Singh et al. 2017). Therefore, volunteer rice is another type of weedy rice. IMI-resistant volunteer rice cannot be controlled by either PRE or POST herbicides. Nonuniform establishment of volunteer rice can affect grain quality, especially if plants are from different crop seasons. Grains with a different pericarp color or shape from that of the current cultivar affect a crop's commercial classification. Also, high numbers of volunteer plants can result in overpopulation, with the volunteers being less productive, and reduces overall rice yield.

An additional concern is that herbicide-resistant volunteer rice plants, such as CL rice, may become agents for the flow of resistance genes to weedy rice populations (Sudianto et al. 2014). The difficulties in chemical control of volunteer CL rice highlight the importance of management before and after cultivation. Before cultivation, management should focus on preventing the emergence of volunteer plants. Postharvest management should prevent plants that were not eliminated during rice cultivation from reproducing and contributing to the seedbank.

In areas where rice is cultivated after CL rice, the producers in RS till the fields during the fallow season and delay rice planting to minimize the population of volunteer rice. Where crop rotation is practiced, the volunteer CL rice is controlled by burndown and PRE herbicides. One important practice is to minimize seed shattering during harvest and to not incorporate the seeds in the soil, allowing for predation and seed deterioration (Noldin et al. 2006). In Brazil there is lack of information on volunteer plants from hybrids and inbred cultivars. In Arkansas, Singh et al. (2016) report that after overwintering and burial for 130 d and 160 d, the survival rate of hybrid rice seeds (53% and 13%, respectively) was higher than that of conventional rice seeds (27% and 8%, respectively) across various depths of seed burial. It is important to study the behavior of volunteer rice to improve the management of weedy rice.

Soil Management in the Winter

Winter soil management affects several aspects of rice cultivation by creating a more suitable environment for rice planting in the next season. In addition, soil management during winter provides weed control and increases IMI herbicide degradation (see the "Herbicide Carryover" section). The primary winter management practice in RS is to allow livestock to graze on rice regrowth after harvest (Denardin et al. 2020), often without establishing a cover crop. Fallow and livestock are the predominant systems adopted during winter according to 44% and 49% of rice consultants' responses, respectively (Ulguim et al. 2021). After most of the rice plant regrowth is consumed by cattle, the soil is tilled, because rice straw degrades slowly during winter, and the soil is rutted by the rice combine harvesters (Botta et al. 2015).

A wide variety of species are suitable for use as cover crops during the winter in southern Brazil. However, few of these species are adapted to saturated or flooded soil, a common situation in rice production areas (Menezes et al. 2001). The native serradella [*Ornithopus micranthus* (Benth.) Arechav.], an species endemic to southern Brazil, northeastern Argentina, and Uruguay (Stępkowski et al. 2018), can be used as a cover crop during the winter in rice-producing areas, because it is well adapted to wet soils and can establish symbiosis with nitrogen-fixing bacteria (Menezes et al. 2001). Furthermore, its use results in increased rice

grain yield compared with fallow fields (Correia et al. 2018). Another option for cover crops is legumes such as white clover (*Trifolium repens* L.), red clover (*Trifolium pratense* L.), and Persian clover (*Trifolium resupinatum* L.), which are well suited for use in lowland cattle grazing (Weinert et al. 2019). However, the most common cover crop is ryegrass, because it grows well in the lowland areas and can be used as a cover crop or as forage for cattle grazing (Ulguim et al. 2019a).

Cover crop production in the winter favors nutrient cycling while adding crop residues in the soil, improving the system's biodiversity (Carvalho et al. 2010). However, if the cover crop produces a high amount of biomass, it can reduce rice emergence and initial growth and reduce yield. Some cover crops such as ryegrass also produce allelochemicals (Silveira et al. 2019), which can harm rice seedlings, affecting initial rice development (Bohnen et al. 2005).

The use of winter cover crops is beneficial for weed management in the CL system. In a study comparing areas in which rice was cultivated for five consecutive growing seasons without rotation to areas that use an integrated crop–livestock system, the number and diversity of weeds were higher in fields without crop rotation (Ulguim et al. 2018). In particular, the population of *C. iria* was 50% lower in fields with crop rotation. The authors also observed an increase in the number and diversity of weeds when no winter cover crop was used, with a high incidence of Indian goosegrass [*Eleusine indica* (L.) Gaertn.], compared with fields with ryegrass. Unfortunately, most rice producers and extension agents in southern Brazil do not consider winter cover crops to be essential in weed management, and they therefore do not adopt this practice, contributing to higher weed populations and the evolution of herbicide-resistant weeds (Fruet et al. 2020; Ulguim et al. 2019b). Even though scientists have indicated several essential practices for winter land management, such as drainage, soil preparation, and use of cover crops, few producers have adopted these technologies, preferring to leave the area fallow over the winter instead (Ulguim et al. 2021).

Herbicide Carryover

IMI herbicides are persistent in the soil. The half-life of imazethapyr is 60 to 90 d, of imazapyr 25 to 142 d, and of imazapic 120 d (Shaner 2014). Until 2009, the information available about the fate of IMI herbicides was primarily for aerobic conditions driven by soil aerobic microorganisms (Kraemer et al. 2009). Much more has been learned about the environmental fate of IMI in the last decade, especially in studies of rice cultivation in RS (Pinto et al. 2009a, 2009b, 2011). The persistence of herbicides in the field depends on the amount of herbicide applied and the dissipation process. The continuous use of the CL system may increase the carryover effect of IMI herbicides, as accelerated degradation has not been observed thus far (Bundt et al. 2015b).

Herbicide dissipation in lowland rice fields is quite different than in upland areas. Although continuous flooding (the most common rice irrigation method in southern Brazil) creates a favorable environment for surface transport of herbicides, minimal herbicide movement via runoff occurs in rice fields, because the areas have minimal slope or are practically flat (Martini et al. 2013). The lack of oxygen in such systems reduces microorganism activity, thus reducing herbicide degradation. Most IMI herbicides degrade slowly in anaerobic soils (Flint and Witt 1997). Alternative water management systems, such as intermittent flooding (Martini et al.

2013) and sprinkler irrigation (Helgueira et al. 2019), may be beneficial in reducing the persistence of IMI herbicides.

Water availability during the winter is another factor that affects herbicide carryover. Herbicide carryover in rice fields is higher during extremely dry or extremely wet winters. Excess water in the soil can move herbicides closer to the rice roots (Bundt et al. 2010). In dryer soils, herbicides may not be available for degradation, as moisture affects herbicide sorption, and the herbicide moves upward in the soil profile with water evaporation (Bundt et al. 2013). Practices that can enhance herbicide degradation include tilling the soil after harvest and establishing drainage systems during the winter months.

Soil moisture during the winter determines the effect of the herbicide on winter cover crops such as ryegrass (Avila et al. 2010a) and the extent of herbicide carryover. Drainage is crucial for establishing winter cover crops and can be an effective strategy to reduce herbicide carryover. Drainage improves soil aeration, while cover crops stimulate microbial activity, which in turn increases herbicide degradation. In areas with a high concentration of IMI herbicides in the soil, Ulguim et al. (2019a) observed that ryegrass and white clover reduced herbicide carryover to soybeans. This shows the phytoremediation ability of some plant species (Souto et al. 2013) and the stimulation of soil microbial populations that metabolize herbicides (Souto et al. 2020).

Soil pH also affects the persistence of IMI herbicides. Low soil pH increases sorption of IMI herbicide, thereby increasing persistence (Su et al. 2019). Lowland soils, where rice is grown in RS, are naturally acidic (Boeni et al. 2010). Although soil pH increases after flooding, this happens together with a lower redox potential in the soil that reduces aerobic microbial activity, and the pH returns to normal levels as soon as the field is drained for harvesting. Best practices, therefore, include the application of lime to raise soil pH. Other hypotheses pertaining to herbicide carryover include the reduced ability of plants to overcome herbicide injuries in less fertile soils (which has been demonstrated in some instances) and herbicide concentrating in the root zone in shallow soils (Bundt et al. 2015a).

Depending on the relative contributions of the processes described, residual IMI herbicides may injure sensitive crops. In RS, injury to non-tolerant crops planted after CL rice has been observed, including injury to cover crops such as ryegrass (Avila et al. 2010a; Bundt et al. 2015a). Residual IMI herbicide effect on non-tolerant rice planted 1 yr after CL rice is manifested in reduced grain yield between 30% (Avila et al. 2010b) and 55% (Marchesan et al. 2010). *Sorghum bicolor* (L.) Moench is one of the most sensitive crops, with a reduction in aboveground dry mass of up to 94% (Souza et al. 2016). Residual IMI herbicides in soil reduced soybean shoot dry mass and leaf area by about 50% (Fraga et al. 2019).

It is important to use integrated crop management to avoid, or at least reduce, herbicide carryover in lowland areas. This management should include crop rotation, preferably soybean, which is most tolerant to IMI carryover. Furthermore, producers should plant a cover crop during the winter, correct the soil pH, and drain the field during the winter. For a detailed discussion of this topic, refer to Gehrke et al. (2021).

Complementary Methods for Weedy Rice Control

Producers have abandoned complementary weed control practices such as roguing, wick bar, and mechanical weeding, due to the

availability of CL rice (Agostinetto et al. 2001). The abandonment of complementary weedy rice control methods is due to several factors, including the low availability and increasing cost for hand weeding.

Before the CL system was introduced, the producers had no other option than to use every tool they had available for weedy rice control; after the CL system was introduced, rice producers began to rely only on herbicides for weedy rice control. Powles and Gaines (2016) called this “herbicide-only syndrome,” and this is the main factor driving producers’ failure to use complementary weed control methods.

Without a complementary control method, the remaining uncontrolled plants have resulted in outcrossing with CL rice and the fast evolution of IMI-resistant weedy rice populations. We have learned that complementary methods for weedy rice control are vital to improving the efficacy and longevity of the CL rice system.

Special Case: CL Rice in the Pre-germinated Rice System

Rice producers in southern Brazil, mainly in SC and part of RS, have adopted the water-seeded system with pre-germinated seeds to suppress weedy rice infestation. However, the selection pressure from decades of monoculture rice cultivation has also resulted in weedy rice plants that can now germinate and emerge under the water, even with continuous flooding after seeding (Kaspary et al. 2020). Rice growers are in urgent need of new technologies to manage weedy rice.

Since the first CL rice cultivar for water-seeded rice was released by Epagri, growers have focused on how to manage the CL rice crop (Eberhardt et al. 2007). Recommendations include rotation of the establishment systems, especially for producers intending to plant water-seeded rice in fields with high infestations of weedy rice, and to avoid ratoon from CL cultivars (Eberhardt et al. 2007). However, most rice growers are unable to change their planting systems due to the soil type or climatic conditions being unfavorable for crop rotations. Producers have also avoided planting conventional cultivars after CL rice due to herbicide carryover and others have cultivated ratoon rice. As a result, the majority of the rice-growing areas in SC have been heavily infested with a diverse weedy rice population resistant to IMI herbicides, including plants that mimic commercial cultivars. Semi-dwarf weedy rice plants with long grains have become a major concern in certified seed production areas, because they are difficult to identify in the field. Although researchers, extension agents, and consultants have repeatedly advised growers in SC to rotate CL and conventional cultivars and to avoid ratooning the CL cultivars, most growers have not followed these recommendations (Eberhardt et al. 2007, 2015). The result is that any surviving, uncontrolled weedy rice plants flower simultaneously with the CL rice varieties, resulting in gene flow. Although, herbicide carryover is a primary concern in RS (Pinto et al. 2009a, 2009b, 2011), it is not a problem in water-seeded systems in SC.

Over the past 18 yr, we have learned the difficulties of teaching producers about best management practices and effective increased adoption rates of such practice. We still need to identify strategies that researchers and extension agents can use to communicate more effectively with growers. Knowledge we have gained concerning weedy rice management in the CL system in Brazil (Figure 3) could be applied in other countries where this system is just beginning to be developed and commercialized.



Figure 3. Pragmatic recommendations for sustainable use of the Clearfield™ (CL) rice production system.

Conclusions

CL rice or any other weed management technology cannot be used as a “silver bullet” but must instead be part of an integrated weed management strategy. This is especially true for weedy rice, as this weed can outcross with HR rice cultivars and quickly generate herbicide-resistant weedy populations. The CL system must not be used in areas without a weedy rice infestation.

One of the most important aspects of a weed management strategy like the CL system is to train producers. This requires close cooperation between researchers, extension agents, and the industry to develop a coordinated message and promote a broader

understanding of the system. We also need to train extension agents and producers more effectively on the optimal management of this technology to extend its period of usefulness.

Before launching any technology with resistant varieties to control weedy rice, it is mandatory to establish a meticulous rice seed production system free of weedy rice and to educate the growers on the importance of planting certified seeds for the sustainable production of rice.

To avoid IMI herbicide carryover to non-tolerant crops, including non-tolerant rice, producers should adopt the following practices: (1) do not use IMI herbicides for more than two consecutive growing seasons; (2) use the recommended label rate of herbicides;

(3) drain the field before harvest, if possible, till and level the field, and drain the field during the winter; (4) plant cover crops to enhance herbicide phytodegradation; (5) lime the soil to raise its pH to enhance herbicide availability and degradation in the soil; (6) practice intermittent flooding, if possible, in the last season of CL rice production; (7) plant the least IMI-susceptible non-CL rice variety; and (8) rotate rice with other crops.

Adopting integrated weed management practices is essential to avoid the evolution of resistance in weedy rice and other weeds. These practices reduce selection pressures by rotating herbicides with different MOAs, applying herbicides sequentially, using herbicides with lower selection pressure, using mixtures of herbicides with different MOAs, and performing site-specific management with direct application of herbicides to surviving plants before the flowering stage. The occurrence of resistant populations can be further minimized by adopting practices such as crop rotation, rotation of control methods, rotation of cultivation methods, monitoring weed shifts, and preventing suspect plants from producing seeds.

New technologies for weedy rice control will likely become available in the near future. Based on what we have learned over the last 18 yr with the CL system in Brazil, we can apply these tools more wisely while continuing to use CL rice as a tool for weedy rice management.

Acknowledgments. We are grateful to CNPq for research fellowships granted to several authors, as follows: LAdA, N.Proc. 310538/2015-7 CNPq; AM, Proc. 303379/2019-7; EM, Proc. N. 305230/2018-2 CNPq; and DA, Proc. 308363/2018-3. We also thank Jesus Juarez de Oliveira Pinto for his work during his career, which helped us to understand the fate of IMI herbicides in rice fields and how to manage the CL system. We thank Alcido Elenor Wander for building the map showing rice production in Brazil (Figure 1). The authors declare that most of them have received partial funding to conduct experiments from BASF. We also declare that this funding was only used in research activities and did not affect the authors' ability to write this review, which was strictly based on the science behind the technology.

References

- Agostinetto D, Fleck NG, Rizzardi MA, Merotto Jr A, Vidal RA (2001) Red rice: ecophysiology and strategies of control. *Cienc Rural* 31:341–349. Original in Portuguese
- Agostinetto D, Magro TD, Vargas L, Noldin JA (2011) *Cyperus difformis* L. resistance to pyrazosulfuron-ethyl herbicide and option control. *Semin Ciencias Agrar* 32:839–848
- Alister C, Kogan M (2005) Efficacy of imidazolinone herbicides applied to imidazolinone-resistant maize and their carryover effect on rotational crops. *Crop Prot* 24:375–379
- Andres A, Concenço G, Theisen G, Galon L, Tesio F (2012) Management of red rice (*Oryza sativa*) and barnyardgrass (*Echinochloa crus-galli*) grown with sorghum with reduced rate of atrazine and mechanical methods. *Exp Agric* 48:587–596
- Avila LA de, Andres A, Marchesan E, Menezes VG (2000a) Red rice seed bank under seeding systems of flooded rice. *Cienc Rural* 30:773–777. Original in Portuguese
- Avila LA, Marchesan E, Machado SL de O, Silva RP da (2000b) Red rice seed bank evolution under different systems of lowland utilization. *Planta Daninha* 18:217–230. Original in Portuguese
- Avila LA, Marchezan M, François T, Cezimbra DM, Souto KM, Refatti JP (2010a) Injury caused by the formulated mixture of the herbicide imazethapyr and imazapic in ryegrass as affected by soil moisture. *Planta Daninha* 28:1041–1046. Original in Portuguese
- Avila LA, Noldin JA, Mariot CHP, Massoni PFS, Fipke M V., Gehrke VR, Merotto A, Tomita FM, Matos AB, Facioni G, Vieira EB, Rosa ES, Santos RP, Camargo ER, Theisen G, Roma-Burgos N (2021) Status of weedy rice (*Oryza sativa* L.) infestation and management practices in southern Brazil. *Weed Sci*, doi: 10.1017/wsc.2021.24
- Avila LA, Teló GM, Ferreira RB, Marchesan E, Machado SLO, Rossato TL, Cezimbra DM, Rigão G (2010b) Non-tolerant rice production after using the Clearfield™ rice system. *Planta Daninha* 28:123–129. Original in Portuguese
- BASF (2020) Clearfield™ Rice System: The Main Rice Production System. <https://agriculture.basf.com/br/pt/protacao-de-cultivos-e-sementes/produtos/clearfield.html>. Accessed: July 7, 2020. Original in Portuguese
- Boeni M, Anghinoni I, Junior S, Filho B (2010) Soil Fertility Evolution of Rice Cultivated Soils of the Rio Grande do Sul. Cachoeirinha, RS, Brazil: IRGA. 40 p. Original in Portuguese
- Bohnen H, Da Silva LS, Macedo VRM, Marcolin E (2005) Organic acids in the soil solution of a gley soil cropped with lowland rice under different systems. *Rev Bras Cienc do Solo* 29:475–480. Original in Portuguese
- Botta GF, Tolón-becerra A, Lastra-bravo X, Hidalgo R, Rivero D, Agnes D (2015) Alternatives for handling rice (*Oryza sativa* L.) straw to favor its decomposition in direct sowing systems and their incidence on soil compaction. *Geoderma* 239–240:213–222
- Bundt A da C, Avila LA de, Agostinetto D, Nohatto MA, Souto KM, Senseman SA (2010) Depth of placement of the herbicide imazethapyr + imazapic in soil profile on non-tolerant rice injury. *Cienc Rural* 40:1867–1873. Original in Portuguese
- Bundt A da C, Avila LA de, Pinto JJ de O, Santos TT dos, Agostinetto D, Martins K (2013) Upward movement of the formulated mixture of imazethapyr and imazapic in response to height of the water table. *Cienc Rural* 43:1597–1604. Original in Portuguese
- Bundt ADC, Avila LA, Agostinetto D, Nohatto MA, Vargas HC (2015a) Carryover of imazethapyr + imazapic on ryegrass and non-tolerant rice as affected by thickness of soil profile. *Planta Daninha* 33:357–364
- Bundt ADC, Avila LAA, Pivetta A, Agostinetto D, Dick DPP, Bureau P (2015b) Persistence of imidazolinones in soils under a Clearfield™ system of rice cultivation. *Planta Daninha* 33:341–349
- Burgos NR, Norsworthy JK, Scott RC, Smith KL (2008) Red rice (*Oryza sativa*) status after 5 years of imidazolinone-resistant rice technology in Arkansas. *Weed Technol* 22:200–208
- Busi R, Nguyen NK, Chauhan BS, Vidotto F, Tabacchi M, Powles SB (2017) Can herbicide safeners allow selective control of weedy rice infesting rice crops? *Pest Manag Sci* 73:71–77
- Carvalho PC de F, Anghinoni I, Moraes A, Souza ED, Sulc RM, Lang CR, Flores JPC, Terra Lopes ML, da Silva JLS, Conte O, de Lima Wesp C, Levien R, Fontaneli RS, Bayer C (2010) Managing grazing animals to achieve nutrient cycling and soil improvement in no-till integrated systems. *Nutr Cycl Agroecosyst* 88:259–273
- [CONAB] Companhia Nacional do Abastecimento (2020) Mapping of paddy rice production in Brazil—CONAB. <https://www.conab.gov.br/institucional/publicacoes/outras-publicacoes/item/14089-mapeamento-do-arroz-irrigado-no-brasil-2020>. Accessed: September 21, 2020. Original in Portuguese
- Correia S da L, da Silva PRF, Boeni M, Bredemeier C, Anghinoni I, Menegati GB, Maass MB, Miozzo LC (2018) Performance of flooded rice grown in succession to winter cover crops. *Rev Bras Cienc do Solo*:e0160461
- Counce PA, Keisling TC, Mitchell AJ (2000) A uniform, objective, and adaptive system for expressing rice development. *Crop Sci* 40:436–443
- Croughan T, Utomo H, Asnders D, Braverman M (1996) Herbicide resistant rice offers potential solution to red rice problem. *Louisiana Agric* 39:10–12
- Dal Magro T, Agostinetto D, Pinto JJO, Galon L, Rezende AL (2006) Simulated drift effects of an ALS inhibitor herbicide on yield components of flooded rice. *Planta Daninha* 24:805–812. Original in Portuguese
- Dalazen G, Markus C, Merotto A Jr (2018) Differential expression of genes associated with degradation enhancement of imazethapyr in barnyardgrass (*Echinochloa crus-galli*). *J Agric Sci* 10:389–401
- Denardin LG de O, Martins AP, Carmona F de C, Veloso MG, Carmona GI, Carvalho PC de F, Anghinoni I (2020) Integrated crop–livestock systems in paddy fields: new strategies for flooded rice nutrition. *Agron J* 112:2219–2229
- Drescher MS, Reinert DJ, Denardin JE, Gubiani PI, Faganello A, Silva BR da, Zardin MC (2017) Fertilizer shanks to promote soil decompaction in the seeding operation. *Cienc Rural* 47:e20160026

- Eberhardt DS, Noldin JA, Schiocchet MA, Bacha RE, Knoblauch R, Marschalek R, Vieira J, Leites A, Jezus NB de (2007) Clearfield™ Production System for Pre-germinated Rice. Itajaí, Brazil: EPAGRI. 14 p. Original in Portuguese
- Eberhardt DS, Noldin JA, Schiocchet MA, Knoblauch R, Marschalek R, Leites A, Guma JM, Wickert E, Andrade A de, Scheuermann KK, Hickel ER (2015) Clearfield™ Production System for Pre-germinated and Dry Seeded Rice. Itajaí, Brazil: EPAGRI. 20 p. Original in Portuguese
- Eberhardt DS, Oliveira Neto AM, Noldin JA, Vanti RM (2016) Barnyardgrass with multiple resistance to synthetic auxin, ALS and Accase inhibitors. *Planta Daninha* 34:823–832
- Flint JL, Witt WW (1997) Microbial degradation of imazaquin and imazethapyr. *Weed Sci* 45:586–591
- Fraga DS, Agostinetto D, Langaro AC, Oliveira C, Ulguim AR, Silva JDG (2019) Morphological and metabolic changes in soybean plants cultivated in irrigated rice rotation and as affected by imazapyr and imazapic herbicides carryover. *Planta Daninha* 37:1–10. Original in Portuguese
- Fruet B de L, Merotto A, Ulguim A da R (2020) Survey of rice weed management and public and private consultant characteristics in southern Brazil. *Weed Technol* 34:351–356
- Gealy DR, Mitten DH, Rutger JN (2003) Gene flow between red rice (*Oryza sativa*) and herbicide-resistant rice (*O. sativa*): implications for weed management. *Weed Technol* 17:627–645
- Gehrke VR, Fipke MV, Avila LA de, Camargo ER (2021) Understanding the opportunities to mitigate carryover of imidazolinone herbicides in lowland rice. *Agriculture* 11:299
- Giacomeli R, Marchesan E, Sartori GMS, Donato G, da Silva PRF, Kaiser DR, Aramburu EBB (2016) Deep tillage and furrow opener seeders for corn cropping in Planosols. *Pesqui Agropecu Bras* 51:261–270. Original in Portuguese
- Gomes AS, Petrini JA, Fagundes PRR (2004) Racional rice crop management: “Programa Marca”. Pelotas, Brazil: EMBRAPA. 203 p. Original in Portuguese
- Goulart ICGR, Borba TCO, Menezes VG, Merotto A (2014) Distribution of weedy red rice (*Oryza sativa*) resistant to imidazolinone herbicides and its relationship to rice cultivars and wild *Oryza* species. *Weed Sci* 62:280–293
- Goulart ICGR, Matzenbacher FO, Merotto A (2012) Differential germination pattern of rice cultivars resistant to imidazolinone herbicides carrying different acetolactate synthase gene mutations. *Weed Res* 52:224–232
- Heap I (2021) The International Herbicide-Resistant Weed Database. www.weedscience.org. Accessed: December 2, 2020
- Helgueira DB, Rosa TD, Moura DS, Galon L, Pinto JJO (2019) Leaching of imidazolinones in irrigation systems in rice cultivation: sprinkling and flooding. *Planta Daninha* 37:e019179877
- Henry CG, Sartori GMS, Gaspar JP, Marchesan E, Hirsh SM, Horton AP, Espinoza L, James H (2018) Deep tillage and gypsum amendments on fully, deficit irrigated, and dryland Soybean. *Agron J* 110:737–748
- Hicks HL, Comont D, Coutts SR, Crook L, Hull R, Norris K, Neve P, Childs DZ, Freckleton RP (2018) The factors driving evolved herbicide resistance at a national scale. *Nat Ecol Evol* 2:529–536
- Kalsing A, dos Reis Goulart ICG, Mariot CHP, Menezes VG, de Oliveira Matzenbacher F, Merotto A (2019) Spatial and temporal evolution of imidazolinone-resistant red rice in “Clearfield” rice cultivations. *Pesqui Agropecu Bras* 54:e00215
- Kaspary TE, Roma-Burgos N, Merotto A (2020) Snorkeling strategy: tolerance to flooding in rice and potential application for weed management. *Genes (Basel)* 11:975
- Kraemer AF, Marchesan E, Avila LA, Machado SLO, Grohs M (2009) Environmental fate of imidazolinone herbicides—a review. *Planta Daninha* 27:629–639. Original in Portuguese
- Krausz RF, Kapusta G (1998) Total postemergence weed control in imidazolinone-resistant corn (*Zea mays*). *Weed Technol* 12:151–156
- Langevin SA, Clay K, Grace JB (1990) The incidence and effects of hybridization between cultivated rice and its related weed red rice (*Oryza sativa* L.). *Evolution (NY)* 44:1000–1008
- Lopes MCB, Rosso AF de, Lopes SIG (2003) A new cultivar developed by IRGA for the Clearfield™ system. Pages 3–5 in *Proceedings of the Brazilian Rice Congress*. Itajaí, Brazil: EPAGRI. Original in Portuguese
- Magalhães AM Jr, Fagundes PRR, Franco D, Andres A, Rangel PH, Moraes OP, Moura Neto FM, Severo A (2011) BRS Sinuelo CL: Rice Cultivar for the Clearfield™ System. Pelotas, Brazil: EMBRAPA. 8 p
- Marchesan E, Dos Santos FM, Grohs M, De Avila LA, Machado SLO, Senseman SA, Massoni PFS, Sartori GMS (2010) Carryover of imazethapyr and imazapic to nontolerant rice. *Weed Technol* 24:6–10
- Marchesan E, Massoni PFS, Grohs M, Villa SCC, Avila LA, Roso R (2011a) Imidazolinone-tolerant rice: red rice seed bank and gene flow. *Planta Daninha* 29:1099–1105. Original in Portuguese
- Marchesan E, Massoni PFS, Villa SCC, Grohs M, de Avila LA, Sartori GMS, Bruck RF (2011b) Productivity, injury and control of red rice in succession of irrigated rice cultivation in system Clearfield. *Cienc Rural* 41:17–24. Original in Portuguese
- Marchesan E, Menezes NL de, Siqueira C do A (2001) Control of quality of the flooded rice seeds in Santa Maria/RS. *Cienc Rural* 31:375–379. Original in Portuguese
- Marchesan E, Oliveira APBB, Avila LA, Bundt ALP (2003) Red rice seed bank dynamics affected by cattle trampling and fallow duration. *Planta Daninha* 21:55–62. Original in Portuguese
- Martini LFD, Mezzomo RF, Avila LA de, Massey JH, Marchesan E, Zanella R, Peixoto SC, Refatti JP, Cassol GV, Marques M (2013) Imazethapyr and imazapic runoff under continuous and intermittent irrigation of paddy rice. *Agric Water Manag* 125:26–34
- Martins GN, Noldin JA, Lucietti D, Oliveira DG, Haverroth HS, Souza LV, Fernandes RH (2017) Rate of use and quality of rice seeds used in Santa Catarina, Brazil. Pages 187–190 in *Rio Grande do Sul Rice Institute, ed. Proceedings of the Southern Brazilian Rice Congress*. Cachoeirinha, RS, Brazil: IRGA. Original in Portuguese
- Massoni PFS, Marchesan E, Grohs M, Roso R, Coelho LL, MacHado SLO, Teló GM, Dal’Col Lúcio A (2013) Influence of post-harvest management of rice crop on red rice seed bank. *Planta Daninha* 31:89–98. Original in Portuguese
- Matzenbacher FO, Bortoly ED, Kalsing A, Merotto A (2015) Distribution and analysis of the mechanisms of resistance of barnyardgrass (*Echinochloa crus-galli*) to imidazolinone and quinclorac herbicides. *J Agric Sci* 153:1044–1058
- Menezes VG (2003) Clearfield™ rice system, a new alternative for red rice control. Pages 824–826 in *Proceedings of the Brazilian Rice Congress*. Balneário Camboriu, SC, Brazil: SOSBAI. Original in Portuguese
- Menezes VG, Macedo VRM, Anahinoni I (2004) Projeto 10—Crop Strategy to Enhance Yield, Competitiveness and Sustainability of Rice Farms in RS. Porto Alegre, Brazil: IRGA. 32 p
- Menezes VG, Mariot CHP, Kalsing A, Goulart ICGR (2009) Red rice (*Oryza sativa*) resistant to the herbicides imidazolinones. *Planta Daninha* 27:1047–1052. Original in Portuguese
- Menezes VG, Mariot CHP, Lopes MCB, Silva PRF da, Teichmann LL, Da Silva PRF, Teichmann LL, Silva PRF da, Teichmann LL, Da Silva PRF, Teichmann LL (2001) No-tillage system of flooded rice genotypes in succession to winter cover crops. *Pesqui Agropecu Bras* 36:1107–1115. Original in Portuguese
- Merotto Jr A, Goulart ICGR, Nunes AL, Kalsing A, Markus C, Menezes VG, Wander AE (2016) Evolutionary and social consequences of introgression of nontransgenic herbicide resistance from rice to weedy rice in Brazil. *Evol Appl* 9:837–846
- [MAPA] Ministry of Agriculture—Brazil (2005) Normative Instruction MAPA IN 24/2015, 16/12/2005. Establish patterns of identity and quality for production and commercialization of seeds and seedlings. <https://www.defesa.agricultura.sp.gov.br/legislacoes/instrucao-normativa-24-de-16-12-2005,725.html>. Original in Portuguese. Accessed: June 26, 2020
- [MAPA] Ministry of Agriculture—Brazil (2013) Normative Instruction MAPA 45/2013-18/09/2013. Establish patterns of identity and quality for production and commercialization of seeds and seedlings. https://www.gov.br/agricultura/pt-br/assuntos/insumos-agropecuarios/insumos-agricolas/sementes-e-mudas/publicacoes-sementes-e-mudas/copy_of_INN45de17-desetembrede2013.pdf. Original in Portuguese. Accessed: June 26, 2020
- [MAPA] Ministry of Agriculture—Brazil (2020). National registration of cultivars. https://sistemas.agricultura.gov.br/snpc/cultivarweb/cultivares_registradas.php Accessed: June 26, 2020
- Moraes P V.D., Agostinetto D, Vignolo GK, Santos LS, Panozzo LE (2009) Cover crop management and weed control in corn. *Planta Daninha* 27:289–296. Original in Portuguese

- Noldin JA, Chandler JM, McCauley GN (2006) Seed longevity of red rice ecotypes buried in soil. *Planta Daninha* 24:611–620
- Noldin JA, Knoblauch R, Dal Piva CA, Alfonso-Morel D (1997) Rice seed quality in Santa Catarina. Pages 487–490 in *Proceedings of the Brazilian Rice Congress*. Itajaí, SC, Brazil: Sociedade Brasileira do Arroz Irrigado. Original in Portuguese
- Noldin JA, Yokoyama S, Antunes P, Luzzardi R (2002) Outcrossing potential of glufosinate-resistant rice to red rice. *Planta Daninha* 20:243–251. Original in Portuguese
- Norsworthy JK, Bond J, Scott RC (2013) Weed management practices and needs in Arkansas and Mississippi rice. *Weed Technol* 27:623–630
- Norsworthy JK, Ward SM, Shaw DR, Llewellyn RS, Nichols RL, Webster TM, Bradley KW, Frisvold G, Powles SB, Burgos NR, Witt WW, Barrett M (2012) Reducing the risks of herbicide resistance: best management practices and recommendations. *Weed Sci* 60:31–61
- Pellerin KJ, Webster EP, Zhang W, Blouin DC (2003) Herbicide mixtures in water-seeded imidazolinone-resistant rice (*Oryza sativa*). *Weed Technol* 17:836–841
- Pellerin KJ, Webster EP, Zhang W, Blouin DC (2004) Potential use of imazethapyr mixtures in drill-seeded imidazolinone-resistant rice. *Weed Technol* 18:1037–1042
- Peske ST (2021) Rice seed system in Brazil. *Seed News* 21:18–22. Original in Portuguese
- Pinto JJO, Noldin JA, Pinho CF, Rossi F, Galon L, Almeida GF (2009a) Field persistence of (imazethapyr+imazapic) to grain sorghum (*Sorghum bicolor*) planted in rotation after irrigated rice. *Planta Daninha* 27:1015–1024
- Pinto JJO, Noldin JA, Rosenthal MD, Pinho CF, Rossi F, Machado A, Piveta L, Galon L (2009b) Residual activity of (imazethapyr+imazapic) on ryegrass (*Lolium multiflorum*), following Clearfield™ rice. *Planta Daninha* 27:609–619. Original in Portuguese
- Pinto JJO, Noldin JA, Sousa CP, Agostinetto D, Piveta L, Donida A (2011) Residual soil activity of imazethapyr + imazapic to rice planted in rotation after Clearfield rice. *Planta Daninha* 29:205–216. Original in Portuguese
- Piveta LB, Noldin JA, Roma-Burgos N, Viana VE, Benedetti L, Pinto JJO, Lamego FP, Avila LA de (2021) Weedy rice diversity in southern Brazil. *Weed Sci*, doi: 10.1017/wsc.2021.23
- Powles SB, Gaines TA (2016) Exploring the potential for a regulatory change to encourage diversity in herbicide use. *Weed Sci* 64:649–654
- Qiu J, Jia L, Wu D, Weng X, Chen L, Sun J, Chen M, Mao L, Jiang B, Ye C, Turra GM, Guo L, Ye G, Zhu QH, Imaizumi T, Song BK, Scarabel L, Merotto A, Olsen KM, Fan L (2020) Diverse genetic mechanisms underlie worldwide convergent rice fertilization. *Genome Biol* 21:70
- [IRGA] Rio Grandense Rice Research Institute (2020) Home page. <https://irga.rs.gov.br/inicial>. Accessed: June 26, 2020. Original in Portuguese
- Roso AC, Merotto Jr A, Delatorre CA (2010a) Bioassays for diagnosis of resistance to the herbicides imidazolinones in rice plants. *Planta Daninha* 28:411–419. Original in Portuguese
- Roso AC, Merotto Jr A, Delatorre CA, Menezes VG (2010b) Regional scale distribution of imidazolinone herbicide-resistant alleles in red rice (*Oryza sativa* L.) determined through SNP markers. *Field Crop Res* 9:175–182
- Rouse CE, Roma-Burgos N, Norsworthy JK, Tseng TM, Starkey CE, Scott RC (2018) *Echinochloa* resistance to herbicides continues to increase in Arkansas rice fields. *Weed Technol* 32:34–44
- Santos FM, Marchesan E, Machado SLO, Villa SCC, Avila LA, Massoni PFS (2007) Controle químico de arroz-vermelho na cultura do arroz irrigado. *Planta Daninha* 25:405–412
- Sartori GMS, Marchesan E, Azevedo CF, Coelho LL, De Oliveira ML (2014) Germination of irrigated rice and red rice biotypes under different temperatures. *Rev Cienc Agron* 45:319–326. Original in Portuguese
- Schaedler CE, Moraes PVD, Noldin JA (2008) Barnyardgrass resistance to quinclorac in Santa Catarina. *Agropecuária Catarinense* 21:68–72. Original in Portuguese
- Shiocchet MA, Yokoyama S, Eberhardt DS, Noldin JA, Knoblauch R, Marchalek R, Peruch LAM, Bacha RE, Ishiy T, Alfonso-Morel D, Vieira J, Prado HF, Miura L (2007) SCS 115 CL: Paddy Rice Cultivar. Florianópolis, SC, Brazil: EPAGRI. 2 p. Original in Portuguese
- Shaner DL (2014) *Herbicide Handbook*. 10th ed. Champaign, IL: Weed Science Society of America. 213 p
- Silveira LP, Feijó AR, Benetti C, Refatti JP, Fipke M V., Camargo ER, Ziska LH, Avila LA (2019) Elevated CO2 concentrations and water stress affect the ability of Italian ryegrass to remediate herbicides and enhance its allelopathic effect. *Planta Daninha* 37:e019179823
- Singh V, Burgos N, Singh S, Abugho S, Earnest L, Gbur E, Scott R (2016) Herbicide and winter flood treatments for controlling volunteer rice off-season. *Crop Prot* 79:87–96
- Singh V, Burgos NR, Singh S, Gealy DR, Gbur EE, Caicedo AL (2017) Impact of volunteer rice infestation on yield and grain quality of rice. *Pest Manag Sci* 73:604–615
- SOSBAI (2018) Rice Technical Recommendation from Research in Southern Brazil. Embrapa Clima Temperado. Pelotas, Brazil: EMBRAPA. 205 p. Original in Portuguese
- Souto KM, Jacques RJS, de Avila LA, Machado SL de O, Zanella R, Refatti JP (2013) Biodegradation of the herbicides imazethapyr and imazapic in rhizosphere soil of six plant species. *Cienc Rural* 43:1790–1796. Original in Portuguese
- Souto KM, Jacques RJS, Zanella R, Machado SL de O, Balbinot A, Avila LA de (2020) Phytostimulation of lowland soil contaminated with imidazolinone herbicides. *Int J Phytoremediation* 22:774–780
- Souza MF, Neto MDC, Marinho MI, Saraiva DT, Faria AT, Silva AA, Silva DV (2016) Persistence of imidazolinones in soils under a Clearfield system of rice cultivation. *Planta Daninha* 34:589–596
- Spatt LL, Dornelles SHB, Sanchotene DM, Brum AB, Carloto BW, Scherer MB (2016) Low-level resistance of *Cyperus iria* L. to ALS-inhibiting herbicides occurring in the State of Rio Grande do Sul. *Científica* 44:532
- Stępkowski T, Banasiewicz J, Granada CE, Andrews M, Passaglia LMP (2018) Phylogeny and phylogeography of rhizobial symbionts nodulating legumes of the tribe genisteae. *Genes (Basel)* 163:2–25
- Su W, Hao H, Ding M, Wu R, Xu H, Xue F, Shen C, Sun L, Lu C (2019) Adsorption and degradation of imazapic in soils under different environmental conditions. *PLoS ONE* 14:e0219462
- Sudianto E, Beng-Kah S, Ting-Xiang N, Saldain NE, Scott RC, Burgos NR (2014) Clearfield® rice: its development, success, and key challenges on a global perspective. *Crop Prot* 49:40–51
- Ulguim AR, Agostinetto D, Vargas L, Da Silva JDG, Da Silva BM, Da Rosa Westendorff N (2017) Agronomic factors involved in low-level wild poinsettia resistance to glyphosate. *Rev Bras Ciencias Agrar* 12:51–59
- Ulguim AR, Carlos FS, Santos RAS, Zanon AJ, Werle IS, Beck M (2018) Weed phytosociological in irrigated rice under different cultivation systems and crop rotation intensity. *Cienc Rural* 48:e20180230
- Ulguim AR, Carlos FS, Zanon AJ, Ogoshi C, Bexaira KP, Silva PRF (2019a) Is increasing doses of imazapyr + imazapic detrimental to the main crop rotation alternatives to flooded rice? *Planta Daninha* 37:e019217913
- Ulguim AR, Fruet BL, Merotto A, Silva AL (2021) Status of weed control in imidazolinone-herbicide resistant rice in Rio Grande do Sul. *Adv Weed Sci* 39
- Ulguim AR, Silva BM, Agostinetto D, Avila Neto RC, Zandoná RR (2019b) Resistance mapping of the genus *Cyperus* in Rio Grande do Sul and selection pressure analysis. *Planta Daninha* 37:e019186679
- Vedelago A, Carmona FC, Boeni M, Lange CE, Anghinoni I (2012) Soil Fertility and Fitness for Soybean Production in Rice Production Regions of RS State. Porto Alegre, Brazil: IRGA. 46 p. Original in Portuguese
- Viero JLC, Schaedler CE, de Azevedo EB, Dos Santos JVA, Scalcon R de M, de David DB, da Rosa FQ (2018) Endozoochorous dispersal of seeds of weedy rice (*Oryza sativa* L.) and barnyardgrass (*Echinochloa crus-galli* L.) by cattle. *Cienc Rural* 48:e20170650
- Villa SCC, Marchesan E, Avila LA, Massoni PFS, Telo GM, Machado SLO, Camargo ER (2006) Imidazolinone tolerant rice: red rice control, out-crossing and herbicide carryover to non-tolerant crops. *Planta Daninha* 24:761–768. Original in Portuguese
- Weinert C, Carlos F, Pacheco D, Sousa R, Bortowski E, Lages M (2019) Persian clover production in spring enhances rice yield. *Proceedings of the Latin American Soil Science Congress*. Montevideo, Uruguay, October 7–11, 2019. Original in Portuguese
- Ziska LH, Gealy DR, Burgos N, Caicedo AL, Gressel J, Lawton-Rauh AL, Avila LA, Theisen G, Norsworthy J, Ferrero A, Vidotto F, Johnson DE, Ferreira FG, Marchesan E, Menezes V, et al. (2015) Weedy (red) rice. An emerging constraint to global rice production. *Adv Agron* 129:181–228