

Citation: Sakamoto LS, Souza LL, Gianvecchio SB, de Oliveira MHV, Silva JAlldV, Canesin RC, et al. (2021) Phenotypic association among performance, feed efficiency and methane emission traits in Nellore cattle. PLoS ONE 16(10): e0257964. https://doi.org/10.1371/journal. pone.0257964

Editor: Marcio de Souza Duarte, Universidade Federal de Viçosa, BRAZIL

Received: March 20, 2021

Accepted: September 14, 2021

Published: October 14, 2021

Copyright: © 2021 Sakamoto et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: The authors would like to thank Sao Paulo Research Foundation (FAPESP) for financial support (grant#2017/10630-2 and grant#2017/ 50339-5), and also for providing grants to LSS (grant #2018/17313-5) and LLS (grant #2019/ 11738-7). The authors also thank the Coordination for the Improvement of Higher Education **RESEARCH ARTICLE**

Phenotypic association among performance, feed efficiency and methane emission traits in Nellore cattle

Leandro Sannomiya Sakamoto¹*, Luana Lelis Souza^{1,2}, Sarah Bernardes Gianvecchio¹, Matheus Henrique Vargas de Oliveira³, Josineudson Augusto II de Vasconcelos Silva³, Roberta Carrilho Canesin¹, Renata Helena Branco¹, Melissa Baccan⁴, Alexandre Berndt⁵, Lucia Galvão de Albuquerque², Maria Eugênia Zerlotti Mercadante¹

1 Institute of Animal Science, Beef Cattle Research Center, Sertãozinho, SP, Brazil, 2 São Paulo State University (Unesp), School of Agricultural and Veterinarian Sciences, Jaboticabal, SP, Brazil, 3 São Paulo State University (Unesp), School of Veterinary Medicine and Animal Science (FMVZ), Botucatu, São Paulo, Brazil, 4 Embrapa Environment, Jaguariúna, SP, Brazil, 5 Embrapa Southeast Livestock, São Carlos, SP, Brazil

* leandrossakamoto@gmail.com

Abstract

Enteric methane (CH₄) emissions are a natural process in ruminants and can result in up to 12% of energy losses. Hence, decreasing enteric CH₄ production constitutes an important step towards improving the feed efficiency of Brazilian cattle herds. The aim of this study was to evaluate the relationship between performance, residual feed intake (RFI), and enteric CH₄ emission in growing Nellore cattle (*Bos indicus*). Performance, RFI and CH₄ emission data were obtained from 489 animals participating in selection programs (mid-test age and body weight: 414±159 days and 356±135 kg, respectively) that were evaluated in 12 performance tests carried out in individual pens (n = 95) or collective paddocks (n = 394) equipped with electronic feed bunks. The sulfur hexafluoride tracer gas technique was used to measure daily CH₄ emissions. The following variables were estimated: CH₄ emission rate (g/day), residual methane emission and emission expressed per mid-test body weight, metabolic body weight, dry matter intake (CH₄/DMI), average daily gain, and ingested gross energy (CH₄/GE). Animals classified as negative RFI (RFI<0), i.e., more efficient animals, consumed less dry matter (P < 0.0001) and emitted less g CH₄/day (P = 0.0022) than positive RFI animals (RFI>0). Nonetheless, more efficient animals emitted more CH₄/DMI and CH_4/GE (P < 0.0001), suggesting that the difference in daily intake between animals is a determinant factor for the difference in daily enteric CH₄ emissions. In addition, animals classified as negative RFI emitted less CH₄ per kg mid-test weight and metabolic weight (P = 0.0096 and P = 0.0033, respectively), i.e., most efficient animals could emit less CH₄ per kg of carcass. In conclusion, more efficient animals produced less methane when expressed as g/day and per kg mid-test weight than less efficient animals, suggesting lower emissions per kg of carcass produced. However, it is not possible to state that feed efficiency has a direct effect on enteric CH₄ emissions since emissions per kg of consumed dry matter and the percentage of gross energy lost as CH_4 are higher for more efficient animals.

Personnel (CAPES, Finance Code 001) for providing grants to SBG, and MHVO. NO.

Competing interests: The authors have declared that no competing interests exist.

Introduction

Enteric methane (CH_4) emission is a natural process in ruminants that can result in losses of 2 to 12% of the total energy consumed by the animal [1]. The variation is the result of some factors such as chemical composition of the diet, intake level [2], and even genetic [3] and meta-genomic [4].

Residual feed intake (RFI) has been used as a selection criterion in beef cattle in order to increase individual feed efficiency [5,6]. Most efficient animals (negative RFI) have a significant economic advantage as they consume less dry matter than expected for their weight and weight gain [7]. Consequently, the use of negative RFI animals has the potential to significantly reduce meat production costs.

Generally, the higher the dry matter intake (DMI), the higher the daily enteric emissions of CH_4 since a larger amount of substrate will be available for fermentation in the rumen and consequently more hydrogen will be available for methanogenesis [8,9]. Therefore, the use of more efficient animals may reduce enteric CH_4 emissions proportionally to the lower feed intake [10]. However, it is unclear whether the differences in enteric CH_4 emissions are due to the variation in digestive efficiency between negative and positive RFI animals or simply the result of from the lower DMI associated with negative RFI animals [11,12].

Inconsistencies still exist regarding the relationship between feed efficiency (RFI and FC) and enteric CH₄ emission by cattle. Studies have shown this correlation is positive and favorable in the case of highly digestible diets [10,13,14], while the phenotypic relationship between feed efficiency and enteric CH₄ emission is zero or even negative and unfavorable in diets with low digestibility [14–17], suggesting that individual enteric CH₄ emission may even increase with the improvement of feed efficiency. Furthermore, few studies have investigated *Bos indicus* animals receiving high roughage diets [16,18,19] and there is a lack of studies involving a large number of zebuine animals. The aim of the present study was to evaluate the relationship among performance, feed efficiency and enteric CH₄ emission traits in Nellore cattle (*Bos indicus*). The hypothesis was that the use of animals with low DMI and similar ADG could be a strategy to reduce greenhouse gases emissions in the beef production system.

Materials and methods

Location and animals

The data were collected in 2011, 2012, 2018, 2019 and 2020 in Sertãozinho-SP, Brazil, as well as in Botucatu-SP, Brazil, in 2019. Performance, feed efficiency and enteric CH_4 emission data were obtained from 489 Nellore animals evaluated in performance tests. This study was carried out in strict accordance with the recommendations in the Guidelines for Animal Welfare and Humane Slaughter (São Paulo State, Law Number 11.977). The protocol was approved by the Committee on the Ethics of Animal Experiments of the Institute of Animal Science (Protocol Number 278–19), Nova Odessa-SP, Brazil.

Treatments and management

The performance tests had an average duration of 76.5 ± 12 days preceded by 28 days of adaptation [5]. The animals started the test at 376 ± 164 days of age, from June to December of each year, and were kept in individual pens (n = 95) or collective paddocks equipped with electronic feed bunks (GrowSafe®, Airdrie-AB, Canada; or Intergado®, Contagem-MG, Brazil) for automated recording of individual daily feed intake (n = 394), with *ad libitum* access to diet and water. The animals were weighed at the beginning and end of the test after fasting for 14 h, or at predetermined intervals without previous fasting (Table 1).

Group	Year	Sex category	Days in test	Facility	Collector container ³	Capsule emission (mg SF ₆ /day)	No. of animals	Initial age (days)	Initial weight (kg)	No. of weight recordings
1	2011	Heifers	83	Individual pen	Canister	1.623 ± 0.08	23	294 ± 26	219 ± 28	4
2	2011	Bulls	71	Individual pen	Canister	1.405 ± 0.05	23	268 ± 24	254 ± 34	19
3	2012	Bulls	90	Individual pen	Canister	2.334 ± 0.19	24	264 ± 23	229 ± 34	13
4	2012	Heifers	85	Individual pen	Canister	1.938 ± 0.16	25	325 ± 26	261 ± 28	14
5	2018	Bulls	83	GrowSafe®	Cylinder	3.119 ± 0.27	34	347 ± 28	270 ± 46	6
6	2018	Bulls	83	GrowSafe®	Cylinder	3.145 ± 0.23	36	354 ± 25	275 ± 43	6
7	2019	Bulls	83	GrowSafe®	Cylinder	4.549 ± 0.30	60	249 ± 31	224 ± 33	6
8	2019	Bulls	56	Intergado®	Cylinder	3.471 ± 0.17	58	647 ± 36	465 ± 39	2
9	2019	Bulls	56	Intergado®	Cylinder	3.062 ± 0.09	58	667 ± 35	573 ± 48	2
10	2019	Bulls	83	GrowSafe®	Cylinder	2.471 ± 0.15	62	329 ± 24	285 ± 49	7
11	2020	Bulls	83	GrowSafe®	Cylinder	2.621 ± 0.35	42	237 ± 24	226 ± 42	7
12	2020	Bulls	83	GrowSafe®	Cylinder	2.656 ± 0.34	44	239 ± 22	221 ± 33	7

Table 1. Description of test groups for evaluating the association among performance, feed efficiency and enteric methane emission traits of Nellore (Bos indicus).

https://doi.org/10.1371/journal.pone.0257964.t001

In each test, the animals were fed a single diet that differed among the years. The diets during the performance tests consisted of silage (corn or sorghum), *Brachiaria* hay, sugar cane bagasse, meal (cottonseed, soybean or peanut), corn (ground or wet grain), citrus pulp, mineral premix, salt, ammonium sulfate, and urea (Table 2).

After pre-drying (55 ± 5 °C for 72 h), diet samples were ground in a Willey-type mill (R-TE-650 model, Tecnal Equipamentos Científicos, Piracicaba, São Paulo, Brazil) to pass a 1-mm screen and analyzed for dry matter (method 934.01), ash (method 942.05) and ether extract (method 920.39) contents following the AOAC [22] guidelines. The contents of neutral detergent fiber (NDF) and acid detergent fiber (ADF) were determined by the methodology of Mertens [23] using a Tecnal fiber analyzer (TE-149, Piracicaba, São Paulo, Brazil) using α -amylase and without sodium sulphite. The NDF and ADF were expressed exclusive of ash. The determination of crude protein (method 990.03) was performed by the Dumas method [24] based on the release of nitrogen by combustion at high temperature in pure oxygen in DUMATHERM(R) analyzer. Total carbohydrates were calculated according to the methodology described by Sniffen et al. [25]: CHOT = 100 - (CP + EE + MM); and non-fiber carbohydrates were obtained by subtracting the NDF. The gross energy (GE) determinations were performed in an adiabatic calorimetric pump of the brand IKA WERKE Model C5003 (Parr Instrument Company, Illinois, USA).

The amount of feed was adjusted weekly to guarantee daily leftovers of 5 to 10% of the total amount supplied in order to ensure *ad libitum* intake. The troughs were cleaned and leftovers were removed and discarded three times per week. Intake records were discarded when there were no feed leftovers and in the case of evidence of malfunctioning of the electronic measurement devices. Weekly samples of the ingredients were obtained for determination of the dry matter content of the diet.

Animal performance and feed efficiency

The following traits were calculated as described by Grion et al. [5] and Ceacero et al. [6]: DMI, average daily gain (ADG), metabolic body weight (BW^{0.75}), RFI, and feed conversion (FC). The DMI was obtained as the mean of all valid days during the period. The ADG was

Ingredients (% DM)		Year of performance test							
	2011	2012	2018	2019	2019	2020			
Corn silage	-	53.6	54.0	-	27.6	60.0			
Sorghum silage	-	-	-	60.0	-	-			
Brachiaria hay	44.5	10.1	-	-	-	-			
Sugar cane bagasse	-	-	10.2	-	4.89	-			
Cottonseed meal	21.4	-	-	-	-	-			
Soybean meal	-	11.6	11.7	13.0	-	13.0			
Peanut meal	-	-	-	-	8.01	-			
Ground corn	32.2	21.7	21.9	25.0	-	25.0			
Wet corn	-	-	-	-	44.6	-			
Citrus pulp	-	-	-	-	11.9	-			
Mineral premix	-	-	-	-	1.78	-			
Salt	1.45	2.28	1.70	1.75	-	1.75			
Ammonium sulfate	-	0.072	-	-	-	-			
Urea	0.45	0.648	0.49	0.25	1.16	0.25			
Forage to concentrate ration	65:45	65:45	60:40	60:40	50:50	60:40			
Nutrients									
Dry matter, %	87.4	54.4	60.5	52.4	60.0	52.9			
Crude protein, % DM	11.3	13.9	10.6	11.2	15.6	10.6			
Ash, % DM	3.74	-	3.69	4.63	-	4.08			
Ether extract, % DM	2.84	1.90	1.78	2.13	3.20	3.29			
Neutral detergent fiber, % DM	50.0	50.2	48.1	40.6	26.9	35.6			
Acid detergent fiber, % DM	31.0	22.9	30.7	24.4	-	21.3			
Gross energy, Mcal/kg	4.09	4.16	3.73	3.77	4.11	4.47			
Non-fiber carbohydrates, DM%	32.1	34.0	35.8	41.5	54.0	46.4			
Total digestible nutrients ¹ , DM%	70.5	70.2	65.9	70.2	77.0	75.1			

Table 2. Percentage of ingredients and nutrient com	position of diets offered to the animals during	g the performance test accord	rding each test group

¹Values calculated using the equation of Weiss [20]. DM: Dry matter. The diets were formulated for 0.800 kg/day in 2011 and 2012, for 1.200 kg/day in 2018, 2019 (60:40), and 2020, and for 1.700 kg/day in 2019 (50:50) [21].

https://doi.org/10.1371/journal.pone.0257964.t002

estimated by the linear regression coefficient of weights on days in test (DIT) according to the equation: $yi = \alpha + \beta x \text{ DITi} + \epsilon i$, where yi = weight of the animal in the *i*th observation; $\alpha =$ intercept representing the initial weight of the animal; $\beta =$ linear regression coefficient representing ADG; DITi = days in test in the *i*th observation; $\epsilon i =$ random error associated with each observation. The BW^{0.75} was obtained as follows: BW^{0.75} = (BWi + (0.5 DIT x ADG))^{0.75}, where BWi = initial body weight and DIT = days in test.

The RFI was calculated as the difference between observed and expected DMI, which was estimated by multiple regression of DMI on ADG and $BW^{0.75}$ within the test group [i = 1,..., 12; formed by year of birth, sex (48 females and 441 intact males), facility, site)], using the GLM procedure (SAS Inst., Inc., Cary, NC). The FC was obtained as the ratio between DMI and ADG. Mean residual gain was calculated as the difference between observed and expected ADG, which was estimated by multiple regression of ADG on DMI and BW^{0.75} within the test group, using the same procedure as described above.

Ruminal methane measurement

The modified sulfur hexafluoride (SF₆) tracer gas technique described by Deighton et al. [26] was used for methane collection. The technique uses a permeation tube or capsule



Fig 1. Animals, in a test group, with the apparatus (halter, saddle and cylinder) for methane collection by sulfur hexafluoride (SF_6) tracer gas technique.

https://doi.org/10.1371/journal.pone.0257964.g001

administered to the animal and deposited in the rumen. These capsules were calibrated and prepared specifically for each sampling year. The mean SF_6 gas emission of the capsules used during each sampling period was similar, with minimal variation in mg/day (Table 1).

Before the beginning of the sampling period, the animals were adapted to the sampling apparatus for at least seven days. Methane gas was collected for five consecutive days, with the evacuated sampling polyvinyl chloride canisters (n = 95 animals) [16,18] or stainless-steel cylinder (n = 394 animals) being changed every 24 hours (Fig 1). The gas expelled through the mouth and nostrils of the animal was aspirated under vacuum with a capillary tube fixed in a halter and connected to the collector container, which was attached to the neck of the animal (polyvinyl chloride canister) or to a saddle on the back of the animal (stainless-steel cylinder) (Table 1). Collector tubes were kept in the same environment as the animals to measure back-ground concentrations of CH_4 and SF_6 during the sampling period. After each sampling period, the collectors were sent for gas chromatography analysis and their content was diluted with pure nitrogen to determine the quantities of SF_6 and CH_4 gases. The background concentrations of CH_4 and SF_6 measured by chromatography were subtracted from the concentrations found in the evacuated sampling containers of the animals.

There was a total of 12 CH_4 sampling periods (Table 1). Of the 489 animals evaluated, samples from 481 animals could be used. The losses were due to problems with the capsules. The sampling periods were September 2011; October and December 2012; November and December 2018; June, August and October 2019, and August 2020.

A gas chromatograph (HP6890, Agilent, Wilmington, Delaware, USA) was used for the analysis of CH₄ (ppm, parts per million) and SF₆ (ppt, parts per trillion). The concentrations of CH₄ and SF₆ collected in the evacuated sampling containers were determined with a flame ionization detector at 280°C (HP-Plot Al₂O₃ M column, 30 m length × 0.53 mm i.d. × 15 µm film thickness) and an electron capture detector at 300°C (HP-Plot MoleSieve column, 30 m length × 0.53 mm i.d. × 25 µm film thickness), respectively, with two loops of 0.5 cm³ maintained at 80°C attached to 2 six-way valves. Chromatography analysis was carried out immediately after the end of the field sampling periods, which allowed the reuse of the evacuated containers in the subsequent sampling period.

Methane-related variables

Daily CH₄ emission (g/day) of each animal was obtained as the arithmetic mean of emissions on five consecutive sampling days. Enteric methane emission was also expressed as: CH₄ emission expressed per DMI (CH₄/DMI, g/kg), ADG (CH₄/ADG, g/kg), mid-test body weight (CH₄/MBW, g/kg) and BW^{0.75} (CH₄/BW^{0.75}, g/kg), residual CH₄ emission (observed CH₄ – predicted CH₄ by regression of CH₄ on DMI as described by Donoghue et al. [3]), and CH₄ emission expressed per gross energy intake (CH₄ Mcal/100 Mcal GE, as described by IPCC [27]).

Statistical analysis

The animals were classified as negative RFI (RFI<0) or positive RFI (RFI>0). The variables were analyzed using the MIXED procedure (SAS Inst., Inc., Cary, NC), fitting a model that included the fixed effect of RFI class (i = 1, 2), age of animal at the start of the performance test as covariate (linear effects), and the random effects of test group (i = 1,..., 12), in addition to the residual random effect. The relationships of CH_4 (g/day) with DMI, ADG and MBW were explored by Pearson's correlation and regression analyses using the CORR and GML procedures (SAS Inst., Inc., Cary, NC). The regression model for CH_4 (g/day) included the linear effect of DMI or ADG or MBW as covariate and the random effects of test group and residual. Statistical significance was declared when P<0.05.

Results

The mean weights (initial, mid-test and metabolic) or ADG did not differ between animals classified as negative and positive RFI (<u>Table 3</u>). The mean RFI was -0.556 and 0.565 kg DM/ day for negative and positive RFI animals, respectively, showing a difference in DMI of 1.16 kg/day between the most and least efficient animals. Animals classified as negative RFI

Trait	N	Negative RFI $(n = 246)$	Positive RFI (n = 243)	SEM	Р
Initial age (days)	489	390	389	44.0	0.5353
Initial body weight (kg)	489	317	317	34.2	0.8675
Mid-test body weight (kg)	489	353	354	11.2	0.8498
Dry matter intake (kg/day)	489	7.405	8.550	0.23	< 0.0001
Average daily gain (kg/day)	489	1.228	1.237	0.07	0.7121
Metabolic body weight (kg)	489	79.7	79.8	1.59	0.8937
RFI (kg/day)	489	-0.556	0.565	0.03	< 0.0001
Feed conversion (kg/kg)	489	6.695	7.764	0.453	< 0.0001
Residual average daily gain (kg/day)	489	0.066	-0.064	0.014	< 0.0001
CH ₄ (g/day)	481	179.7	189.8	10.1	0.0022
CH4/DMI (g/kg/day)	481	23.46	21.34	1.09	< 0.0001
CH ₄ /ADG (g/kg/day)	481	169.3	175.2	16.2	0.0724
CH ₄ /MBW (g/kg)	481	0.529	0.548	0.03	0.0096
CH ₄ /BW ^{0.75} (g/kg)	481	2.259	2.353	0.14	0.0033
CH ₄ Res (g/day)	481	4.811	-4.953	1.95	0.0004
CH ₄ /GE (%GE)	481	7.78	7.08	0.41	< 0.0001

Table 3. Mean values of performance, feed efficiency and enteric methane emission traits according to residual feed intake class of Nellore (Bos indicus).

RFI: Residual feed intake; SEM: Standard error of the mean; CH₄: Enteric methane emission; CH₄/DMI: CH₄ emission expressed per dry matter intake; CH₄/ADG: CH₄ emission expressed per average daily gain, CH₄/MBW: CH₄ emission expressed per mid-test body weight; CH₄/BW^{0.75} = CH₄ emission expressed per metabolic body weight; CH₄Res: Residual CH₄ emission; CH₄/GE: % consumed gross energy lost as CH₄.

https://doi.org/10.1371/journal.pone.0257964.t003

consumed on average 13% less DM than animals classified as positive RFI; consequently, FC and residual ADG higher for more efficient animals. There was a 5% reduction of CH_4 emission (g/day) in negative RFI animals compared to animals with positive RFI. In addition, despite a similar performance, more efficient animals emitted less methane expressed as g CH_4 /kg MBW and g CH_4 /kg BW^{0.75}. Conversely, lower CH_4 emission in relation to DMI (g CH_4 /kg DMI), lower residual CH_4 emission and a lower percentage of GE lost as CH_4 were observed in positive RFI animals compared to animals with negative RFI (Table 3).

The simple correlation coefficients of CH₄ (g/day) with DMI, ADG and MBW were 0.77, 0.70, and 0.78 (P<0.0001), respectively. Scatter plots of CH₄ (g/day) with DMI, ADG and MBW and their respective regression equations are shown in Figs 2–4. For each kg of DMI the animals emitted on average 17.5 g CH₄/day, for each kg of ADG the animals emitted on average 58.0 g CH₄/day, and for each kg of MBW the animals emitted 36.0 g CH₄/day. The regression equations of CH₄ on DMI, ADG and MBW within RFI class differed from one another (P = 0.001), accompanying the results shown in Table 3.

Discussion

In general, studies investigating the relationship between feed efficiency and enteric CH_4 emissions in cattle did not include a large number of animals, mainly because of the difficulty in measuring individual enteric CH_4 emissions in the animals, and the ones that did were conducted on *Bos taurus* (Table 4). In contrast, the present study evaluated 489 *Bos indicus* animals and enteric CH_4 was measured individually by the SF₆ tracer gas technique.

Greater reductions in enteric CH₄ emissions (15–30%) were reported in the literature for more efficient taurine animals [10,13,32] compared to the reduction of approximately 5% in the emission of zebuine animals classified as negative RFI in the present study (Table 3).





https://doi.org/10.1371/journal.pone.0257964.g002



Fig 3. Relationship between enteric methane (CH₄) emissions and average daily gain (ADG) of Nellore bulls and heifers classified as negative (triangle) or positive (circle) residual feed intake (RFI). The general linear regression equation of CH₄ on ADG was: $y = 112(\pm 13.7) + 58.0(\pm 5.31)x + residual.$

https://doi.org/10.1371/journal.pone.0257964.g003



Fig 4. Relationship between enteric methane (CH₄) emissions and mid-test body weight (MBW) of Nellore bulls and heifers classified as negative (triangle) or positive (circle) residual feed intake (RFI). The general linear regression equation of CH₄ on MBW was: $y = 35,7(\pm 13.9) + 0.43(\pm 0.030)x + residual.$

https://doi.org/10.1371/journal.pone.0257964.g004

Reference	N	Sex category	Cattle breed	Measurement technique	CH ₄ (g/day)		Р	CH ₄ /DMI (g/ kg/day)		Р
					RFI-	RFI+		RFI-	RFI+	
Nkrumah et al. [<u>13</u>]	19	Steers	Continental x British	Indirect calorimetry	135	180	< 0.05	14.0	15.5	-
Hegarty et al. [<u>10</u>]	20	Steers	Angus	SF ₆	142	190	0.01	16.3	14.7	0.37
Jones et al. [<u>14</u>]	25 48	Pregnant cows Cows	Angus	OP-FTIR	133 182	125 227	- <0.05	13.0 13.9	11.7 16.2	-
Fitzsimons et al. [28]	14	Heifers	Simmental	SF ₆	260	297	0.04	38.0	36.0	0.52
Sharma et al. [29]	6	Calves	Sahiwal	SF ₆	58.7	65.6	< 0.05	15.3	18.9	<0.05
McDonnell et al. [30]	28	Heifers	Limousin x Friesian	SF ₆	156	146	0.11	22.4	20.2	0.034
Alemu et al. [31]	16	Heifers	Crossbred	GreenFeed Respirometry chamber	203 156	222 165	0.02 0.40	27.7 26.5	28.5 26.5	0.25 0.99
Dini et al. [<u>32</u>]	16	Steers	Hereford	SF ₆	194	265	0.009	20.3	28.1	0.021
Flay et al. [33]	56	Heifers	Jersey/Holstein-Friesian	GreenFeed	253	256	0.60	22.7	20.7	<0.01
Manafiazar et al. [<u>34</u>]	314 139	Heifers Cows	Crossbred	GreenFeed	180 233	184 241	0.001 <0.001	24.1 21.1	22.7 19.2	<0.001 <0.001
Batalha et al. [19]	24	Bulls	Nellore	SF ₆	235	249	0.365	25.3	26.2	0.389

SF₆: SF₆ tracer gas technique; OP-FTIR: Open-path Fourier transform infrared spectroscopy; CH₄: Methane emission; RFI-: Negative residual feed intake.; RFI+: Positive residual feed intake.

https://doi.org/10.1371/journal.pone.0257964.t004

Results similar to those of the present study were observed in crossbred taurine heifers and cows classified as more efficient, with a daily CH_4 reduction of 2.5% and 3.7%, respectively, compared to less efficient animals [34]. Lower enteric CH_4 emissions were also reported for negative RFI Angus cows compared to positive RFI cows grazing on high-quality pasture [14]. However, there was no difference in CH_4 emissions (g/day) for animals grazing a pasture of low nutritional quality [14]. On the other hand, Freetly and Brown-Brandl [15], Velazco et al. [17], Flay et al. [33] and Batalha et al. [19] found no differences in CH_4 emissions (g/day) between more and less efficient animals.

The difference in the DMI of the animals might be responsible for the differences in daily enteric CH_4 emissions between RFI classes [9,35], which would explain the results found in the present study (Table 3). Given that they have the same body weight, same weight gain and same amount of body fat, negative RFI animals tend to emit less CH₄ per day because of lower DMI [34]. Some studies evaluating CH_4 emission in animals classified as negative and positive RFI reported differences in emissions per kg of DMI, although they found no differences in CH₄ production [30,33]. However, other studies reported lower CH₄ emissions and lower CH_4 emission per DMI in negative RFI animals [29,32]. One explanation would be a lower particle passage rate in the rumen due to differences in feeding behavior since more efficient animals spend less time feeding and therefore exhibit a higher feeding rate than positive RFI animals [19,32]. Residual feed intake is an intrinsic trait of the individual that reflects maintenance requirements [36]; thus, another explanation for the lower CH_4 emissions would be a lower energy intake of negative RFI animals compared to positive RFI animals [29], since CH_4 emission is positively associated with energy intake and differences in digestibility, CH₄ emissions, heat production and energy retention are the main factors responsible for the variation of RFI between animals [13].

Lower or equal CH_4 emission (g/day), but higher emission per kg of DMI (CH_4 /DMI) and a higher percentage of gross energy lost as CH_4 , in negative RFI animals compared to positive RFI were observed in the present study and have also been reported by other authors [30,33,34]. The reasons for these differences in CH_4 /DMI and CH_4 /GE between RFI classes are still unclear. Possible explanation is an increase in rumen organic matter degradation with consequent increase in H_2 ions availability for methanogenesis in negative RFI animals [30]. Nellore animals classified based on RFI from the same contemporary groups differed in their nutrient digestive capacity [19,37,38]. Although Magnani et al. [37] and Bonilha et al. [38] demonstrated higher digestibility of dry matter (8%), neutral detergent fiber (13 to 19%) and acid detergent fiber (11%) in positive RFI animals compared to negative RFI animals, Batalha et al. [19] found lower digestibility (4.7% to 9%) of dry matter, neutral detergent fiber and acid detergent fiber, as well as of crude protein.

The lower CH₄ emissions per kg of live and metabolic weight observed in animals classified as negative RFI (P = 0.0096 and P = 0.0033 for CH₄/MBW and CH₄/BW^{0.75}, respectively) were similar to the results reported by Nkrumah et al. [13] and Fitzsimons et al. [28]. These findings indicate that, regardless of the effects of DMI, potential selection of cattle for RFI and reduced enteric CH₄ emission is possible [16,28]. The strong relationship between CH₄ and DMI and the divergent results reported in the literature for RFI and CH₄ emissions underscore the lack of evidence of a direct effect of RFI on enteric methane emission [35].

Considering the number of animals evaluated in this study and the fact that negative and positive RFI animals were compared (and not only extreme animals), it is possible to state that CH_4 emissions per kg of live body weight were different, with lower values in more efficient animals. This confirms the hypothesis that negative RFI animals emit less CH_4 per day or per kg of live body weight or per kg of ADG and, furthermore, these animals have strong potential to emit less CH_4 per kg of carcass. Considering that live body weight in the present study is the yearling weight, and the evidence of high genetic (0.55 to 0.89) and phenotypic (0.67 to 0.72) correlation between yearling weight and carcass weight [39–41], live body weight is a real indicator of the carcass weight.

Differences in the number of methanogenics in the rumen between more and less efficient animals may explain the differences observed in the intensity of CH_4 emission, regardless of the diet supplied [42]. In fact, Lopes et al. [43] and Andrade et al. [44] reported differences in the microbial composition of fecal samples between Nellore animals classified as negative and positive RFI. However, these differences in microbial populations between negative and positive RFI cattle may be due to differences in the ruminal passage rate and digestion as a result of different DMI levels [45]. Another approach to explain the variation in CH_4 emission between more and less efficient animals independent of DMI would be to identify differences in the efficiency of feed utilization, such as nutrient absorption, appetite regulation, and cell metabolism [35].

In conclusion, more efficient animals emit less CH_4 expressed as g/day and per kg of live body weight than less efficient animals, suggesting lower emission per kg of carcass. However, it is not possible to state that RFI has a direct effect on enteric CH_4 emissions since emission per kg of consumed dry matter and the percentage of gross energy lost as CH_4 were greater for negative RFI animals.

Supporting information

S1 Table. Description of test groups for evaluating the association among performance, feed efficiency and enteric methane emission traits of Nellore (*Bos indicus*). (DOCX)

S2 Table. Percentage of ingredients and nutrient composition of diets offered to the animals during the performance test according each test group. (DOCX) S3 Table. Mean values of performance, feed efficiency and enteric methane emission traits according to residual feed intake class of Nellore (*Bos indicus*). (DOCX)

S4 Table. Studies in the literature showing the relationship between residual feed intake classes and enteric methane emission. (DOCX)

Author Contributions

Conceptualization: Leandro Sannomiya Sakamoto, Maria Eugênia Zerlotti Mercadante.

- **Data curation:** Leandro Sannomiya Sakamoto, Luana Lelis Souza, Maria Eugênia Zerlotti Mercadante.
- Formal analysis: Leandro Sannomiya Sakamoto, Maria Eugênia Zerlotti Mercadante.

Funding acquisition: Josineudson Augusto II de Vasconcelos Silva, Melissa Baccan, Lucia Galvão de Albuquerque, Maria Eugênia Zerlotti Mercadante.

Investigation: Leandro Sannomiya Sakamoto, Luana Lelis Souza, Sarah Bernardes Gianvecchio, Matheus Henrique Vargas de Oliveira, Roberta Carrilho Canesin, Melissa Baccan.

- Methodology: Leandro Sannomiya Sakamoto, Luana Lelis Souza, Alexandre Berndt, Maria Eugênia Zerlotti Mercadante.
- Project administration: Lucia Galvão de Albuquerque, Maria Eugênia Zerlotti Mercadante.
- **Resources:** Renata Helena Branco, Melissa Baccan, Alexandre Berndt, Lucia Galvão de Albuquerque.
- Supervision: Leandro Sannomiya Sakamoto, Josineudson Augusto II de Vasconcelos Silva, Maria Eugênia Zerlotti Mercadante.
- Validation: Maria Eugênia Zerlotti Mercadante.
- Visualization: Leandro Sannomiya Sakamoto, Maria Eugênia Zerlotti Mercadante.

Writing - original draft: Leandro Sannomiya Sakamoto, Maria Eugênia Zerlotti Mercadante.

Writing – review & editing: Leandro Sannomiya Sakamoto, Roberta Carrilho Canesin, Maria Eugênia Zerlotti Mercadante.

References

- Johnson KA, Johnson DE. Methane emissions from cattle. J Anim Sci. 1995; 73(8):2483–92. <u>https://doi.org/10.2527/1995.7382483x PMID: 8567486</u>.
- Warner D, Bannink A, Hatew B, van Laar H, Dijkstra J. Effects of grass silage quality and level of feed intake on enteric methane production in lactating dairy cows. J Anim Sci. 2017; 95(8):3687. <u>https://doi.org/10.2527/jas.2017.1459</u> PMID: 28805897.
- Donoghue KA, Bird-Gardiner T, Arthur PF, Herd RM, Hegarty RF. Genetic and phenotypic variance and covariance components for methane emission and postweaning traits in Angus cattle. J Anim Sci. 2016; 94(4):1438–45. https://doi.org/10.2527/jas.2015-0065 PMID: 27136003.
- Roehe R, Dewhurst RJ, Duthie C-A, Rooke JA, McKain N, Ross DW, et al. Bovine host genetic variation influences rumen microbial methane production with best selection criterion for low methane emitting and efficiently feed converting hosts based on metagenomic gene abundance. PLoS Genet. 2016; 12 (2):e1005846. https://doi.org/10.1371/journal.pgen.1005846 PMID: 26891056.
- Grion AL, Mercadante MEZ, Cyrillo JNSG, Bonilha SFM, Magnani E, Branco RH. Selection for feed efficiency traits and correlated genetic responses in feed intake and weight gain of Nellore cattle. J Anim Sci. 2014; 92(3):955–65. https://doi.org/10.2527/jas.2013-6682 PMID: 24492579.

- Ceacero TM, Mercadante MEZ, Cyrillo JNSG, Canesin RC, Bonilha SFM, de Albuquerque LG. Phenotypic and genetic correlations of feed efficiency traits with growth and carcass traits in Nellore cattle selected for postweaning weight. PLoS One. 2016; 11(8):e0161366. https://doi.org/10.1371/journal. pone.0161366 PMID: 27537268.
- Carberry CA, Kenny DA, Han S, McCabe MS, Waters SM. Effect of phenotypic residual feed intake and dietary forage content on the rumen microbial community of beef cattle. Appl Environ Microbiol. 2012; 78(14):4949–58. https://doi.org/10.1128/AEM.07759-11 PMID: 22562991.
- Beauchemin KA, McGinn SM. Methane emissions from feedlot cattle fed barley or corn diets. J Anim Sci. 2005; 83(3):653–61. https://doi.org/10.2527/2005.833653x PMID: 15705762.
- Grainger C, Clarke T, McGinn SM, Auldist MJ, Beauchemin KA, Hannah MC, et al. Methane emissions from dairy cows measured using the sulfur hexafluoride (SF₆) tracer and chamber techniques. J Dairy Sci. 2007; 90(6):2755–66. https://doi.org/10.3168/jds.2006-697 PMID: 17517715.
- Hegarty RS, Goopy JP, Herd RM, McCorkell B. Cattle selected for lower residual feed intake have reduced daily methane production. J Anim Sci. 2007; 85(6):1479–86. https://doi.org/10.2527/jas.2006-236 PMID: 17296777.
- Kelly AK, McGee M, Crews DH Jr, Fahey AG, Wylie AR, Kenny DA. Effect of divergence in residual feed intake on feeding behavior, blood metabolic variables, and body composition traits in growing beef heifers. J Anim Sci. 2010; 88(1):109–23. https://doi.org/10.2527/jas.2009-2196 PMID: 19820067.
- Lawrence P, Kenny DA, Earley B, Crews DH Jr, McGee M. Grass silage intake, rumen and blood variables, ultrasonic and body measurements, feeding behavior, and activity in pregnant beef heifers differing in phenotypic residual feed intake. J Anim Sci. 2011; 89(10):3248–61. https://doi.org/10.2527/jas. 2010-3774 PMID: 21622881.
- Nkrumah JD, Okine EK, Mathison GW, Schmid K, Li C, Basarab JA, et al. Relationships of feedlot feed efficiency, performance, and feeding behavior with metabolic rate, methane production, and energy partitioning in beef cattle. J Anim Sci. 2006; 84(1):145–53. https://doi.org/10.2527/2006.841145x PMID: 16361501.
- Jones FM, Phillips FA, Naylor T, Mercer NB. Methane emissions from grazing Angus beef cows selected for divergent residual feed intake. Anim Feed Sci Technol. 2011; 166–167:302–7. https://doi. org/10.1016/j.anifeedsci.2011.04.020.
- Freetly HC, Brown-Brandl TM. Enteric methane production from beef cattle that vary in feed efficiency. J Anim Sci. 2013; 91(10):4826–31. https://doi.org/10.2527/jas.2011-4781 PMID: 23965389.
- Mercadante MEZ, Caliman AP de M, Canesin RC, Bonilha SFM, Berndt A, Frighetto RTS, et al. Relationship between residual feed intake and enteric methane emission in Nellore cattle. Rev Bras Zootec. 2015; 44(7):255–62. http://dx.doi.org/10.1590/S1806-92902015000700004.
- Velazco JI, Herd RM, Cottle DJ, Hegarty RS. Daily methane emissions and emission intensity of grazing beef cattle genetically divergent for residual feed intake. Anim Prod Sci. 2017; 57(4):627–35. <u>https://doi.org/10.1071/AN15111</u>.
- Oliveira LF, Ruggieri AC, Branco RH, Cota OL, Canesin RC, Costa HJU, et al. Feed efficiency and enteric methane production of Nellore cattle in the feedlot and on pasture. Anim Prod Sci. 2018; 58 (5):886. https://doi.org/10.1071/AN16303.
- Batalha CDA, Morelli M, Branco RH, Cyrillo JNSG, Carrilho RC, Mercadante MEZ, et al. Association between residual feed intake, digestion, ingestive behavior, enteric methane emission and nitrogen metabolism in Nellore beef cattle. Anim Sci J. 2020; 91(1):e13455. <u>https://doi.org/10.1111/asj.13455</u> PMID: 33025683.
- Weiss WP, editor. Energy prediction equations for ruminant feeds. Proceedings of the 61st Cornell Nutrition Conference for Feed Manufactures (Cornell University, Ithaca) pp 176–85; 1999.
- 21. NRC. Nutrient requirements of beef cattle. 7th rev. ed. Natl. Acad. Press, Washington, DC. 2000.
- AOAC. Association of Official Analytical Chemistry. Official methods of analysis. 15th ed. Assoc. Off. Anal. Chem., Arlington, VA. 1990.
- Mertens D.R. Gravimetric determination of amylase-treated neutral detergent fiber in feeds with refluxing in beakers or crucibles: collaborative study. J AOAC Int. 2002; 85(6):1217–40. PMID: 12477183.
- Etheridge RD, Pesti GM, Foster EH. A comparison of nitrogen values obtained utilizing the Kjeldahl nitrogen and Dumas combustion methodologies (LECO CNS 2000) on samples typical of an animal nutrition analytical laboratory. Anim Feed Sci Technol. 1998; 73:21–8. https://doi.org/10.1016/S0377-8401(98)00136-9.
- Sniffen CJ, O'Connor JD, Van Soest PJ. A net carboydrate and protein for evaluating catlle diets. II. Carbohydrate and protein availability. J Anim Sci. 1993; 70:3562–77. https://doi.org/10.2527/1992. 70113562x PMID: 1459919.

- Deighton MH, Williams SRO, Hannah MC, Eckard RJ, Boland TM, Wales WJ, et al. A modified sulphur hexafluoride tracer technique enables accurate determination of enteric methane emissions from ruminants. Anim Feed Sci Technol. 2014; 197:47–63. https://doi.org/10.1016/j.anifeedsci.2014.08.003.
- IPCC (2006) Emissions from livestock and manure management, chapter 10. In: Guidelines for National Greenhouse Gas Inventories, Vol 4, Prepared by the National Greenhouse Gas Inventories Programme (eds Eggleston HS, Buendia L, Miwa K, Ngara T, Tanabe K), 10.7–10.84. IGES, Japan.
- Fitzsimons C, Kenny DA, Deighton MH, Fahey AG, McGee M. Methane emissions, body composition, and rumen fermentation traits of beef heifers differing in residual feed intake. J Anim Sci. 2013; 91 (12):5789–800. https://doi.org/10.2527/jas.2013-6956 PMID: 24146149.
- Sharma VC, Mahesh MS, Mohini M, Datt C, Nampoothiri VM. Nutrient utilisation and methane emissions in Sahiwal calves differing in residual feed intake. Arch Anim Nutr. 2014; 68(5):345–57. https:// doi.org/10.1080/1745039X.2014.951193 PMID: 25156936.
- McDonnell RP, Hart KJ, Boland TM, Kelly AK, McGee M, Kenny DA. Effect of divergence in phenotypic residual feed intake on methane emissions, ruminal fermentation, and apparent whole-tract digestibility of beef heifers across three contrasting diets. J Anim Sci. 2016; 94(3):1179–93. https://doi.org/10.2527/ jas.2015-0080 PMID: 27065279.
- Alemu AW, Vyas D, Manafiazar G, Basarab JA, Beauchemin KA. Enteric methane emissions from lowand high-residual feed intake beef heifers measured using GreenFeed and respiration chamber techniques. J Anim Sci. 2017; 95(8):3727. https://doi.org/10.2527/jas.2017.1501 PMID: 28805902.
- Dini Y, Cajarville C, Gere JI, Fernandez S, Fraga M, Pravia MI, et al. Association between residual feed intake and enteric methane emissions in Hereford steers. Transl anim sci. 2019; 3(1):239–46. <u>https:// doi.org/10.1093/tas/txy111</u> PMID: 32704795.
- Flay HE, Kuhn-Sherlock B, Macdonald KA, Camara M, Lopez-Villalobos N, Donaghy DJ, et al. Hot topic: Selecting cattle for low residual feed intake did not affect daily methane production but increased methane yield. J Dairy Sci. 2019; 102(3):2708–13. https://doi.org/10.3168/jds.2018-15234 PMID: 30639015.
- Manafiazar G, Baron VS, McKeown L, Block H, Ominski K, Plastow G, et al. Methane and carbon dioxide emissions from yearling beef heifers and mature cows classified for residual feed intake under drylot conditions. Can J Anim Sci. 2020; 100(3):522–35. https://doi.org/10.1139/cjas-2019-0032.
- Kenny DA, Fitzsimons C, Waters SM, McGee M. Invited review: Improving feed efficiency of beef cattle-the current state of the art and future challenges. Animal. 2018; 12(9):1815–26. https://doi.org/10. 1017/S1751731118000976 PMID: 29779496.
- Arthur JPF, Herd RM. Residual feed intake in beef cattle. Rev Bras Zootec. 2008; 37(spe):269–79. http://dx.doi.org/10.1590/S1516-35982008001300031.
- Magnani E, Nascimento CF, Branco RH, Bonilha SFM, Ribeiro EG, Mercadante MEZ. Relações entre consumo alimentar residual, comportamento ingestivo e digestibilidade em novilhas Nelore. Bol Ind Anim. 2013; 70(2):187–94. https://doi.org/10.17523/bia.v70n2p187.
- Bonilha SFM, Branco RH, Mercadante MEZ, Cyrillo JNSG, Monteiro FM, Ribeiro EG. Digestion and metabolism of low and high residual feed intake Nellore bulls. Trop Anim Health Prod. 2017; 49(3):529– 35. https://doi.org/10.1007/s11250-017-1224-9 PMID: 28124731.
- Meyer K, Johnson DJ, Graser H-U. Estimates of the complete genetic covariance matrix for traits in multi-trait genetic evaluation of Australian Hereford cattle. Aust J Agric Res. 2004; 55(2):195–210. https://doi.org/10.1071/AR03164.
- Bergen R, Miller SP, Wilton JW. Genetic correlations among indicator traits for carcass composition measured in yearling beef bulls and finished feedlot steers. Can J Anim Sci. 2005; 85(4):463–73. https://doi.org/10.4141/A05-013.
- Tonussi RL, Espigolan R, Gordo DGM, Magalhães AFB, Venturini GC, Baldi F, et al. raits in Nellore cattle. Genet Mol Res. 2015; 14(4):18713–19. https://doi.org/10.4238/2015.December.28.20 PMID: 26782521.
- Carberry CA, Waters SM, Kenny DA, Creevey CJ. Rumen methanogenic genotypes differ in abundance according to host residual feed intake phenotype and diet type. Appl Environ Microbiol. 2014; 80 (2):586–94. https://doi.org/10.1128/AEM.03131-13 PMID: 24212580.
- 43. Lopes DRG, La Reau AJ, Duarte M de S, Detmann E, Bento CBP, Mercadante MEZ, et al. The bacterial and fungal Microbiota of Nelore steers is dynamic across the gastrointestinal tract and its fecal-associated Microbiota is correlated to feed efficiency. Front Microbiol. 2019; 10:1263. <u>https://doi.org/10.3389/fmicb.2019.01263 PMID: 31293524</u>.
- Andrade BGN, Bressani FA, Cuadrat RRC, Tizioto PC, de Oliveira PSN, Mourão GB, et al. The structure of microbial populations in Nelore GIT reveals inter-dependency of methanogens in feces and rumen. J Anim Sci Biotechnol. 2020; 11(1):6. https://doi.org/10.1186/s40104-019-0422-x PMID: 32123563.

45. Freetly HC, Lindholm-Perry AK, Hales KE, Brown-Brandl TM, Kim M, Myer PR, et al. Methane production and methanogen levels in steers that differ in residual gain. J Anim Sci. 2015; 93(5):2375–81. https://doi.org/10.2527/jas.2014-8721 PMID: 26020333.