

Assessing the diversity of whiteflies infesting cassava in Brazil

Cesar A.D. Xavier¹, Angélica Maria Nogueira¹, Vinicius Henrique Bello², Luís Fernando Maranhão Watanabe², Tarsiane Mara Carneiro Barbosa¹, Miguel Alves Júnior³, Leonardo Barbosa⁴, José E.A. Beserra-Júnior⁵, Alessandra Boari⁶, Renata Calegario⁷, Eduardo Silva Gorayeb⁸, Jaime Honorato Júnior⁹, Gabriel Koch⁷, Gaus Silvestre de Andrade Lima¹⁰, Cristian Lopes⁴, Raquel Neves de Mello¹¹, Késsia Pantoja⁶, Fábio Nascimento Silva⁸, Roberto Ramos Sobrinho¹⁰, Enilton Nascimento Santana¹², José Wilson Pereira da Silva¹³, Renate Krause-Sakate² and Francisco M. Zerbini¹

¹ Dep. de Fitopatologia/BIOAGRO, Universidade Federal de Viçosa, Viçosa, MG, Brazil

² Dep. de Proteção Vegetal, Universidade Estadual Paulista, Botucatu, SP, Brazil

³ Faculdade de Engenharia Agrônômica, Universidade Federal do Pará, Altamira, PA, Brazil

⁴ Instituto Federal do Sudeste de Minas Gerais, Rio Pomba, MG, Brazil

⁵ Dep. de Fitotecnia, Universidade Federal do Piauí, Teresina, PI, Brazil

⁶ Embrapa Amazônia Oriental, Belém, PA, Brazil

⁷ Dep. de Fitotecnia e Fitossanidade, Universidade Federal do Paraná, Curitiba, PR, Brazil

⁸ Centro de Ciências Agroveterinárias, Universidade do Estado de Santa Catarina, Lages, SC, Brazil

⁹ Centro Multidisciplinar do Campus de Barra, Universidade Federal do Oeste da Bahia, Barra, BA, Brazil

¹⁰ Centro de Ciências Agrárias/Fitossanidade, Universidade Federal de Alagoas, Rio Largo, AL, Brazil

¹¹ Embrapa Arroz e Feijão, Santo Antonio de Goiás, GO, Brazil

¹² Instituto Capixaba de Pesquisa e Extensão Rural, Linhares, ES, Brazil

¹³ Faculdade de Engenharia Florestal, Universidade Federal do Pará, Altamira, PA, Brazil

ABSTRACT

Background. The necessity of a competent vector for transmission is a primary ecological factor driving the host range expansion of plant arthropod-borne viruses, with vectors playing an essential role in disease emergence. Cassava begomoviruses severely constrain cassava production in Africa. Curiously, begomoviruses have never been reported in cassava in South America, the center of origin for this crop. It has been hypothesized that the absence of a competent vector in cassava is the reason why begomoviruses have not emerged in South America.

Methods. We performed a country-wide whitefly diversity study in cassava in Brazil. Adults and/or nymphs of whiteflies were collected from sixty-six cassava fields in the main agroecological zones of the country. A total of 1,385 individuals were genotyped based on mitochondrial cytochrome oxidase I sequences.

Results. A high species richness was observed, with five previously described species and two putative new ones. The prevalent species were *Tetraleurodes acaciae* and *Bemisia tuberculata*, representing over 75% of the analyzed individuals. Although we detected, for the first time, the presence of *Bemisia tabaci* Middle East-Asia Minor 1 (*BtMEAM1*) colonizing cassava in Brazil, it was not prevalent. The species composition varied across regions, with fields in the Northeast region showing a higher diversity. These results expand our knowledge of whitefly diversity in cassava and support the hypothesis that begomovirus epidemics have not occurred in cassava in Brazil due to the absence of

Submitted 4 February 2021

Accepted 17 June 2021

Published 15 July 2021

Corresponding authors

Renate Krause-Sakate,
renate.krause@unesp.br
Francisco M. Zerbini, zerbini@ufv.br

Academic editor

Joseph Gillespie

Additional Information and
Declarations can be found on
page 20

DOI 10.7717/peerj.11741

© Copyright

2021 Xavier et al.

Distributed under

Creative Commons CC-BY 4.0

OPEN ACCESS

competent vector populations. However, they indicate an ongoing adaptation process of *BtMEAM1* to cassava, increasing the likelihood of begomovirus emergence in this crop.

Subjects Agricultural Science, Biodiversity, Entomology, Virology, Zoology

Keywords Geminivirus, Virus vector, *Manihot esculenta*

INTRODUCTION

Cassava (*Manihot esculenta* Crantz) is a perennial shrub of the Euphorbiaceae family with great economic and social importance, especially in Africa, Asia, and Latin America. Currently, cassava is the third most important source of calories after rice and corn and is a staple food for more than one billion people living mainly in developing countries (Montagnac, Davis & Tanumihardjo, 2009). Although the botanical and geographical origin of *M. esculenta* is still debated, studies based on genetic markers and archaeological evidence suggest that domesticated cassava originated from the wild relative progenitor *M. esculenta* ssp. *flabellifolia* in the Amazon basin, with the domestication center located at the southern border of the Amazon in Brazil (Clement et al., 2016; Leotard et al., 2009; Olsen & Schaal, 1999; Watling et al., 2018). After its introduction in west Africa by Portuguese traders during the 16th century, cassava quickly disseminated throughout tropical Africa and Asia (Carter et al., 1997). Currently, the African continent is the world's biggest cassava producer, followed by Asia and South America (www.fao.org/faostat/en/#data/QC). Due to its high resilience to adverse environmental conditions, especially drought, high yield per unit of land and low level of management and inputs required during its life cycle, cassava is a suitable crop for poor and small farmers, partially ensuring food security in many African countries (Alves & Setter, 2004; El-Sharkawy, 2004; Gleadow, Pegg & Blomstedt, 2016).

Nevertheless, cassava may be affected by several pathogens and pests. Whiteflies (Hemiptera: Aleyrodidae) are one of the major constraints to its production in developing countries (Herrera-Campo, Hyman & Bellotti, 2011). Whiteflies comprise a diverse group of phloem-feeding insects, with more than 1,500 species assigned to 126 genera of which over 20 species have been reported to colonize cassava worldwide (Vasquez-Ordóñez et al., 2015). In addition to the direct damage due to feeding in the plant phloem, whiteflies cause indirect damage by deposition of honeydew, favoring the growth of sooty mold fungi on the leaf surface, and mainly by transmission of a broad range of viruses (Navas-Castillo, Fiallo-Olivé & Sánchez-Campos, 2011). Currently, species included in the genera *Aleurodicus*, *Aleurothrixus*, *Bemisia* and *Trialeurodes* have been shown to constitute effective vectors of plant viruses classified in five families (Chandrashekar et al., 2020; Maruthi et al., 2017; Navas-Castillo, Fiallo-Olivé & Sánchez-Campos, 2011; Njoroge et al., 2017). *Aleurodicus dispersus* and *Aleurothrixus trachoides* each transmit only one virus from the genera *Ipomovirus* and *Begomovirus*, respectively, while *Trialeurodes vaporariorum* and *T. abutilonea* transmit a few viruses included in the genera *Crinivirus* and *Torradovirus*. On the other hand, the *Bemisia tabaci* complex comprises one of the most important

group of plant virus vectors, transmitting over 450 viruses, the majority included in the genus *Begomovirus* (Navas-Castillo, Fiallo-Olivé & Sánchez-Campos, 2011; Zerbini et al., 2017) but including also viruses classified in the genera *Carlavirus*, *Crinivirus*, *Ipomovirus*, *Polerovirus* and *Torradorvirus* (Ghosh et al., 2019; Gilbertson et al., 2015; Navas-Castillo, Fiallo-Olivé & Sánchez-Campos, 2011; Whitfield, Falk & Rotenberg, 2015).

Over the last decade, advances in the use of molecular markers has led to a deep reappraisal of the taxonomic status of *B. tabaci* (De Barro et al., 2011; Dinsdale et al., 2010). Based on molecular phylogeny of the mitochondrial cytochrome oxidase I (mtCOI) gene, it has been proposed that *B. tabaci* consists of a complex of more than 40 cryptic (morphologically indistinguishable) species (De Barro et al., 2011; Dinsdale et al., 2010; Lee et al., 2013; Mugerwa et al., 2018; Vyskočilová, Seal & Colvin, 2019). Partial or complete reproductive isolation and biological and ecological differences among distinct species within the complex support the proposed classification (De Marchi et al., 2017; Gilbertson et al., 2015; Malka et al., 2018; Qin, Pan & Liu, 2016). The global dissemination of polyphagous and invasive species, such as *B. tabaci* Middle East-Asia Minor 1 (*Bt*MEAM1) and *B. tabaci* Mediterranean (*Bt*MED), have caused major changes in the epidemiology of crop-infecting begomoviruses such as *Tomato yellow leaf curl virus* (TYLCV), currently present in all the main tomato producing areas of the world (Lefeuvre et al., 2010; Mabvakure et al., 2016; Pan et al., 2012). In addition, the dissemination of polyphagous whiteflies has favored the transfer of indigenous begomoviruses from wild reservoir hosts to cultivated plants, as occurred in tomato crops in Brazil after the introduction of *Bt*MEAM1 in the mid-1990's (Ribeiro et al., 1998; Rocha et al., 2013).

Specific associations between endemic populations of *B. tabaci* and indigenous begomoviruses have also led to the emergence of severe epidemics in crops (Fauquet & Fargette, 1990; Pan et al., 2018). Cassava mosaic disease (CMD) is considered the most significant constraint to cassava production in Africa (Jacobson, Duffy & Sseruwagi, 2018); (Rey & Vanderschuren, 2017) and has been expanding to south and southeast Asia in recent years (Minato et al., 2019; Wang et al., 2019; Wang et al., 2016). CMD is caused by viruses of the genus *Begomovirus* (family *Geminiviridae*), which are transmitted in a circulative manner by whiteflies of the *Bemisia tabaci* cryptic species complex (Zerbini et al., 2017). Also, because cassava can be vegetatively propagated, transmission by infected root stock is important for the long-distance spread of CMD (Wang et al., 2020). To date, nine cassava mosaic begomoviruses (CMBs) have been reported in association with CMD, seven of them in Africa and two in the Indian subcontinent (Jacobson, Duffy & Sseruwagi, 2018; Legg et al., 2015; Patil & Fauquet, 2009). The emergence of CMD seems to have been the result of the transfer of indigenous begomoviruses from wild reservoir hosts to cassava, probably mediated by endemic populations of *B. tabaci* that have adapted to feed in cassava since its introduction from South America (Fauquet & Fargette, 1990; Legg & Fauquet, 2004). In Africa, CMD is transmitted primarily by *B. tabaci* Sub-Saharan Africa (SSA1-3, SSA6 and SSA9) and *B. tabaci* Indian Ocean (IO) (Jacobson, Duffy & Sseruwagi, 2018; Kunz et al., 2019). *B. tabaci* MEAM1 has been detected in some countries and is associated mostly with the transmission of begomoviruses in tomato (Jacobson, Duffy & Sseruwagi, 2018). Although wild reservoir hosts and a possible ancestral progenitor of current begomoviruses

causing CMD have not been found, the absence of cassava-infecting begomoviruses in the Americas supports an African origin for those viruses, and the presence of cassava-adapted *B. tabaci* species being restricted to Africa reinforces that hypothesis. The high species diversity and high level of molecular variation observed in viral populations causing CMD strongly suggests Africa as a diversification center for CMBs, with distinct CMBs recurrently emerging and evolving for a long time (De Bruyn et al., 2016; De Bruyn et al., 2012; Ndunguru et al., 2005; Tiendrebeogo et al., 2012).

Even if CMD in Africa is caused by indigenous viruses, the fact that cassava in South America has not been affected by begomoviruses is puzzling (Carabali, Belloti & Montoya-Lerma, 2010; Carabali, Montoya-Lerma & Bellotti, 2008; Patil & Fauquet, 2009). Carabali et al. (2005) suggested that the absence of cassava-infecting begomoviruses in the Americas would be due to lack of competent *B. tabaci* species that efficiently colonize cassava (Fauquet & Fargette, 1990; Legg & Fauquet, 2004). In Colombia, the inability of *B. tabaci* MEAM1 to colonize *M. esculenta* efficiently has been demonstrated under experimental conditions, reinforcing the above hypothesis (Carabali, Belloti & Montoya-Lerma, 2010). In addition, a recent study also from Colombia failed to detect any whitefly species of the *B. tabaci* complex in cassava (Gómez-Díaz, Montoya-Lerma & Muñoz Valencia, 2019).

Although whitefly diversity in Brazil has been surveyed extensively in recent years (Barbosa et al., 2015; Marubayashi et al., 2014; Marubayashi et al., 2013; Moraes et al., 2017; Moraes et al., 2018; Rocha et al., 2011), no study has been carried out specifically to explore the composition of whitefly communities colonizing cassava. Those studies carried out in other crops demonstrated that *B. tabaci* MEAM1 is the predominant species across Brazil in crops such as common bean, cotton, pepper, tomato and soybean. Furthermore, *B. tabaci* MED, which was recently introduced in Brazil, has quickly spread and currently is present in five states from the South and Southeast regions (Barbosa et al., 2015; Moraes et al., 2017; Moraes et al., 2018). A small number of whitefly samples from cassava were analyzed in those studies, with *B. tuberculata* and *Tetraleurodes acaciae* prevalent and detected exclusively in cassava (Marubayashi et al., 2014; Moraes et al., 2018). A large survey addressing whitefly diversity in cassava in its domestication center could provide clues to understand the absence of a CMD-like disease in the Americas. Moreover, this knowledge would be useful to anticipate the potential of emergence of begomoviruses in the crop and to help anticipate a management strategy.

Given this context, the objective of this work was to evaluate whitefly diversity in cassava across Brazil to infer about the absence of begomovirus occurrence in cassava. Our results demonstrated that the most prevalent species in cassava were *T. acaciae* and *B. tuberculata*. In addition, we detected for the first time the presence of *Bt*MEAM1 colonizing cassava in Brazil. The possible implications of these findings are discussed considering the absence of CMD and the potential for its emergence in cassava fields in Brazil.

MATERIALS & METHODS

Whitefly and cassava samples

Whiteflies were collected exclusively from cassava (*M. esculenta*) plants across 12 Brazilian states representative of the five macroregions (North, Northeast, Midwest, Southeast and

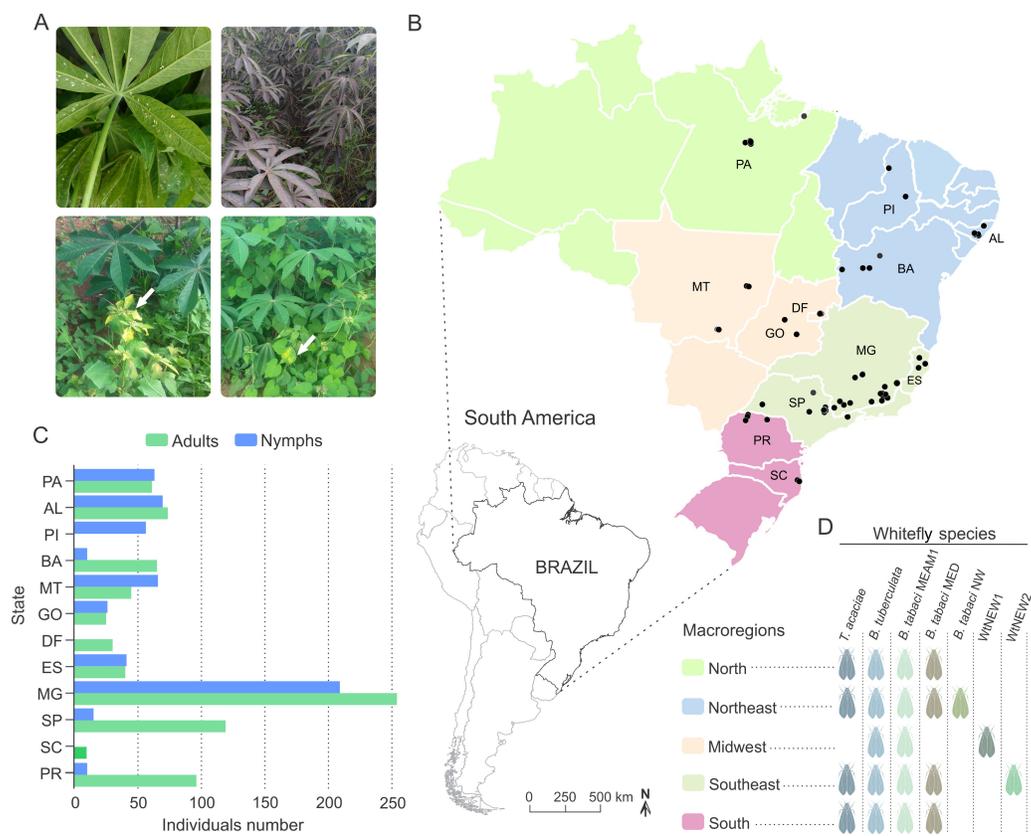


Figure 1 Overview of the survey locations and whitefly species detected. (A) Clockwise from top-left: adults and nymphs of *Bemisia tuberculata* colonizing cassava in Mogi Mirim, São Paulo state. Growth of sooty mould fungus on the leaf surface due to the deposition of honeydew by whiteflies. Presence of begomovirus-infected *Blainvillea rhomboidea* (family Asteraceae) in a cassava field in Minas Gerais state. Presence of begomovirus-infected *Euphorbia heterophylla* (family Euphorbiaceae) in a cassava field in Minas Gerais state. (B) Map of Brazil showing the locations where whiteflies samples were collected. The map is colored according to the regions as indicated in the legend. Black dots correspond to the sampled sites. Scale bar is only for Brazil map. (C) Number of adults and nymphs analyzed from each sampled site according to state. (D) Specie distribution according to region. AL, Alagoas; BA, Bahia; DF, Distrito Federal; ES, Espírito Santo; GO, Goiás; MG, Minas Gerais; MT, Mato Grosso; PA, Pará; Piauí; PR, Paraná; SC, Santa Catarina; SP, São Paulo.

Full-size DOI: [10.7717/peerj.11741/fig-1](https://doi.org/10.7717/peerj.11741/fig-1)

South; Fig. 1) between March 2016 and February 2019 (Table 1). To gather evidence of whether a given species was colonizing cassava, adults and nymphs from the same field were collected whenever possible (Table 1). Samples were obtained from commercial and non-commercial (subsistence) crops. Whitefly adults were sampled using a hand-held aspirator and nymphs were collected with the aid of a needle. Insects were preserved in 95% ethanol and stored at -20°C until being used for molecular identification of the species.

To verify the presence of begomoviruses infecting cassava, foliar samples were also collected at some sampled sites (Table S1). The samples were collected randomly regardless

Table 1 Sampled sites and whiteflies species detected in cassava in Brazil.

Sample	Date of collection	Location	Region	Geographical coordinates		Alt.	Number of samples			Whiteflies species ^c	Reference
				Latitude	Longitude		Ad ^b	Ny	T		
AL1	July 2018	Arapiraca, AL ^a	Northeast	09°48'27.66''S	36°36'40.68''W	241	15	15	30	Btu, Ta	This study
AL2	April 2018	TeotônioVilela, AL	Northeast	09°53'16.86''S	36°23'05.52''W	167	15	15	30	BtM, BtNW, Btu, Ta	This study
AL3	April 2018	União dos Palmares, AL	Northeast	09°11'51.84''S	36°01'51.90''W	154	14	10	24	BtM, BtNW, Btu, Ta	This study
AL4	April 2018	Arapiraca, AL	Northeast	09°47'29.46''S	36°25'36.00''W	143	14	15	29	BtM, Btu, Ta	This study
AL5	April 2018	Arapiraca, AL	Northeast	09°43'09.30''S	36°40'45.60''W	298	16	15	31	BtM, Ta	This study
BA1	December 2017	Barra, BA	Northeast	11°20'52.37''S	43°12'57.44''W	409	28	0	28	BtM, Btu, Ta	This study
BA2	March 2018	Luis Eduardo Magalhães, BA	Northeast	12°20'10.00''S	45°49'12.00''W	744	10	10	20	BtM, Btu	This study
BA3	June 2018	Wanderley, BA	Northeast	12°13'42.04''S	43°55'50.00''W	875	14	0	14	BtM, Ta	This study
BA4	March 2018	Cristópolis, BA	Northeast	12°13'56.08''S	44°23'11.50''W	694	13	0	13	BtM, Ta	This study
DF1	March 2018	Planaltina, DF	Midwest	15°30'59.00''S	47°16'09.00''W	809	10	0	10	BtM	This study
DF2	March 2018	Planaltina, DF	Midwest	15°28'57.00''S	47°20'06.00''W	757	10	0	10	BtM	This study
DF3	March 2018	Planaltina, DF	Midwest	15°31'17.00''S	47°21'22.00''W	874	10	0	10	BtM	This study
ES1	January 2018	Sooretama, ES	Southeast	19°06'52.02''S	40°04'46.30''W	60	11	11	22	Ta	This study
ES2	January 2018	Marilândia, ES	Southeast	19°24'22.04''S	40°32'21.70''W	111	14	15	29	BtM, Ta	This study
ES3	January 2018	Pinheiros, ES	Southeast	18°40'83.20''S	40°28'63.40''W	88	15	15	30	BtM, Btu, Ta	This study
GO1	March 2018	Bela Vista, GO	Midwest	16°59'50.00''S	48°57'56.00''W	808	10	11	21	BtM	This study
GO2	March 2018	Itaberaí, GO	Midwest	15°56'58.00''S	49°47'07.00''W	765	15	15	30	BtM, Btu	This study
MG1	February 2018	Ouro Fino, MG	Southeast	22°16'44.00''S	46°29'33.00''W	846	10	10	20	Ta	This study
MG2	February 2018	Pouso Alegre, MG	Southeast	22°15'01.00''S	46°58'31.00''W	677	15	15	30	BtM, Btu, Ta	This study
MG3	February 2018	Careaçu, MG	Southeast	22°04'37.00''S	45°41'49.00''W	813	15	15	30	Btu, Ta	This study
MG4	February 2018	Lambari, MG	Southeast	21°56'05.00''S	45°15'49.00''W	864	15	15	30	BtM, Ta	This study
MG5	February 2018	Lima Duarte, MG	Southeast	21°50'35.00''S	43°47'01.00''W	751	11	11	22	Ta	This study
MG6	February 2018	Rio Pomba, MG	Southeast	21°15'50.00''S	43°09'59.00''W	504	15	15	30	Btu, WtNEW2	This study
MG7	March 2018	Florestal, MG	Southeast	19°54'13.00''S	44°25'48.00''W	793	10	10	20	Ta	This study
MG8	March 2018	Florestal, MG	Southeast	19°51'20.00''S	44°23'58.00''W	704	10	10	20	Ta	This study
MG9	March 2018	Florestal, MG	Southeast	19°53'39.00''S	44°24'55.00''W	769	15	15	30	BtM, Ta	This study
MG10	March 2018	Florestal, MG	Southeast	19°52'38.00''S	44°25'21.00''W	780	16	13	29	BtM, Btu, Ta	This study
MG11	March 2018	Florestal, MG	Southeast	19°52'38.00''S	44°25'21.00''W	780	15	14	29	Ta	This study
MG12	March 2018	Divinópolis, MG	Southeast	20°06'21.00''S	44°55'36.00''W	782	15	15	30	Btu, Ta	This study
MG13	March 2018	Viçosa, MG	Southeast	20°46'06.00''S	42°52'14.00''W	661	14	11	25	BtM, BtNW, Btu, Ta	This study
MG14	April 2018	Piraúba, MG	Southeast	21°16'22.71''S	43°02'31.28''W	366	10	0	10	Ta	This study
MG15	May 2018	Descoberto, MG	Southeast	21°28'06.28''S	42°58'05.53''W	351	15	15	30	Btu, Ta	This study
MG16	June 2018	Mar de Espanha, MG	Southeast	21°46'07.26''S	43°04'26.62''W	508	15	15	30	Btu, Ta	This study
MG17	July 2018	Leopoldina, MG	Southeast	21°33'50.15''S	42°40'42.97''W	254	9	0	9	Ta	This study
MG18	August 2018	Dona Euzébia, MG	Southeast	21°19'18.83''S	42°48'37.07''W	246	17	0	17	Btu, Ta	This study
MG19	May 2018	Caparaó, MG	Southeast	20°31'48.00''S	41°54'00.36''W	864	12	10	22	Ta	This study

(continued on next page)

Table 1 (continued)

Sample	Date of collection	Location	Region	Geographical coordinates		Alt.	Number of samples			Whiteflies species ^c	Reference
				Latitude	Longitude		Ad ^b	Ny	T		
MT1	December 2017	Canarana, MT	Midwest	13° 31' 16.00" S	52° 25' 03.00" W	312	15	16	31	WtNEW1	This study
MT2	December 2017	Canarana, MT	Midwest	13° 33' 47.00" S	52° 15' 53.00" W	415	0	20	20	Btu	This study
MT4	January 2018	Pedra Preta, MT	Midwest	16° 38' 29.00" S	54° 25' 41.00" W	241	10	10	20	Btu	This study
MT5	January 2018	Pedra Preta, MT	Midwest	16° 39' 07.00" S	54° 22' 27.00" W	255	10	10	20	BtM, Btu	This study
MT6	January 2018	Pedra Preta, MT	Midwest	16° 39' 28.00" S	54° 20' 20.00" W	303	10	10	20	BtM, Btu	This study
PA1	January 2018	Brasil Novo, PA	North	03° 12' 23.07" S	52° 30' 13.80" W	170	11	10	21	Ta	This study
PA2	January 2018	Vitória doXingu, PA	North	03° 04' 51.01" S	52° 10' 08.80" W	130	10	12	22	Ta	This study
PA3	January 2018	Altamira, PA	North	03° 18' 15.03" S	52° 07' 26.00" W	176	15	15	30	Btu, Ta	This study
PA4	January 2018	Altamira, PA	North	03° 09' 14.60" S	52° 07' 49.00" W	128	15	16	31	Btu, Ta	This study
PA5	August 2018	Belém, PA	North	01° 18' 20.00" S	48° 26' 46.00" W	46	10	10	20	BtM	This study
PI1	April 2018	Picos, PI	Northeast	07° 04' 79.50" S	41° 25' 57.80" W	214	0	29	29	Btu, Ta	This study
PI2	May 2018	Teresina, PI	Northeast	05° 02' 41.94" S	42° 47' 18.84" W	71	0	27	27	Btu, Ta	This study
PR1	March 2018	Santo Antônio do Caiuá, PR	South	22° 41' 07.00" S	52° 19' 06.00" W	342	30	0	30	Btu, Ta	This study
PR2	March 2018	Santo Antônio do Caiuá, PR	South	22° 49' 03.00" S	52° 21' 25.00" W	439	30	0	30	BtM, Btu	This study
PR3	March 2018	Paraná, PR	South	23° 06' 03.29" S	52° 29' 10.00" W	471	26	0	26	Btu, Ta	This study
PR4	March 2018	Sertãozinho, PR	South	23° 02' 54.00" S	50° 59' 54.00" W	343	10	10	20	BtM, BtMED	This study
SC2	March 2018	Agrônoma, SC	South	27° 34' 40.00" S	48° 32' 08.00" W	34	10	0	10	BtM	This study
SP1	July 2016	Holambra, SP	Southeast	22° 36' 26.00" S	47° 02' 50.00" W	634	10	0	10	BtMED	Moraes et al. (2018)
SP2	July 2016	Mogi Mirim, SP	Southeast	22° 28' 08.00" S	46° 56' 25.00" W	652	10	0	10	Btu	Moraes et al. (2018)
SP3	July 2016	Mogi Mirim, SP	Southeast	22° 24' 59.00" S	46° 59' 19.00" W	678	10	0	10	Btu	Moraes et al. (2018)
SP4	July 2016	Mogi Mirim, SP	Southeast	22° 26' 44.00" S	47° 04' 11.00" W	689	10	0	10	Btu	Moraes et al. (2018)
SP5	July 2016	Mogi Mirim, SP	Southeast	22° 27' 05.00" S	47° 04' 56.00" W	710	10	0	10	Btu	Moraes et al. (2018)
SP6	July 2016	São Pedro, SP	Southeast	22° 34' 08.00" S	48° 05' 22.00" W	525	10	0	10	Btu	Moraes et al. (2018)
SP7	July 2017	Montalvão, SP	Southeast	22° 02' 23.00" S	51° 19' 53.00" W	404	10	0	10	Btu	Moraes et al. (2018)
SP8	July 2017	Pindamonhangaba, SP	Southeast	22° 56' 05.00" S	45° 26' 25.00" W	562	10	0	10	BtM	Moraes et al. (2018)
SP9	March 2016	Casa Branca, SP	Southeast	21° 11' 32.00" S	47° 48' 44.00" W	598	10	0	10	BtM	Moraes et al. (2018)
SP10	January 2019	São Pedro do Turvo, SP	Southeast	22° 36' 32.00" S	49° 45' 29.00" W	563	4	10	14	BtM, Btu, BtMED	This study
SP11	January 2019	Oleo, SP	Southeast	22° 56' 32.00" S	49° 26' 15.00" W	584	10	0	10	BtMED	This study
SP12	February 2019	Mogi Mirim, SP	Southeast	22° 28' 32.30" S	47° 00' 47.60" W	675	5	5	10	Btu	This study
SP13	July 2016	Casa Branca, SP	Southeast	21° 49' 08.00" S	46° 58' 23.00" W	612	10	0	10	Btu	Moraes et al. (2018)
Total							819	566	1385		

Notes.

^aBrazilian states where samples were collected: AL, Alagoas; BA, Bahia; DF, Distrito Federal; ES, Espírito Santo; GO, Goiás; MG, Minas Gerais; MT, Mato Grosso; PA, Pará; PI, Piauí; PR, Paraná; SC, Santa Catarina; SP, São Paulo.

^bAd, Adults; Ny, Nymphs; T, Total.

^cBtu, *Bemisia tuberculata*; BtM, *Bemisia tabaci* MEAM1; BtMED, *B. tabaci* MED; BtNW, *B. tabaci* New World; Ta, *Tetraleuroides acaciae*; WtNEW1, whitefly new species 1; WtNEW2, whitefly new species 2.

of the presence of virus-like symptoms. The leaves were press-dried and stored at room temperature as herbarium-like samples until being used for DNA extraction.

Whitefly species identification

Whitefly species were identified by sequencing of a mtCOI fragment, as previously described (Moraes *et al.*, 2018). When enough adults and nymphs were collected at a given sampled site, ten individuals from each stage were analyzed, and when only one stage was obtained, 20 individuals were tested (Table 1). An initial assessment of whitefly diversity was done using PCR-RFLP of the mtCOI fragment. When variation in the RFLP pattern was observed in the first screening, suggesting that more than one species could be present in that site, approximately five additional individuals for each stage were sequenced according to sample availability.

Total DNA was extracted from single individual whiteflies following a Chelex protocol (Walsh, Metzger & Higuchi, 1991). Briefly, adults or nymphs were ground in 30 μ l of Chelex buffer (5% Chelex in 1 \times Tris-EDTA) using a toothpick in a 600 μ l tube. Samples were vortexed for 30 s and incubated at 99 $^{\circ}$ C for 8 min in a PTC-100 thermocycler (MJ Research). Next, the tubes were centrifuged at 14,000 g for 5 min and 20 μ l of the supernatant was collected and transferred to a new tube. One microliter of the supernatant was used as a template for PCR amplification of a 800 bp fragment of the mtCOI gene using primers C1-J-2195 and L2-N-3014 (Frohlich *et al.*, 1999; Simon *et al.*, 1994). PCR was performed using 0.2 μ M of forward and reverse primers in a final volume of 25 μ l using GoTaq Colorless Master Mix (Promega), following the manufacturer's instructions. The PCR cycles consisted of an initial denaturing step at 95 $^{\circ}$ C for 5 min, followed by 35 cycles at 95 $^{\circ}$ C for 30 s, 42 $^{\circ}$ C for 45 s and 72 $^{\circ}$ C for 1 min, with a final extension at 72 $^{\circ}$ C for 10 min. Amplified products were visualized in 0.8% agarose gels stained with ethidium bromide and directly used for RFLP analysis (Bosco *et al.*, 2006).

RFLP analysis of the amplicons consisted of 5 μ l of each PCR product digested with 0.1 unit of *Taq* I (Promega) in a final volume of 20 μ l. Reactions were performed at 65 $^{\circ}$ C for 2 h and visualized in 1.2% agarose gels stained with ethidium bromide. To verify whether the predicted mtCOI restriction pattern corresponded to a given species according to *in silico* prediction, a subset of PCR products from adults and nymphs representative of distinct patterns from different sampled sites were selected and sequenced. PCR products were precipitated with 100% ethanol and 3 M sodium acetate pH 5.2 (Sambrook & Russel, 2001) and sequenced commercially (Macrogen Inc.) in both directions using primers C1-J-2195/L2-N-3014.

For a small subset of samples that failed to yield a PCR product using primers C1-J-2195 and L2-N-3014, a second screening, using a recently described primer set with improved specificity for species of the *B. tabaci* complex and *B. afer* (2195Bt and C012/Bt-sh2), was performed (Mugerwa *et al.*, 2018). Samples that still failed to amplify or had unexpected RFLP pattern were analyzed with specific primers for *T. vaporariorum* (TvapF and Wfrev) (Scott *et al.*, 2007).

Sequence comparisons and phylogenetic analysis

Nucleotide sequences were first checked for quality and assembled using Geneious v. 8.1 (Kearse et al., 2012). mtCOI sequences were initially analyzed with the BLAST n algorithm (Altschul et al., 1990) to determine the whitefly species with which they shared greatest similarity. Pairwise comparisons between all mtCOI sequences obtained here and those with higher similarities (as determined by the BLAST n search) were performed with the program SDT v. 1.2 (Muhire, Varsani & Martin, 2014) using the MUSCLE alignment option (Edgar, 2004).

For phylogenetic analyses, the final dataset was composed of 142 sequences: 95 obtained in this work and 47 sequences representative of species in the family Aleyrodidae. Sequences were retrieved from GenBank and from the updated mtCOI reference dataset for species of the *Bemisia tabaci* complex (Boykin, Savill & De Barro, 2017). Multiple sequence alignments were prepared using the MUSCLE option in MEGA7 (Kumar, Stecher & Tamura, 2016). Alignments were checked and manually adjusted when necessary. Phylogenetic trees were constructed using Bayesian inference performed with MrBayes v. 3.0b4 (Ronquist & Huelsenbeck, 2003). The program MrModeltest v. 2.2 (Nylander, 2004) was used to select the nucleotide substitution model with the best fit in the Akaike Information Criterion (AIC). The analyses were carried out running 50,000,000 generations with sampling at every 1,000 generations and a burn-in of 25%. The convergence was assumed when average standard deviation of split frequencies was lower than 0.001. Trees were visualized and edited using FigTree (tree.bio.ed.ac.uk/software/figtree) and CorelDRAW X5, respectively.

Virus detection in foliar samples

Total DNA was extracted as described (Doyle & Doyle, 1987) and used as a template for PCR using the DNA-A universal primer pair PAL1v1978 and PAR1c496 (Rojas et al., 1993). PCR was performed in a final volume of 25 μ l using *Taq* DNA Polymerase (Invitrogen) following the manufacturer's instructions. The PCR cycles consisted of an initial denaturing step at 95 °C for 5 min, followed by 35 cycles at 95 °C for 1 min, 52 °C for 1 min and 72 °C for 1 min, with a final extension at 72 °C for 10 min. PCR products were visualized in 0.8% agarose gels stained with ethidium bromide. In addition, rolling-circle amplification (Inoue-Nagata et al., 2004) followed by digestion with MspI was performed in a subset of the samples.

Diversity index and statistical analysis

Simpson's index of diversity (1-D) was calculated to verify if there was any difference in whitefly diversity across macroregions. This index represents the probability that two randomly chosen individuals in a given sampled site will belong to distinct species (Morris et al., 2014). Simpson's index was chosen as its value increases with increasing diversity and assigns more weight to more abundant species in a sample. We assume that species colonizing cassava will be in abundance, whereas rare species that briefly visit the plant without colonizing it will be underrepresented. Simpson's index was calculated for each sampled site separately and then pooled according to macroregions. To assess the

statistical significance of the differences in diversity among regions, the non-parametric Kruskal-Wallis test followed by *post hoc* multiple comparison test using Fisher's least significant difference was calculated, using the function `kruskal` implemented in the `Agricolae` package in R software (R Core Team, 2017). Non-parametric Spearman's rank correlation coefficient analysis was performed using the `ggpubr` package in R software (R Core Team, 2017).

RESULTS

High whitefly species richness in cassava in Brazil

To verify the composition of whitefly communities colonizing cassava in Brazil, sampling was performed across the country, including the main agroecological zones. A total of 66 sites from 12 states were sampled (Fig. 1; Table 1). Out of 1,385 individuals submitted to PCR-RFLP analysis, 58 adults and 37 nymphs from different locations and representing distinct restriction patterns were sequenced (Table S2). The combination of PCR-RFLP followed by sequencing showed reliability and consistency for species identification without misidentification due to incongruence between the two methods.

Based on pairwise comparisons and molecular phylogeny of the partial mtCOI gene, we identified the presence of at least seven species comprising the whitefly community in cassava (Fig. 2; Table 1). Among them, *T. acaciae* and *B. tuberculata*, both previously reported in this crop, were the most prevalent, representing over 75% of the analyzed individuals. In addition, based on the criterion of 3.5% divergence to differentiate species within the *B. tabaci* complex, three *B. tabaci* species were identified, with *Bt*MED previously reported, and *Bt*MEAM1 and *Bt*NW identified for the first time in cassava fields in Brazil (Fig. 2; Table 1). The species *Bt*MEAM1 represented 18% of the total individuals analyzed, followed by *Bt*MED (1.6%) and *Bt*NW (0.21%).

Furthermore, two putative new species were identified (Fig. 2), provisionally named whitefly new species 1 and 2 (WtNEW1 and WtNEW2). The WtNEW1 mtCOI sequence (KY249522) showed highest identity (80.65%) and clustered close to the *T. acaciae* clade, comprised of individuals reported here and three other previously reported sequences from cassava in Brazil (Fig. 2). For WtNEW2, two mtCOI sequences obtained from an adult (JX678666) and a nymph (DQ989531) shared 97.81% among them and showed highest identity with *B. tabaci* (adult: 82.11%; nymph: 81.68%) and clustered as a basal sister clade to the genus *Bemisia* (Fig. 2). Although whitefly taxonomy is predominantly based on puparial characters (Hodges & Evans, 2005) and there is no taxonomic criterion established based in mtCOI sequences for most of the groups, as has been proposed for the *B. tabaci* complex, the level of divergence between the two proposed new species with the closest species is similar to the level of divergence observed between species already described within the Aleyrodidae, as demonstrated in pairwise comparisons (Table S3) and phylogenetic analysis (Fig. 2). Nevertheless, further molecular and morphological characterization should be performed. Together, these results indicate the existence of a high whitefly species richness in cassava in Brazil.

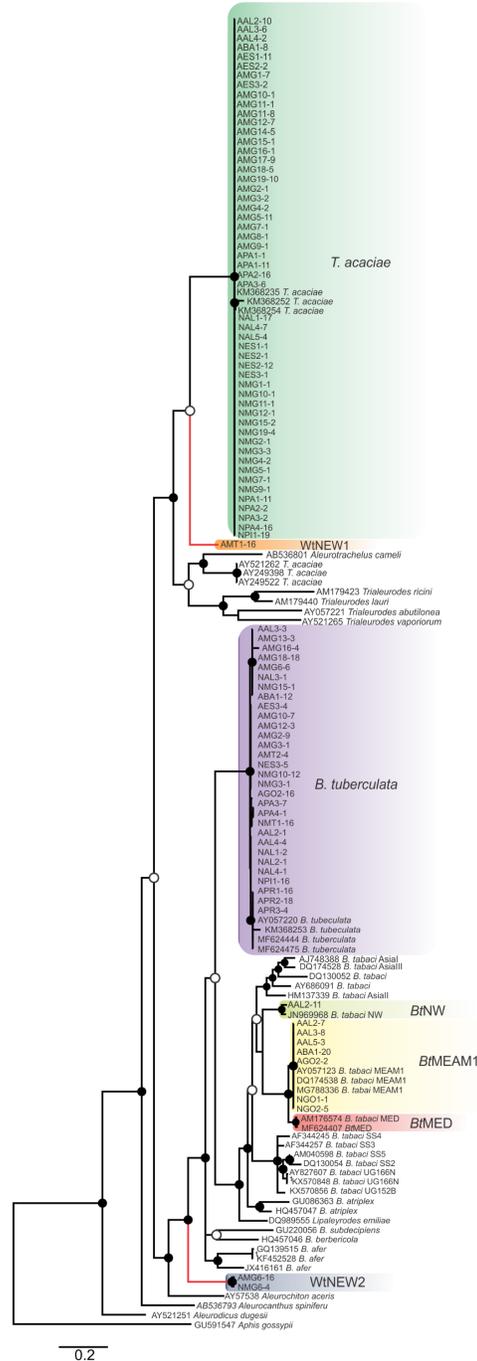


Figure 2 Phylogenetic relationships among whitely species, including the ones detected in cassava in Brazil, based on the sequence of the mtCOI gene. Bayesian phylogenetic tree based on partial nucleotide sequences of the mitochondrial cytochrome oxidase (mtCOI) gene of representative individuals of each whitely species detected in this study and reference sequences retrieved from GenBank. The tree was rooted with the aphid *Aphis gossypii*. Bayesian posterior probabilities are shown at the nodes. The scale bar represents the number of nucleotide substitutions per site. Nodes with posterior probability values between 0.60 and 0.80 are indicated by empty circles and nodes with values equal to or greater than 0.81 are indicated by filled circles. Clades highlighted with different colors indicate the species detected in this study. Branches highlighted in red indicate the putative new species detected here.

Full-size [DOI: 10.7717/peerj.11741/fig-2](https://doi.org/10.7717/peerj.11741/fig-2)

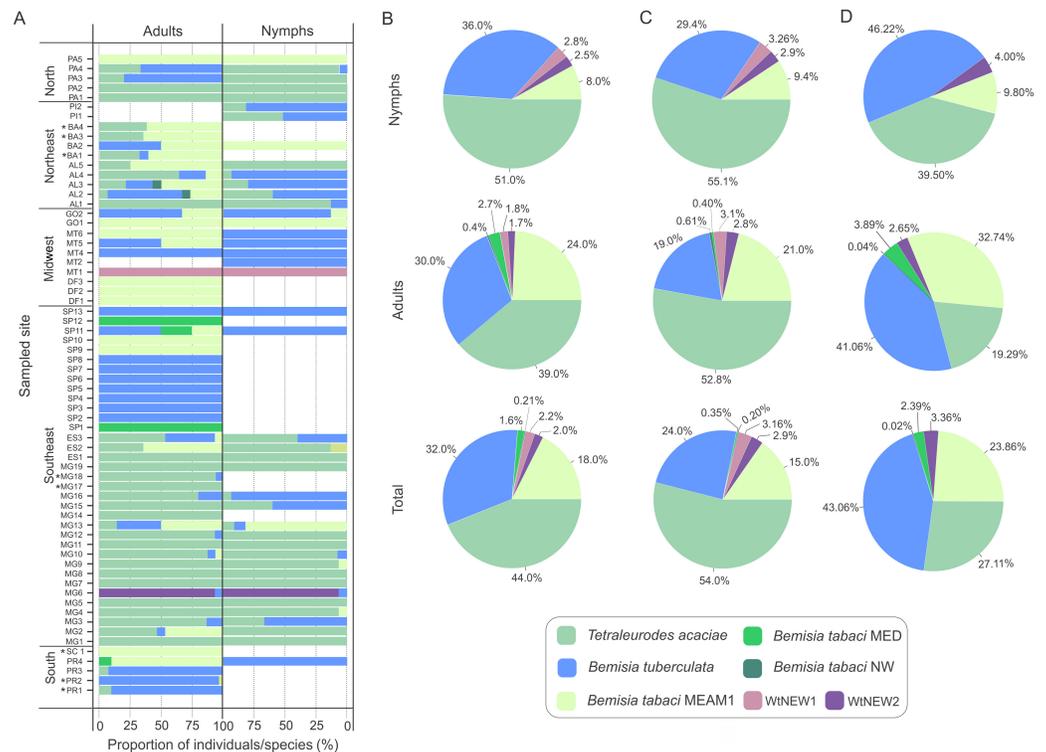


Figure 3 Composition of whitefly populations colonizing cassava in Brazil. (A) Species composition at each sampled site according to stage of development (adult and nymphs). Asterisks indicate that nymphs were not detected. (B, C, D) Species distribution of the 1,385 individuals genotyped in this study considering the samples from all sites (B) or only samples from sites where both adults and nymphs were sampled (C) or without samples from Minas Gerais state (D).

Full-size [DOI: 10.7717/peerj.11741/fig-3](https://doi.org/10.7717/peerj.11741/fig-3)

Both the prevalence and the capacity to colonize cassava differ among species

Nymphs were collected for samples identified as *T. acaciae*, *B. tuberculata*, *Bt*MEAM1 and the two new putative species (Fig. 3A), suggesting that these species may colonize cassava. Nymphs were not obtained at two sites where *Bt*MED was prevalent (SP1 and SP12). Although it could be suggested that this species has the potential to colonize cassava due to the high prevalence of adults at these two sites, the lack of nymphs suggests otherwise. Moreover, at the sites PR4 and MT6, *Bt*MEAM1 predominated among adults but 100% of the nymphs were *B. tuberculata*, suggesting that the predominance at one stage does not necessarily mean predominance in another stage. Indeed, correlation analysis between the number of adults and nymphs, performed for all sites where both stages were sampled, showed no significant correlation between them (Fig. S1). Further sampling in those sites or free-choice experiments are necessary to confirm the potential of *Bt*MED to colonize cassava. Considering the whole sampling, we detected only three adults of *Bt*NW, suggesting an inability of this specie to colonize cassava.

To verify if prevalence differs among species across distinct developmental stages, the data were separated according to stage and the proportions of individuals were compared

for the three most abundant species (Figs. 3B, 3C). Considering the entire data set, *T. acaciae* was the prevalent species, followed by *B. tuberculata* and *BtMEAM1* ($x_2^2 = 152.63$, $P < 2.2 \times 10^{-16}$). The same was true according to stage, either adults ($x_2^2 = 28.61$, $P < 6.1 \times 10^{-07}$) or nymphs ($x_2^2 = 169.44$, $P < 2.2 \times 10^{-16}$; Fig. 3B). However, caution is needed to interpret these results as only adults were sampled at some sites where *BtMEAM1* and *B. tuberculata* were prevalent (Fig. 3A), which could bias the analysis, causing an underestimation of the number of nymphs for those species. Therefore, we also analyzed the data considering only those sites where both adults and nymphs were obtained. Again, *T. acaciae* was the predominant species followed by *B. tuberculata* and *BtMEAM1* considering either the entire data set ($x_2^2 = 258.61$, $P < 2.2 \times 10^{-16}$) or only nymphs ($x_2^2 = 164.47$, $P < 2.2 \times 10^{-16}$). When only adults were considered, *T. acaciae* was still predominant ($x_2^2 = 113.52$, $P < 2.2 \times 10^{-16}$) but no difference between *B. tuberculata* and *BtMEAM1* was observed ($x_1^2 = 0.505$, $P = 0.477$; Fig. 3C). Moreover, it could be argued that samples from Minas Gerais (MG) were overrepresented in our sampling (Fig. 1C), which could also bias the results presented above due to the predominance of *T. acaciae* in this state (Fig. 3A). To test this possibility, we analyzed the data excluding the samples from MG. In this case, when both stages were considered, *B. tuberculata* was predominant ($x_2^2 = 62.09$, $P = 3.3 \times 10^{-14}$) but no difference between *T. acaciae* and *BtMEAM1* was observed ($x_1^2 = 1.91$, $P = 0.166$). When each stage was considered separately, *B. tuberculata* was predominant followed by *BtMEAM1* and *T. acaciae* (adults: $x_2^2 = 43.94$, $P = 2.9 \times 10^{-10}$; nymphs: $x_2^2 = 84.19$, $P < 2.2 \times 10^{-16}$). Together, these results indicate that the potential to colonize cassava differs among species, which could be due either to lower preference for the plant or to differences in the competitive ability among species during cassava colonization. In addition, they reinforce the low efficiency of *BtMEAM1* to colonize cassava.

Competitive interference does not explain the differences in prevalence

Interestingly, at least two species were detected co-occurring at 51% of the sampled sites (Fig. 3A). To verify the possibility of competition among *T. acaciae*, *B. tuberculata* and *BtMEAM1* to explain the observed differences in prevalence (instead of differences in host preference), the competitive capacity of these three species was inferred based on the analysis of predominance at the sites where they occurred together. Initially, we verified if there were any differences in incidence, defined here as the number of sampled sites where at least one individual belonging to one of the three species was detected (Fig. 4A). The results demonstrate that there were no differences in incidence among them ($x_2^2 = 1.25$, $P = 0.537$; Fig. 4A). In addition, no differences were observed when the proportion of sites where whitefly species occurred alone or in different combinations was compared ($x_6^2 = 3.26$, $P = 0.776$; Fig. 4B). However, when we compared the occurrence between *BtMEAM1* and non-*B. tabaci* species at the sites where they occur alone, the number of sites with non-*B. tabaci* species was higher ($x_1^2 = 6.53$, $P = 0.011$; Fig. 4B). Thus, the competitive capacity was inferred based on the proportion of individuals from each species at the fields where these species were detected co-occurring in different combinations (Fig. 4C). Interestingly, at the sites where *BtMEAM1* and *B. tuberculata* were sampled together, *B. tuberculata*

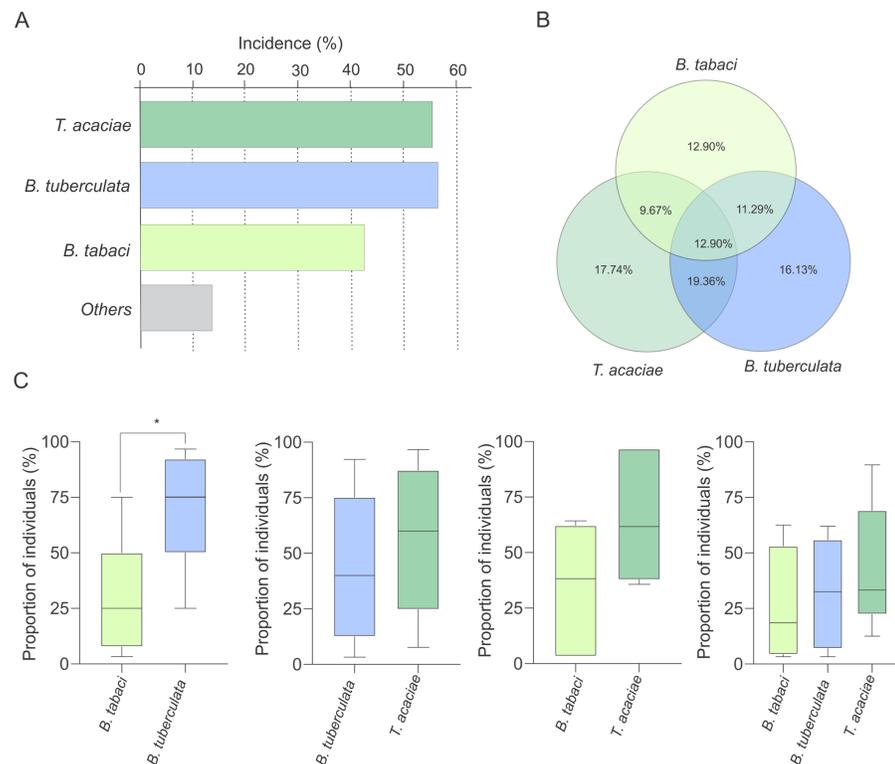


Figure 4 Incidence of *Trialeurodes acaciae*, *Bemisia tuberculata* and *B. tabaci* MEAM1 in cassava fields in Brazil. (A) Incidence of *Trialeurodes acaciae*, *Bemisia tuberculata* and *B. tabaci* MEAM1 in cassava fields in Brazil, measured as the percentage of sampled sites where at least one individual belonging to each one of the three species was detected. Other species detected at low incidence are shown together as "others". (B) Venn diagram showing the proportion of sites where each one of the three whitefly species occur alone or in different combinations. (C) Competitive capacity inferred based on the prevalence of individuals from each of two species in fields where those two species were detected co-occurring. The horizontal line inside the box corresponds to the median. The asterisk indicates a significant difference according to the non-parametric Kruskal-Wallis test ($p < 0.05$).

Full-size DOI: 10.7717/peerj.11741/fig-4

predominated over *Bt* MEAM1, suggesting higher competitive potential (Fig. 4C). For all other species combinations, no evidence of differences in competitive capacity were observed (Fig. 4C). Together, these results suggest that, rather than competition, lower host preference by *Bt* MEAM1 explains its non-prevalence compared to *T. acaciae* and *B. tuberculata*, resulting in low colonization rate as indicated by the low number of *Bt* MEAM1 nymphs detected in cassava (Fig. 3).

Composition and species diversity of whiteflies differ among Brazilian regions

The predominance of species composing the whitefly community across macroregions varied considerably. While *T. acaciae* predominated in the North, Southeast and Northeast, it was not detected in the Midwest (Fig. 5A). In addition, *B. tuberculata* was detected in all regions, and was prevalent in the South and Midwest. *Bt* MEAM1, although not prevalent in any of the regions, was also detected in all regions. Although the number of species

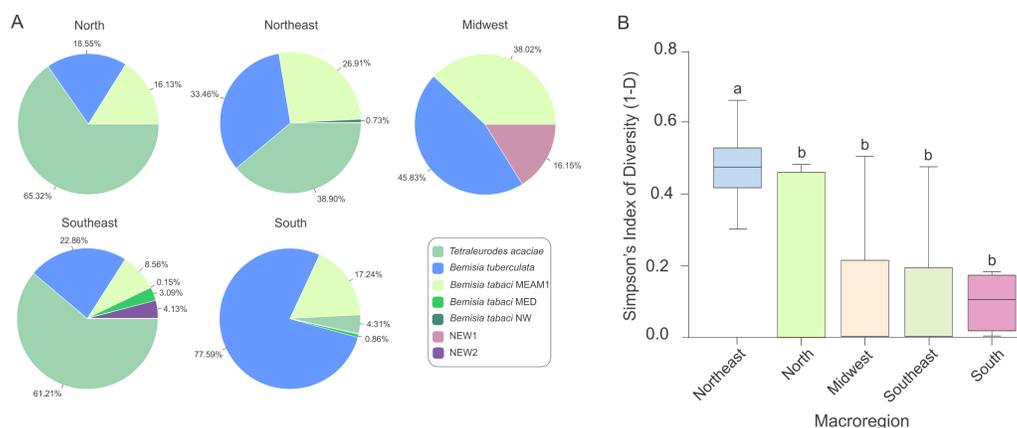


Figure 5 Composition and species diversity of whitefly populations differ among Brazilian regions. (A) Pie charts represent the distribution of the 1,385 individuals genotyped in this study in the five geographic regions of Brazil. (B) Boxplots correspond to Simpson's index of diversity (1-D) calculated for each geographic region. The index was first calculated for each sampled site and grouped by geographic region. Different letters indicate significant differences between groups according to the non-parametric Kruskal-Wallis test followed by *post hoc* multiple comparison test ($p < 0.05$).

Full-size DOI: [10.7717/peerj.11741/fig-5](https://doi.org/10.7717/peerj.11741/fig-5)

detected was higher in the Southeast, where six species out seven were detected, whitefly diversity was significantly higher in fields in the Northeast according to Simpson's index of diversity (Fig. 5B), with no differences among the other four regions.

No begomoviruses detected infecting cassava

To verify the presence of begomoviruses infecting cassava, we analyzed leaves sampled in some of the fields where whiteflies were collected (Table S1). Based on PCR detection using universal primers for begomoviruses, all plants were negative. Although the possibility of false negatives cannot be completely discarded, it is unlikely since the PCR assay used primers known to detect CMBs (Berrie, Rybicki & Rey, 2001; Rojas et al., 1993; Zhou et al., 1997) and none of the plants displayed symptoms (Table S1). Nevertheless, as a confirmatory step, we performed RCA followed by digestion with MspI in five samples, all with negative results (Table S1).

DISCUSSION

Vectors play an essential role during the life cycle of plant viruses, directly affecting their ecology and evolution (Gallet, Michalakos & Blanc, 2018; Gutierrez & Zwart, 2018; Sacristan et al., 2003). Usually, a group of plant viruses establishes a very specific interaction with only one or a few related species of vectors, making virus ecology strongly dependent on that of its vector (Gallet, Michalakos & Blanc, 2018). It has been suggested that the natural host range of a virus is dependent on its vector's host range, as most plant viruses have greater specificity for the vector than for the plant host (Dietzgen, Mann & Johnson, 2016; Elena, Fraile & Garcia-Arenal, 2014). Indeed, the existence of a competent vector for transmission and able to colonize potential reservoir and recipient new hosts is a primary ecological factor driving host range expansion of viruses. Thus, vectors play an essential

role during viral disease emergence and epidemics ([Elena, Fraile & Garcia-Arenal, 2014](#); [Feres, 2015](#); [Gilbertson et al., 2015](#); [Navas-Castillo, Fiallo-Olivé & Sánchez-Campos, 2011](#)). Understanding ecological factors, such as vector species dynamics in crops, might provide important clues about historical and current events of emergence or re-emergence of viral diseases, and even anticipate the potential for new ones to occur ([Legg et al., 2014](#)).

Although it could be suggested that there are no begomoviruses capable of infecting cassava in the Americas, the high diversity of begomoviruses reported in a broad range of cultivated and non-cultivated plants in several botanical families, including the Euphorbiaceae, make this highly unlikely ([Albuquerque et al., 2012a](#); [Albuquerque et al., 2012b](#); [Castillo-Urquiza et al., 2008](#); [Fernandes et al., 2008](#); [Fernandes et al., 2011](#); [Macedo et al., 2018](#); [Mar et al., 2017b](#); [Paz-Carrasco et al., 2014](#); [Rocha et al., 2013](#); [Rodríguez-Negrete et al., 2019](#)). Besides, CMBs could be introduced from infected rootstocks, as demonstrated by the introduction of Sri Lankan cassava mosaic virus into China ([Wang et al., 2020](#); [Wang et al., 2019](#)). Thus, the absence of a competent vector able to colonize cassava and transfer begomoviruses from wild plants to cassava, as previously suggested ([Carabali et al., 2005](#)), seems to be a more plausible hypothesis to explain the lack of begomovirus epidemics in this crop.

Our country-wide survey of whiteflies associated with cassava in Brazil uncovered a high degree of species diversity and showed that *T. acaciae* and *B. tuberculata* are the prevalent species across the country. Non-*B. tabaci* species, including *B. tuberculata*, have been shown to be prevalent also in Colombia ([Gómez-Díaz, Montoya-Lerma & Muñoz Valencia, 2019](#)). In contrast, in Africa, endemic species of the *B. tabaci* complex are prevalent in cassava ([Jacobson, Duffy & Sseruwagi, 2018](#); [Legg et al., 2014](#); [Tocko-Marabena et al., 2017](#)). Previous studies surveying whitefly diversity in South American countries failed to detect *T. acaciae* and *B. tuberculata* in crops other than cassava, indicating a very narrow host range, which may in fact be restricted to cassava or at least to cultivated plants ([Alemndri et al., 2015](#); [Marubayashi et al., 2014](#); [Moraes et al., 2018](#)).

BtMEAM1 and *BtNW* are reported here for the first time in cassava in Brazil. *BtMEAM1* was the third most prevalent species, representing 18% of the genotyped individuals, and with similar incidence to *T. acaciae* and *B. tuberculata*. The failure of previous studies to detect *BtMEAM1* in cassava may have been due to the small number of samples analyzed. The wide distribution and prevalence of *BtMEAM1* in the main agroecological zones in Brazil has been well established, mostly in association with annual crops such as soybean, cotton, common bean and tomato ([Moraes et al., 2018](#)). In these crops, *BtMEAM1* has a great reproductive capacity, rapidly increasing its population. Interestingly, our data showed the higher prevalence of *BtMEAM1* to be in the Midwest, where extensive agriculture predominates. The harvest of annual crops in the Midwest might cause the migration of the insect to semiperennial hosts such as cassava, which could explain why in some sites where *BtMEAM1* predominated among adults, it was not detected as nymphs (e.g., sites MT5, MT6, PR4).

It will be important to monitor *BtMEAM1* populations in cassava over the next years, to assess its possible adaptation to this host. The fact that we collected *BtMEAM1* nymphs at several locations suggests that this process may already be under way. We also detected

BtMED, a worrying result given the recent introduction of this species in the Brazil and its potential to displace other species, including *BtMEAM1* (Liu et al., 2012; Sun et al., 2013; Watanabe et al., 2019). *BtMED* has disseminated quickly in the country, mainly in association with ornamental plants in greenhouses (Moraes et al., 2018). Even though we detect *BtMED* associated to cassava, we cannot infer its potential to effectively colonize this host since only adults were collected. The third species detected is the indigenous *BtNW*. Although *BtMEAM1* partially displaced *BtNW* in Brazil, this species can still be sporadically detected, mostly in association with non-cultivated hosts (Marubayashi et al., 2014; Marubayashi et al., 2013; Moraes et al., 2018). It has been recurrently detected in *Euphorbia heterophylla*, suggesting a potential to colonize other species in the family Euphorbiaceae. However, the very low frequency with which it was detected and the absence of nymphs indicate that *BtNW* is poorly adapted to cassava.

The identification of two putative new species highlights the remarkable genetic diversity of whiteflies. Interestingly, one of the new species was collected in the state of Mato Grosso, which corresponds to the region considered to be the domestication center of cassava (Clement et al., 2016; Leotard et al., 2009; Olsen & Schaal, 1999; Watling et al., 2018). Further studies are needed to explore plant biodiversity in this region (Nassar, 2001; Olsen, 2004), which might reveal a similar diversity of whiteflies which may be specifically adapted to non-cultivated plant species due to long term co-evolution. The close phylogenetic relationship of the new species with non-*B. tabaci* whiteflies suggests that they are not virus vectors.

Whitefly species richness in cassava is just starting to be assessed and may be greater than reported here. Based on morphological characters, Alonso, Racca-Filho & Lima (2012) reported the presence of *Aleurothrixus aepim* and *Trialeurodes manihoti* colonizing cassava in the state of Rio de Janeiro. Although we did not analyze samples from that region, the failure to detect these species in other states suggests a restricted occurrence. Moreover, morphological characters alone are not always sufficient to classify whiteflies at the species level, and additional studies using molecular tools are needed to assess these molecularly uncharacterized whiteflies species (Dickey et al., 2015).

Host suitability has been shown to be an important factor influencing the competitive capacity among species of the *B. tabaci* complex (Luan et al., 2012; Sun et al., 2013; Watanabe et al., 2019). Watanabe et al. (2019) demonstrated that displacement capacity between two invasive *B. tabaci* species was dependent on host suitability. While *BtMEAM1* displaced *BtMED* only on tomato, *BtMED* displaced *BtMEAM1* on sweet pepper and common bean. Luan et al. (2012) demonstrated that even in a host plant poorly suitable for *BtMEAM1*, it was able to displace an indigenous species challenger. These authors demonstrated that even though host suitability may affect the speed of displacement, it may not affect the direction, as *BtMEAM1* always won the challenge (Luan et al., 2012). Interestingly, two or more species occurring sympatrically were detected in 51% of the fields analyzed in our study. In sites where *BtMEAM1* and *B. tuberculata* co-occurred, *B. tuberculata* predominated, suggesting a higher competitive capacity. Nonetheless, in all other combinations of co-occurring species, no differences in prevalence were observed. Thus, competitive capacity is unlikely to explain the low prevalence of *BtMEAM1*, or

the differences observed between *T. acaciae* and *B. tuberculata*. However, we collected whiteflies in a single point in time at each location. Since competition is a dynamic process, additional surveys should be conducted in the future to further confirm these observations.

Host adaptation may be a more important component affecting the low predominance of *Bt* MEAM1 in cassava, as previously suggested (Carabali *et al.*, 2005). The inability of *Bt*MEAM1 and *Bt*MED to colonize domesticated cassava efficiently has been demonstrated under experimental conditions (Carabali, Belloti & Montoya-Lerma, 2010; Carabali, Montoya-Lerma & Bellotti, 2008; Milenovic *et al.*, 2019; Vyskočilová, Seal & Colvin, 2019). Carabali, Montoya-Lerma & Bellotti (2008), evaluating the colonization potential of *Bt*MEAM1 in three commercial cassava genotypes, demonstrated that only in one of them did *Bt*MEAM1 complete its development cycle from eggs to adult, and even then, at very low rates (0.003%). Using an electrical penetration graph assay, Milenovic *et al.* (2019) demonstrated the inability of *Bt*MED to feed in cassava plants. Adults of this species spent a very short time ingesting cassava phloem sap compared to sap from a suitable host, suggesting that they would die by starvation in the field. Furthermore, the low efficiency of whiteflies of the *Bt*MED mitochondrial subgroups Q1 and Q2 in using cassava as a host has also been demonstrated (Vyskočilová, Seal & Colvin, 2019). Oviposition and adult survival rates were very low, and development from eggs to adults was not observed. Although these studies were conducted under experimental conditions, the low predominance of *Bt*MEAM1 and *Bt*MED shown here and in other field surveys in Africa (Ghosh, Bouvaine & Maruthi, 2015; Tajebe *et al.*, 2015; Tocko-Marabena *et al.*, 2017) strongly indicates a low adaptation of these species to cassava.

Nevertheless, our results indicate an ongoing adaptation process of *Bt*MEAM1 to cassava, with the detection of nymphs and adults in the same field. Interestingly, Carabali *et al.* (2005) demonstrated a gradual increase in the rate of reproduction and development of *Bt*MEAM1 after successive passages on plants phylogenetically related to the genus *Manihot* (*Euphorbia pulcherrima* and *Jatropha gossypifolia*), indicating the potential of this whitefly species to become adapted to cassava through intermediate hosts. Furthermore, successful reproduction in the wild relative *M. esculenta ssp. flabellifolia* indicates that this plant may constitute an intermediate host leading to adaptation (Carabali, Belloti & Montoya-Lerma, 2010). This plant has been reported to be widely spread in the Amazon basin and the Midwest region of Brazil (Olsen, 2004). Interestingly, our data showed the higher prevalence of *Bt*MEAM1 to be in the Midwest. Although we cannot establish a cause and effect relationship, it is reasonable to speculate that *M. esculenta ssp. flabellifolia* could be acting as an intermediate host mediating adaptation. A survey addressing whitefly diversity in this host should be necessary to test this hypothesis.

In Brazil, cassava is predominantly grown as a subsistence crop, usually side by side with other vegetables and with a high incidence of weeds. Growing cassava in a heterogeneous environment, especially in the presence of related plants, may increase the adaptation potential of *Bt*MEAM1 and other species of the complex such as *Bt*MED, which we also detected in the open field. A high diversity of plants in cassava fields may allow an overlapping of ecological niches for distinct whitefly species, which under enough selection pressure may gradually adapt to new hosts. The sympatric occurrence of *T.*

acaciae, *B. tuberculata* and *BtMEAM1*, supports the role of botanical heterogeneity in shaping the composition of whitefly populations associated with cassava. A similar pattern was observed in Colombia, with 66% of the surveyed sites showing at least two species occurring sympatrically (Gómez-Díaz, Montoya-Lerma & Muñoz Valencia, 2019). Moreover, a predominance of one species in a given developmental stage and a different one in another stage (e.g., nymphs vs adults) at the same site suggests that other hosts may sustain reproduction and development, with adults migrating to cassava.

Euphorbia heterophylla (family Euphorbiaceae) is an invasive weed widely spread across Brazil and associated with several crops (Mar et al., 2017b; Wilson, 1981). The presence of *E. heterophylla* plants in association with cassava (Fig. 1A) and the fact that it was the most suitable host for *BtMEAM1* in Brazil out seven tested (Sottoriva, Lourenção & Colombo, 2014) shows its potential to act as an intermediate host mediating *BtMEAM1* adaptation. *E. heterophylla* has been frequently associated with the begomovirus *Euphorbia yellow mosaic virus* (EuYMV) (Mar et al., 2017b). Barreto et al. (2013) demonstrated that this plant is also a host of *Tomato severe rugose virus* (ToSRV), which even at a very low titer was transmitted to tomato plants, demonstrating the potential of *E. heterophylla* to act as a reservoir host. Surprisingly, considering that *E. heterophylla* and tomato belong to distinct botanical families, EuYMV is able to infect tomato (Barreto et al., 2013). The closer botanical relationship between *E. heterophylla* and cassava may indicate a higher potential of EuYMV to infect cassava. The presence of EuYMV-infected *E. heterophylla* in cassava fields, as observed in this study (Fig. 1A), its suitability as a host for *BtMEAM1*, and the high efficiency of EuYMV transmission by *BtMEAM1* (Mar et al., 2017a), suggest that EuYMV may have spillover potential to cassava. Experiments are ongoing in our laboratory to assess this spillover potential.

The emergence of begomoviruses in tomato crops in Brazil followed the introduction of *BtMEAM1* (Ribeiro et al., 1998; Rocha et al., 2013), demonstrating the role of vector populations in promoting viral host range expansion and consequently epidemics. Thus, the adaptation of whiteflies to cassava could facilitate the emergence of begomoviruses in this crop. The establishment of management strategies to prevent or at least delay the adaptation process is therefore necessary. *Bemisia tabaci* species may disperse across long distances via international trade routes (Hadjistylli, Roderick & Brown, 2016). Thus, preventing the introduction of cassava-adapted *B. tabaci* species from Africa should also be a priority.

CONCLUSIONS

We hypothesized that the absence of cassava-infecting begomoviruses in Brazil would be due to lack of competent *B. tabaci* vector species that efficiently colonize cassava. The results of our country-wide survey provide support to this hypothesis, with the most prevalent species being the non-vectors *Tetraleurodes acaciae* and *Bemisia tuberculata*. However, we did detect the presence of adult insects of *Bemisia tabaci* MEAM1, suggesting an ongoing adaptation process of this species to cassava which could facilitate the emergence of begomoviruses in this crop. Management strategies to prevent or at least delay the adaptation process are necessary.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was funded by CAPES (Financial code 01), CNPq (grants 409599/2016-6 to Francisco M. Zerbini and 405684/2018-5 to Renate Krause-Sakate), Fapemig (grant APQ-03276-18 to Francisco M. Zerbini), and FAPESP (grant 2017/21588-7 to Renate Krause-Sakate). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

CAPES: 01.

CNPq: 409599/2016-6, 405684/2018-5.

Fapemig: APQ-03276-18.

FAPESP: 2017/21588-7.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Cesar A.D. Xavier conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Angélica Maria and Vinic Nogueira conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
- Luís Fernando Maranhão Watanabe conceived and designed the experiments, performed the experiments, prepared figures and/or tables, and approved the final draft.
- Tarsiane Mara Carneiro Barbosa performed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Miguel Alves Júnior, Leonardo Barbosa, José E.A. Beserra-Júnior, Alessandra Boari, Renata Calegario, Eduardo Silva Gorayeb, Jaime Honorato Júnior, Gaus Silvestre de Andrade Lima, Raquel Neves de Mello, Fábio Nascimento Silva and Enilton Nascimento Santana performed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Gabriel Koch, Cristian Lopes, Késsia Pantoja, Roberto Ramos Sobrinho and José Wilson Pereira da Silva performed the experiments, prepared figures and/or tables, and approved the final draft.
- Renate Krause-Sakate and Francisco M. Zerbini conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The whitefly mtCOI sequences are available in GenBank: [MT901081](#) to [MT901172](#), [MT904381](#), and [MT904382](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.11741#supplemental-information>.

REFERENCES

- Albuquerque LC, Inoue-Nagata AK, Pinheiro B, Resende RO, Moriones E, Navas-Castillo J. 2012a. Genetic diversity and recombination analysis of sweepoviruses from Brazil. *Virology Journal* **9**:241 DOI [10.1186/1743-422X-9-241](https://doi.org/10.1186/1743-422X-9-241).
- Albuquerque LC, Varsani A, Fernandes FR, Pinheiro B, Martin DP, Ferreira PTO, Lemos TO, Inoue-Nagata AK. 2012b. Further characterization of tomato-infecting begomoviruses in Brazil. *Archives of Virology* **157**:747–752 DOI [10.1007/s00705-011-1213-7](https://doi.org/10.1007/s00705-011-1213-7).
- Alemandri V, Vaghi Medina CG, Dumon AD, Arguello Caro EB, Mattio MF, Garcia Medina S, Lopez Lambertini PM, Truol G. 2015. Three members of the *Bemisia tabaci* (Hemiptera: Aleyrodidae) cryptic species complex occur sympatrically in Argentine horticultural crops. *Journal of Economic Entomology* **108**:405–413 DOI [10.1093/jee/tov017](https://doi.org/10.1093/jee/tov017).
- Alonso RS, Racca-Filho F, Lima AF. 2012. Occurrence of whiteflies (Hemiptera: Aleyrodidae) on cassava (*Manihot esculenta* Crantz) crops under field conditions in the state of Rio de Janeiro, Brazil. *EntomoBrasilis* **5**:78–79.
- Altschul SE, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *Journal of Molecular Biology* **215**:403–410 DOI [10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
- Alves AAC, Setter TL. 2004. Response of cassava leaf area expansion to water deficit: cell proliferation, cell expansion and delayed development. *Annals of Botany* **94**:605–613 DOI [10.1093/aob/mch179](https://doi.org/10.1093/aob/mch179).
- Barbosa LF, Yuki VA, Marubayashi JM, De Marchi BR, Perini FL, Pavan MA, Barros DR, Ghanim M, Moriones E, Navas-Castillo J, Krause-Sakate R. 2015. First report of *Bemisia tabaci* Mediterranean (Q biotype) species in Brazil. *Pest Management Science* **71**:501–504 DOI [10.1002/ps.3909](https://doi.org/10.1002/ps.3909).
- Barreto SS, Hallwass M, Aquino OM, Inoue-Nagata AK. 2013. A study of weeds as potential inoculum sources for a tomato-infecting begomovirus in central Brazil. *Phytopathology* **103**:436–444 DOI [10.1094/phyto-07-12-0174-r](https://doi.org/10.1094/phyto-07-12-0174-r).
- Berrie LC, Rybicki EP, Rey MEC. 2001. Complete nucleotide sequence and host range of South African cassava mosaic virus: further evidence for recombination amongst begomoviruses. *Journal of General Virology* **82**:53–58 DOI [10.1099/0022-1317-82-1-53](https://doi.org/10.1099/0022-1317-82-1-53).
- Bosco D, Loria A, Sartor C, Cenis JL. 2006. PCR-RFLP identification of *Bemisia tabaci* biotypes in the Mediterranean Basin. *Phytoparasitica* **34**:243 DOI [10.1007/BF02980951](https://doi.org/10.1007/BF02980951).
- Boykin LM, Savill A, De Barro P. 2017. Updated mtCOI reference dataset for the *Bemisia tabaci* species complex. *F1000 Research* **6**:1835 DOI [10.12688/f1000research.12858.1](https://doi.org/10.12688/f1000research.12858.1).

- Carabali A, Bellotti AC, Montoya-Lerma J. 2010.** Biological parameters of *Bemisia tabaci* (Gennadius) biotype B (Hemiptera: Aleyrodidae) on *Jatropha gossypifolia*, commercial (*Manihot esculenta*) and wild cassava (*Manihot flabellifolia* and *M. carthagenensis*) (Euphorbiaceae). *Neotropical Entomology* **39**:562–567 DOI [10.1590/S1519-566X2010000400015](https://doi.org/10.1590/S1519-566X2010000400015).
- Carabali A, Bellotti AC, Montoya-Lerma J, Cuellar ME. 2005.** Adaptation of *Bemisia tabaci* biotype B (Gennadius) to cassava, *Manihot esculenta* (Crantz). *Crop Protection* **24**:643–649 DOI [10.1016/j.cropro.2004.11.008](https://doi.org/10.1016/j.cropro.2004.11.008).
- Carabali A, Montoya-Lerma J, Bellotti AC. 2008.** Development and reproduction of *Bemisia tabaci* B (Hemiptera: Aleyrodidae) on cassava (*Manihot esculenta*) genotypes. *Revista Colombiana de Entomologia* **34**:28–32.
- Carter SE, Fresco LO, Jones PG, Fairbairn JN. 1997.** *Introduction and diffusion of cassava in Africa. IITA Research Guide no. 49.* Ibadan: International Institute of Tropical Agriculture (IITA).
- Castillo-Urquiza GP, Beserra Jr JEA, Bruckner FP, Lima ATM, Varsani A, Alfenas-Zerbini P, Zerbini FM. 2008.** Six novel begomoviruses infecting tomato and associated weeds in Southeastern Brazil. *Archives of Virology* **153**:1985–1989 DOI [10.1007/s00705-008-0172-0](https://doi.org/10.1007/s00705-008-0172-0).
- Chandrashekar K, Rao A, Gorane A, Verma R, Tripathi S. 2020.** Aleurothrixus trachoides (Back) can transmit begomovirus from *Duranta* to potato, tomato and bell pepper. *Journal of Biosciences* **45**:36 DOI [10.1007/s12038-020-0012-2](https://doi.org/10.1007/s12038-020-0012-2).
- Clement CR, Rodrigues DP, Alves-Pereira A, Mühlen GS, Md Cristo-Araújo, Moreira PA, Lins J, Reis VM. 2016.** Crop domestication in the upper Madeira River basin. *Boletim Do Museu Paraense Emílio Goeldi Ciências Humanas* **11**:193–205 DOI [10.1590/1981.81222016000100010](https://doi.org/10.1590/1981.81222016000100010).
- De Barro PJ, Liu SS, Boykin LM, Dinsdale AB. 2011.** *Bemisia tabaci*: a statement of species status. *Annual Review of Entomology* **56**:1–19 DOI [10.1146/annurev-ento-112408-085504](https://doi.org/10.1146/annurev-ento-112408-085504).
- De Bruyn A, Harimalala M, Zinga I, Mabvakure BM, Hoareau M, Ravigne V, Walters M, Reynaud B, Varsani A, Harkins GW, Martin DP, Lett JM, Lefeuvre P. 2016.** Divergent evolutionary and epidemiological dynamics of cassava mosaic geminiviruses in Madagascar. *BMC Evolutionary Biology* **16**:182 DOI [10.1186/s12862-016-0749-2](https://doi.org/10.1186/s12862-016-0749-2).
- De Bruyn A, Villemot J, Lefeuvre P, Villar E, Hoareau M, Harimalala M, Abdoul-Karime AL, Abdou-Chakour C, Reynaud B, Harkins GW, Varsani A, Martin DP, Lett JM. 2012.** East African cassava mosaic-like viruses from Africa to Indian ocean islands: molecular diversity, evolutionary history and geographical dissemination of a bipartite begomovirus. *BMC Evolutionary Biology* **12**:228 DOI [10.1186/1471-2148-12-228](https://doi.org/10.1186/1471-2148-12-228).
- De Marchi BR, Marubayashi JM, Favara GM, Yuki VA, Watanabe LFM, Barbosa LF, Pavan MA, Krause-Sakate R. 2017.** Comparative transmission of five viruses by *Bemisia tabaci* NW2 and MEAM1. *Tropical Plant Pathology* **42**:495–499 DOI [10.1007/s40858-017-0186-9](https://doi.org/10.1007/s40858-017-0186-9).

- Dickey AM, Stocks IC, Smith T, Osborne L, McKenzie CL. 2015. DNA barcode development for three recent exotic whitefly (Hemiptera: Aleyrodidae) invaders in Florida. *Florida Entomologist* **98**:473–478 DOI 10.1653/024.098.0213.
- Dietzgen RG, Mann KS, Johnson KN. 2016. Plant virus-insect vector interactions: current and potential future research directions. *Viruses* **8**:303 DOI 10.3390/v8110303.
- Dinsdale A, Cook L, Riginos C, Buckley YM, De Barro P. 2010. Refined global analysis of *Bemisia tabaci* (Hemiptera: Sternorrhyncha: Aleyrodoidea: Aleyrodidae) mitochondrial cytochrome oxidase 1 to identify species level genetic boundaries. *Annals of the Entomological Society of America* **103**:196–208 DOI 10.1603/an09061.
- Doyle JJ, Doyle JL. 1987. A rapid DNA isolation procedure for small amounts of fresh leaf tissue. *Phytochemical Bulletin* **19**:11–15.
- Edgar RC. 2004. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* **5**:1–19 DOI 10.1186/1471-2105-5-113.
- El-Sharkawy MA. 2004. Cassava biology and physiology. *Plant Molecular Biology* **56**:481–501 DOI 10.1007/s11103-005-2270-7.
- Elena SF, Fraile A, Garcia-Arenal F. 2014. Evolution and emergence of plant viruses. *Advances in Virus Research* **88**:161–191 DOI 10.1016/B978-0-12-800098-4.00003-9.
- Fauquet C, Fargette D. 1990. African cassava mosaic virus: etiology, epidemiology and control. *Plant Disease* **74**:404–411 DOI 10.1094/PD-74-0404.
- Fereres A. 2015. Insect vectors as drivers of plant virus emergence. *Current Opinion in Virology* **10**:42–46 DOI 10.1016/j.coviro.2014.12.008.
- Fernandes FR, Albuquerque LC, Giordano LB, Boiteux LS, Ávila AC, Inoue-Nagata AK. 2008. Diversity and prevalence of Brazilian bipartite begomovirus species associated to tomatoes. *Virus Genes* **36**:251–258 DOI 10.1007/s11262-007-0184.
- Fernandes FR, Albuquerque LC, Oliveira CL, Cruz ARR, Rocha WB, Pereira TG, Naito FYB, Dias ND, Nagata T, Faria JC, Zerbini FM, Aragão FJL, Inoue-Nagata AK. 2011. Molecular and biological characterization of a new Brazilian begomovirus, euphorbia yellow mosaic virus (EuYMV), infecting *Euphorbia heterophylla* plants. *Archives of Virology* **156**:2063–2069 DOI 10.1007/s00705-011-1070-4.
- Frohlich DR, Torres-Jerez II, Bedford ID, Markham PG, Brown JK. 1999. A phylogeographical analysis of the *Bemisia tabaci* species complex based on mitochondrial DNA markers. *Molecular Ecology* **8**:1683–1691 DOI 10.1046/j.1365-294x.1999.00754.x.
- Gallet R, Michalakakis Y, Blanc S. 2018. Vector-transmission of plant viruses and constraints imposed by virus-vector interactions. *Current Opinion in Virology* **33**:144–150 DOI 10.1016/j.coviro.2018.08.005.
- Ghosh S, Bouvaine S, Maruthi MN. 2015. Prevalence and genetic diversity of endosymbiotic bacteria infecting cassava whiteflies in Africa. *BMC Microbiology* **15**:93 DOI 10.1186/s12866-015-0425-5.
- Ghosh S, Kanakala S, Lebedev G, Kontsedalov S, Silverman D, Alon T, Mor N, Sela N, Luria N, Dombrovsky A, Mawassi M, Haviv S, Czosnek H, Ghanim M. 2019. Transmission of a new polerovirus infecting pepper by the whitefly *Bemisia tabaci*. *Journal of Virology* **15**:e00488-00419.

- Gilbertson RL, Batuman O, Webster CG, Adkins S. 2015.** Role of the insect supervectors *Bemisia tabaci* and *Frankliniella occidentalis* in the emergence and global spread of plant viruses. *Annual Review of Virology* **2**:67–93
DOI [10.1146/annurev-virology-031413-085410](https://doi.org/10.1146/annurev-virology-031413-085410).
- Gleadow R, Pegg A, Blomstedt CK. 2016.** Resilience of cassava (*Manihot esculenta* Crantz) to salinity: implications for food security in low-lying regions. *Journal of Experimental Botany* **67**:5403–5413 DOI [10.1093/jxb/erw302](https://doi.org/10.1093/jxb/erw302).
- Gómez-Díaz JS, Montoya-Lerma J, Muñoz Valencia V. 2019.** Prevalence and diversity of endosymbionts in cassava whiteflies (Hemiptera: Aleyrodidae) from Colombia. *Journal of Insect Science* **19**:12 DOI [10.1093/jisesa/iez047](https://doi.org/10.1093/jisesa/iez047).
- Gutierrez S, Zwart MP. 2018.** Population bottlenecks in multicomponent viruses: first forays into the uncharted territory of genome-formula drift. *Current Opinion in Virology* **33**:184–190 DOI [10.1016/j.coviro.2018.09.001](https://doi.org/10.1016/j.coviro.2018.09.001).
- Hadjistrylli M, Roderick GK, Brown JK. 2016.** Global population structure of a world-wide pest and virus vector: genetic diversity and population history of the *Bemisia tabaci* sibling species group. *PLOS ONE* **11**:e0165105
DOI [10.1371/journal.pone.0165105](https://doi.org/10.1371/journal.pone.0165105).
- Herrera-Campo BV, Hyman G, Bellotti A. 2011.** Threats to cassava production: known and potential geographic distribution of four key biotic constraints. *Food Security* **3**:329 DOI [10.1007/s12571-011-0141-4](https://doi.org/10.1007/s12571-011-0141-4).
- Hodges GS, Evans GA. 2005.** An identification guide to the whiteflies (Hemiptera: Aleyrodidae) of the southeastern United States. *Florida Entomologist* **88**:518–534
DOI [10.1653/0015-4040\(2005\)88\[518:AIGTTW\]2.0.CO;2](https://doi.org/10.1653/0015-4040(2005)88[518:AIGTTW]2.0.CO;2).
- Inoue-Nagata AK, Albuquerque LC, Rocha WB, Nagata T. 2004.** A simple method for cloning the complete begomovirus genome using the bacteriophage phi29 DNA polymerase. *Journal of Virological Methods* **116**:209–211
DOI [10.1016/j.jviromet.2003.11.015](https://doi.org/10.1016/j.jviromet.2003.11.015).
- Jacobson AL, Duffy S, Sseruwagi P. 2018.** Whitefly-transmitted viruses threatening cassava production in Africa. *Current Opinion in Virology* **33**:167–176
DOI [10.1016/j.coviro.2018.08.016](https://doi.org/10.1016/j.coviro.2018.08.016).
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012.** Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* **28**:1647–1649
DOI [10.1093/bioinformatics/bts199](https://doi.org/10.1093/bioinformatics/bts199).
- Kumar S, Stecher G, Tamura K. 2016.** MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* **33**:1870–1874
DOI [10.1093/molbev/msw054](https://doi.org/10.1093/molbev/msw054).
- Kunz D, Tay WT, Elfekih S, Gordon KHJ, De Barro PJ. 2019.** Take out the rubbish—Removing NUMTs and pseudogenes from the *Bemisia tabaci* cryptic species mtCOI database. *BioRxiv*. 724765.

- Lee W, Park J, Lee GS, Lee S, Akimoto SI. 2013. Taxonomic status of the *Bemisia tabaci* complex (Hemiptera: Aleyrodidae) and reassessment of the number of its constituent species. *PLOS ONE* 8:e63817 DOI 10.1371/journal.pone.0063817.
- Lefevre P, Martin DP, Harkins G, Lemey P, Gray AJA, Meredith S, Lakay F, Monjane A, Lett JM, Varsani A, Heydarnejad J. 2010. The spread of tomato yellow leaf curl virus from the Middle East to the world. *PLOS Pathogens* 6:e1001164 DOI 10.1371/journal.ppat.1001164.
- Legg J, Fauquet C. 2004. Cassava mosaic geminiviruses in Africa. *Plant Molecular Biology* 56:585–599 DOI 10.1007/s11103-004-1651-7.
- Legg JP, Lava Kumar P, Makeskumar T, Tripathi L, Ferguson M, Kanju E, Ntawurhunga P, Cuellar W. 2015. Cassava virus diseases: biology, epidemiology, and management. *Advances in Virus Research* 91:85–142 DOI 10.1016/bs.aivir.2014.10.001.
- Legg JP, Sseruwagi P, Boniface S, Okao-Okuja G, Shirima R, Bigirimana S, Gashaka G, Herrmann HW, Jeremiah S, Obiero H, Ndyetabula I, Tata-Hangy W, Masembe C, Brown JK. 2014. Spatio-temporal patterns of genetic change amongst populations of cassava *Bemisia tabaci* whiteflies driving virus pandemics in East and Central Africa. *Virus Research* 186:61–75 DOI 10.1016/j.virusres.2013.11.018.
- Leotard G, Duputie A, Kjellberg F, Douzery EJ, Debain C, De Granville JJ, McKey D. 2009. Phylogeography and the origin of cassava: new insights from the northern rim of the Amazonian basin. *Molecular Phylogenetics and Evolution* 53:329–334 DOI 10.1016/j.ympev.2009.05.003.
- Liu BM, Yan FM, Chu D, Pan HP, Jiao XG, Xie W, Wu QJ, Wang SL, Xu BY, Zhou XG, Zhang YJ. 2012. Difference in feeding behaviors of two invasive whiteflies on host plants with different suitability: implication for competitive displacement. *International Journal of Biological Sciences* 8:697–706 DOI 10.7150/ijbs.4108.
- Luan J-b, Xu J, Lin K-k, Zalucki MP, Liu S-s. 2012. Species exclusion between an invasive and an indigenous whitefly on host plants with differential levels of suitability. *Journal of Integrative Agriculture* 11:215–224 DOI 10.1016/S2095-3119(12)60006-9.
- Mabvakure B, Martin DP, Kraberg S, Cloete L, Van Brunschot S, Geering AD, Thomas JE, Bananej K, Lett JM, Lefevre P, Varsani A, Harkins GW. 2016. Ongoing geographical spread of *Tomato yellow leaf curl virus*. *Virology* 498:257–264 DOI 10.1016/j.virol.2016.08.033.
- Macedo MA, Albuquerque LC, Maliano MR, Souza JO, Rojas MR, Inoue-Nagata AK, Gilbertson RL. 2018. Characterization of tomato leaf curl purple vein virus, a new monopartite New World begomovirus infecting tomato in Northeast Brazil. *Archives of Virology* 163:737–743 DOI 10.1007/s00705-017-3662-0.
- Malka O, Santos-Garcia D, Feldmesser E, Sharon E, Krause-Sakate R, Delatte H, Van Brunschot S, Patel M, Visendi P, Mugerwa H, Seal S, Colvin J, Morin S. 2018. Species-complex diversification and host-plant associations in *Bemisia tabaci*: a plant-defence, detoxification perspective revealed by RNA-Seq analyses. *Molecular Ecology* 27:4241–4256 DOI 10.1111/mec.14865.
- Mar TB, Mendes IR, Lau D, Fiallo-Olive E, Navas-Castillo J, Alves MS, Zerbini FM. 2017a. Interaction between the New World begomovirus *Euphorbia yellow*

- mosaic virus* and its associated alphasatellite: effects on infection and transmission by the whitefly *Bemisia tabaci*. *Journal of General Virology* **98**:1552–1562 DOI [10.1099/jgv.0.000814](https://doi.org/10.1099/jgv.0.000814).
- Mar TB, Xavier CAD, Lima ATM, Nogueira AM, Silva JCF, Ramos-Sobrinho R, Lau D, Zerbini FM. 2017b.** Genetic variability and population structure of the New World begomovirus *Euphorbia yellow mosaic virus*. *Journal of General Virology* **98**:1537–1551 DOI [10.1099/jgv.0.000784](https://doi.org/10.1099/jgv.0.000784).
- Marubayashi JM, Kliot A, Yuki VA, Rezende JA, Krause-Sakate R, Pavan MA, Ghanim M. 2014.** Diversity and localization of bacterial endosymbionts from whitefly species collected in Brazil. *PLOS ONE* **9**:e108363 DOI [10.1371/journal.pone.0108363](https://doi.org/10.1371/journal.pone.0108363).
- Marubayashi JM, Yuki VA, Rocha KCG, Mituti T, Pelegrinotti FM, Ferreira FZ, Moura MF, Navas-Castillo J, Moriones E, Pavan MA, Krause-Sakate R. 2013.** At least two indigenous species of the *Bemisia tabaci* complex are present in Brazil. *Journal of Applied Entomology* **137**:113–121 DOI [10.1111/j.1439-0418.2012.01714](https://doi.org/10.1111/j.1439-0418.2012.01714).
- Maruthi MN, Jeremiah SC, Mohammed IU, Legg JP. 2017.** The role of the whitefly, *Bemisia tabaci* (Gennadius), and farmer practices in the spread of cassava brown streak ipomoviruses. *Journal of Phytopathology* **165**:707–717 DOI [10.1111/jph.12609](https://doi.org/10.1111/jph.12609).
- Milenovic M, Wosula EN, Rapisarda C, Legg JP. 2019.** Impact of host plant species and whitefly species on feeding behavior of *Bemisia tabaci*. *Frontiers in Plant Science* **10**:1 DOI [10.3389/fpls.2019.00001](https://doi.org/10.3389/fpls.2019.00001).
- Minato N, Sok S, Chen S, Delaquis E, Phirun I, Le VX, Burra DD, Newby JC, Wyckhuys KAG, De Haan S. 2019.** Surveillance for Sri Lankan cassava mosaic virus (SLCMV) in Cambodia and Vietnam one year after its initial detection in a single plantation in 2015. *PLOS ONE* **14**:e0212780 DOI [10.1371/journal.pone.0212780](https://doi.org/10.1371/journal.pone.0212780).
- Montagnac JA, Davis CR, Tanumihardjo SA. 2009.** Nutritional value of cassava for use as a staple food and recent advances for improvement. *Comprehensive Reviews in Food Science and Food Safety* **8**:181–194 DOI [10.1111/j.1541-4337.2009.00077](https://doi.org/10.1111/j.1541-4337.2009.00077).
- Moraes LA, Marubayashi JM, Yuki VA, Ghanim M, Bello VH, De Marchi BR, Barbosa LF, Boykin LM, Krause-Sakate R, Pavan MA. 2017.** New invasion of *Bemisia tabaci* Mediterranean species in Brazil associated to ornamental plants. *Phytoparasitica* **45**:517–525 DOI [10.1007/s12600-017-0607-9](https://doi.org/10.1007/s12600-017-0607-9).
- Moraes LA, Muller C, Bueno R, Santos A, Bello VH, De Marchi BR, Watanabe LFM, Marubayashi JM, Santos BR, Yuki VA, Takada HM, Barros DR, Neves CG, Silva FN, Goncalves MJ, Ghanim M, Boykin L, Pavan MA, Krause-Sakate R. 2018.** Distribution and phylogenetics of whiteflies and their endosymbiont relationships after the Mediterranean species invasion in Brazil. *Scientific Reports* **8**:14589 DOI [10.1038/s41598-018-32913-1](https://doi.org/10.1038/s41598-018-32913-1).
- Morris EK, Caruso T, Buscot F, Fischer M, Hancock C, Maier TS, Meiners T, Müller C, Obermaier E, Prati D, Socher SA, Sonnemann I, Wäschke N, Wubet T, Wurst S, Rillig MC. 2014.** Choosing and using diversity indices: insights for ecological applications from the German Biodiversity Exploratories. *Ecology and Evolution* **4**:3514–3524 DOI [10.1002/ece3.1155](https://doi.org/10.1002/ece3.1155).

- Mugerwa H, Seal S, Wang H-L, Patel MV, Kabaalu R, Omongo CA, Alicai T, Tairo F, Ndunguru J, Sseruwagi P, Colvin J. 2018. African ancestry of New World, Bemisia tabaci-whitefly species. *Scientific Reports* 8:2734 DOI 10.1038/s41598-018-20956-3.
- Muhire BM, Varsani A, Martin DP. 2014. SDT: A virus classification tool based on pairwise sequence alignment and identity calculation. *PLOS ONE* 9:e108277 DOI 10.1371/journal.pone.0108277.
- Nassar N. 2001. Cassava, Manihot esculenta Crantz and wild relatives: their relationships and evolution. *Genetic Resources and Crop Evolution* 48:429–436 DOI 10.1023/a:1012005008645.
- Navas-Castillo J, Fiallo-Olivé E, Sánchez-Campos S. 2011. Emerging virus diseases transmitted by whiteflies. *Annual Review of Phytopathology* 49:219–248 DOI 10.1146/annurev-phyto-072910-095235.
- Ndunguru J, Legg J, Aveling T, Thompson G, Fauquet C. 2005. Molecular biodiversity of cassava begomoviruses in Tanzania: evolution of cassava geminiviruses in Africa and evidence for East Africa being a center of diversity of cassava geminiviruses. *Virology Journal* 2:21 DOI 10.1186/1743-422X-2-21.
- Njoroge MK, Mutisya DL, Miano DW, Kilalo DC. 2017. Whitefly species efficiency in transmitting cassava mosaic and brown streak virus diseases. *Cogent Biology* 3:1311499 DOI 10.1080/23312025.2017.1311499.
- Nylander JAA. 2004. MrModeltest v2. Program distributed by the author. Uppsala: Evolutionary Biology Centre, Uppsala University.
- Olsen KM. 2004. SNPs, SSRs and inferences on cassava's origin. *Plant Molecular Biology* 56:517–526 DOI 10.1007/s11103-004-5043-9.
- Olsen KM, Schaal BA. 1999. Evidence on the origin of cassava: phylogeography of Manihot esculenta. *Proceedings of the National Academy of Sciences of the United States of America* 96:5586–5591 DOI 10.1073/pnas.96.10.5586.
- Pan H, Chu D, Yan W, Su Q, Liu B, Wang S, Wu Q, Xie W, Jiao X, Li R, Yang N, Yang X, Xu B, Brown JK, Zhou X, Zhang Y. 2012. Rapid spread of tomato yellow leaf curl virus in China is aided differentially by two invasive whiteflies. *PLOS ONE* 7:e34817 DOI 10.1371/journal.pone.0034817.
- Pan LL, Cui XY, Chen QF, Wang XW, Liu SS. 2018. Cotton leaf curl disease: which whitefly is the vector? *Phytopathology* 108:1172–1183 DOI 10.1094/PHYTO-01-18-0015-R.
- Patil BL, Fauquet CM. 2009. Cassava mosaic geminiviruses: actual knowledge and perspectives. *Molecular Plant Pathology* 10:685–701 DOI 10.1111/j.1364-3703.2009.00559.x.
- Paz-Carrasco LC, Castillo-Urquiza GP, Lima AT, Xavier CA, Vivas-Vivas LM, Mizubuti ES, Zerbini FM. 2014. Begomovirus diversity in tomato crops and weeds in Ecuador and the detection of a recombinant isolate of rhynchosia golden mosaic Yucatan virus infecting tomato. *Archives of Virology* 159:2127–2132 DOI 10.1007/s00705-014-2046-y.

- Qin L, Pan L-L, Liu S-S. 2016.** Further insight into reproductive incompatibility between putative cryptic species of the *Bemisia tabaci* whitefly complex. *Insect Science* 23:215–224 DOI 10.1111/1744-7917.12296.
- R Core Team. 2017.** R: a language and environment for statistical computing. Available at <https://www.R-project.org/> (accessed on 14 August 2020).
- Rey C, Vanderschuren H. 2017.** Cassava mosaic and brown streak diseases: current perspectives and beyond. *Annual Review of Virology* 4:429–452 DOI 10.1146/annurev-virology-101416-041913.
- Ribeiro SG, Ávila AC, Bezerra IC, Fernandes JJ, Faria JC, Lima MF, Gilbertson RL, Zambolim EM, Zerbini FM. 1998.** Widespread occurrence of tomato geminiviruses in Brazil, associated with the new biotype of the whitefly vector. *Plant Disease* 82:830 DOI 10.1094/PDIS.1998.82.7.830C.
- Rocha CS, Castillo-Urquiza GP, Lima ATM, Silva FN, Xavier CAD, Hora-Junior BT, Beserra-Junior JEA, Malta AWO, Martin DP, Varsani A, Alfenas-Zerbini P, Mizubuti ESG, Zerbini FM. 2013.** Brazilian begomovirus populations are highly recombinant, rapidly evolving, and segregated based on geographical location. *Journal of Virology* 87:5784–5799 DOI 10.1128/JVI.00155-13.
- Rocha KCG, Marubayashi JM, Navas-Castillo J, Yuki VA, Wilcken CF, Pavan MA, Krause-Sakate R. 2011.** Only the B biotype of *Bemisia tabaci* is present on vegetables in São Paulo State, Brazil. *Scientia Agricola* 68:120–123 DOI 10.1590/S0103-90162011000100018.
- Rodríguez-Negrete EA, Morales-Aguilar JJ, Domínguez-Duran G, Torres-Devora G, Camacho-Beltrán E, Leyva-López NE, Voloudakis AE, Bejarano ER, Méndez-Lozano J. 2019.** High-throughput sequencing reveals differential begomovirus species diversity in non-cultivated plants in Northern-Pacific Mexico. *Viruses* 11:594 DOI 10.3390/v11070594.
- Rojas MR, Gilbertson RL, Russell DR, Maxwell DP. 1993.** Use of degenerate primers in the polymerase chain reaction to detect whitefly-transmitted geminiviruses. *Plant Disease* 77:340–347 DOI 10.1094/PD-77-0340.
- Ronquist F, Huelsenbeck JP. 2003.** MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19:1572–1574 DOI 10.1093/bioinformatics/btg180.
- Sacristan S, Malpica JM, Fraile A, Garcia-Arenal F. 2003.** Estimation of population bottlenecks during systemic movement of Tobacco mosaic virus in tobacco plants. *Journal of Virology* 77:9906–9911 DOI 10.1128/jvi.77.18.9906-9911.2003.
- Sambrook J, Russel D. 2001.** *Molecular Cloning - A Laboratory Manual (3a edition)*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.
- Scott IAW, Workman PJ, Drayton GM, Burnip GM. 2007.** First record of *Bemisia tabaci* biotype Q in New Zealand. *New Zealand Plant Protection* 60:264–270 DOI 10.30843/nzpp.2007.60.4601.
- Simon C, Frati F, Beckenbach A, Crespi B, Liu H, Flook P. 1994.** Evolution, weighting and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Annals of the Entomological Society of America* 87:651–701 DOI 10.1093/aesa/87.6.651.

- Sottoriva LDM, Lourenção AL, Colombo CA. 2014.** Performance of *Bemisia tabaci* (Genn.) biotype B (Hemiptera: Aleyrodidae) on weeds. *Neotropical Entomology* 43:574–581 DOI [10.1007/s13744-014-0238-5](https://doi.org/10.1007/s13744-014-0238-5).
- Sun D-B, Liu Y-Q, Qin L, Xu J, Li F-F, Liu S-S. 2013.** Competitive displacement between two invasive whiteflies: insecticide application and host plant effects. *Bulletin of Entomological Research* 103:344–353 DOI [10.1017/s0007485312000788](https://doi.org/10.1017/s0007485312000788).
- Tajebe LS, Boni SB, Guastella D, Cavalieri V, Lund OS, Rugumamu CP, Rapisarda C, Legg JP. 2015.** Abundance, diversity and geographic distribution of cassava mosaic disease pandemic-associated *Bemisia tabaci* in Tanzania. *Journal of Applied Entomology* 139:627–637 DOI [10.1111/jen.12197](https://doi.org/10.1111/jen.12197).
- Tiendrebeogo F, Lefeuvre P, Hoareau M, Harimalala MA, De Bruyn A, Villemot J, Traore VS, Konate G, Traore AS, Barro N, Reynaud B, Traore O, Lett JM. 2012.** Evolution of *African cassava mosaic virus* by recombination between bipartite and monopartite begomoviruses. *Virology Journal* 9:67 DOI [10.1186/1743-422X-9-67](https://doi.org/10.1186/1743-422X-9-67).
- Tocko-Marabena BK, Silla S, Simiand C, Zinga I, Legg J, Reynaud B, Delatte H. 2017.** Genetic diversity of *Bemisia tabaci* species colonizing cassava in Central African Republic characterized by analysis of cytochrome c oxidase subunit I. *PLOS ONE* 12:e0182749 DOI [10.1371/journal.pone.0182749](https://doi.org/10.1371/journal.pone.0182749).
- Vasquez-Ordóñez AA, Hazzi NA, Escobar-Prieto D, Paz-Jojoa D, Parsa S. 2015.** A geographic distribution database of the Neotropical cassava whitefly complex (Hemiptera, Aleyrodidae) and their associated parasitoids and hyperparasitoids (Hymenoptera). *Zookeys* 545:75–87 DOI [10.3897/zookeys.545.6193](https://doi.org/10.3897/zookeys.545.6193).
- Vyskočilová S, Seal S, Colvin J. 2019.** Relative polyphagy of Mediterranean cryptic *Bemisia tabaci* whitefly species and global pest status implications. *Journal of Pest Science* 92:1071–1088 DOI [10.1007/s10340-019-01113-9](https://doi.org/10.1007/s10340-019-01113-9).
- Walsh PS, Metzger DA, Higuchi R. 1991.** Chelex 100 as a medium for simple extraction of DNA for PCR-based typing from forensic material. *Biotechniques* 10:506–513.
- Wang D, Huang G, Shi T, Wang G, Fang R, Zhang X, Ye J. 2020.** Surveillance and distribution of the emergent Sri Lankan cassava mosaic virus in China. *Phytopathology Research* 2:18 DOI [10.1186/s42483-020-00063-w](https://doi.org/10.1186/s42483-020-00063-w).
- Wang D, Yao X, Huang G, Shi T, Wang G, Ye J. 2019.** First report of Sri Lankan cassava mosaic virus infected cassava in China. *Plant Disease* 103:1437.
- Wang HL, Cui XY, Wang XW, Liu SS, Zhang ZH, Zhou XP. 2016.** First report of Sri Lankan cassava mosaic virus infecting cassava in Cambodia. *Plant Disease* 100:1029–1029 DOI [10.1094/pdis-10-15-1228](https://doi.org/10.1094/pdis-10-15-1228).
- Watanabe LFM, Bello VH, De Marchi BR, Silva FB, Fusco LM, Sartori MM, Pavan MA, Krause-Sakate R. 2019.** Performance and competitive displacement of *Bemisia tabaci* MEAM1 and MED cryptic species on different host plants. *Crop Protection* 124:104860 DOI [10.1016/j.cropro.2019.104860](https://doi.org/10.1016/j.cropro.2019.104860).
- Watling J, Shock MP, Mongeló GZ, Almeida FO, Kater T, De Oliveira PE, Neves EG. 2018.** Direct archaeological evidence for Southwestern Amazonia as an early plant domestication and food production centre. *PLOS ONE* 13:e0199868 DOI [10.1371/journal.pone.0199868](https://doi.org/10.1371/journal.pone.0199868).

- Whitfield AE, Falk BW, Rotenberg D. 2015.** Insect vector-mediated transmission of plant viruses. *Virology* **480**:278–289.
- Wilson AK. 1981.** Euphorbia heterophylla: a review of distribution, importance and control. *Tropical Pest Management* **27**:32–38 DOI [10.1080/09670878109414169](https://doi.org/10.1080/09670878109414169).
- Zerbini FM, Briddon RW, Idris A, Martin DP, Moriones E, Navas-Castillo J, Rivera-Bustamante R, Varsani A, ICTV Consortium. 2017.** ICTV virus taxonomy profile: *Geminiviridae*. *Journal of General Virology* **98**:131–133 DOI [10.1099/jgv.0.000738](https://doi.org/10.1099/jgv.0.000738).
- Zhou X, Liu Y, Calvert L, Munoz C, Otim-Nape GW, Robinson DJ, Harrison BD. 1997.** Evidence that DNA-A of a geminivirus associated with severe cassava mosaic disease in Uganda has arisen by interspecific recombination. *Journal of General Virology* **78**:2101–2111 DOI [10.1099/0022-1317-78-8-2101](https://doi.org/10.1099/0022-1317-78-8-2101).