

INTERNATIONAL COCOA RESEARCH CONFERENCE

CONFERENCE INTERNATIONALE SUR LA RECHERCHE CACAOYERE

CONFERENCIA INTERNACIONAL DE PESQUISAS EM CACAU

CONFERENCIA INTERNACIONAL DE INVESTIGACION EN CACAO



9 - 14 OCTOBER 2000 SHANGRI-LA'S. TANJUNG ARU RESORT HOTEL KOTA KINABALU, SABAH MALAYSIA

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Biological Faculty Institute of Applied Botany

Anatomical and Physiological Characteristics of Theobroma spec. Seeds and their Relevance to Processing

S. Müller¹, C. Rohsius¹, C. Reisdorff¹, L. Gasparotto², R. Lieberei¹ (1) University of Hamburg, Germany (2) EMBRAPA Amazônia Ocidental, Manaus, Brazil





Seed Protease Activities



Objective

The Origin of Cocoa Aroma Fermentation



Discussion

Our studies show that T bicolor and T grandiflorum seeds generally meet the biochemical requirements for the generation of aroma precursors comparable to those of cocca seeds. Fermented T. grandiflorum and T. bicolor seeds are of a light colour comparable with those of Criollio varieties. Like those, they might be used for the manufacture of light coloured chocolates.

However, simple application of the fermentation practices known from cocoa will not yield sufficient aroma precursors, since structural differences especially of the seed shells may influence the duration in which the acetic acid infiltrates the seeds. Additionally, the distinct contents of phenolic compounds of T. bicolor seeds most likely result in a differing flavour development during fermentation.

Appropriate fermentation procedures need to be developed for the production of chocolate-like products of a high and reproducible quality from seeds of T. bicolor and T. grandiflorum. First studies are being performed to test new fermentation technologies meeting the particular requisites of seeds of T. grandifiorum.

For further Information please refer to our full paper in the Conference Proceedings or contact us: S. Mueller, team cocce (Prof. Dr. R. Liebetel) Institute of Applied Botany, University of Hamburg, Marseiller Straße 7, 20355 Hamburg, Germany. Fon.: 0049-40-42838-2335, Fax: 0049-40-428386593, email: silke mueller@iangbotumi-hamburg.de

S. MUELLER, C. ROHSIUS, C. REISDORFF, L. GASPAROTTO, R. LIEBEREI Institute of Applied Botany, University of Hamburg Marseiller Strasse 7, 20355 Hamburg, Germany Silke.Mueller@iangbot.uni-hamburg.de

Anatomical and Physiological Characteristics of *Theobroma spec.* Seeds and their Relevance to Processing

1. Introduction

All 22 species of the genus *Theobroma* L. produce berry fruits with large seeds embedded in a pulp. Nevertheless, only *Theobroma cacao* L. became a main tropical crop plant due to its seeds which are processed to raw cocoa, the basic substance of chocolate and related products.

Besides *T. cacao* the seeds of at least 12 other *Theobroma* species are used in the Amazon Basin for the production of home-made chocolate (Cuatrecasas 1964). Among these, *T. grandiflorum* (WILLD. EX SPRENG) SCHUM. and *T. bicolor* H. B. K. are considered to be the most important species.

T. grandiflorum, the cupuaçu tree, is an excellent crop for locally adapted land use systems in the Amazon region. The fruit pulp fetches relatively high market prices while the seeds are not yet commercially used.

T. bicolor is commonly cultivated throughout all humid tropical America. Its pulp is used similar to the one of *T. grandiflorum*, while in some regions the seeds are utilised for the production of chocolate for local markets (CUATRECASAS 1964).

The unique chocolate aroma is formed in the course of fermentation and roasting of the seeds. During fermentation acetic acid is formed by microbial degradation of the fruit pulp. After permeating the seed shell, the acid penetrates the cotyledons where it effects a disintegration of the cellular compartments. A continuous aqueous phase is generated by the subsequent fusion of the cotyledonar liposomes. Within this phase the lowering of the pH activates two types of proteases, which corporately digest the major cocoa storage protein, a vicilin-type globulin. The resulting mixture of amino acids and oligopeptides represents essential aroma precursors.

During drying and roasting the aroma precursors react with reducing sugars to Maillard-products, forming the proper cocoa aroma (BIEHL et *al.* 1982, 1985, 1993, BIEHL and PASSERN 1982, VOIGT et *al.* 1994a, 1994b, 1994c, 1994d, VOIGT and BIEHL 1995, ZIEGLEDER and BIEHL 1988).

Phenols form another group of flavour compounds, causing an astringent and bitter taste. During fermentation, cocoa phenols leak from the vacuoles and tan significant portions of the surrounding proteins (BIEHL et *al.* 1983). The function of the resulting products as flavour components is disputed (ZIEGLEDER and Biehl 1988). However, the tanning prevents large amounts of storage proteins from proteolytic cleavage and thus affects the flavour development.

Besides the physiological aspects the success of fermentation depends on the histological and cytological structure of the seeds. Both the permeability of the

seed coat to acetic acid and the specific characteristics of the cotyledon tissue are to be mentioned in this context (LOPEZ et al. 1987).

The objective of our studies was to find out whether the seeds of T. grandiflorum and T. bicolor have the potential to develop a good chocolate-like aroma and thus can be used to produce a storable and valuable ware. With this intention, we studied physiological and anatomical features relevant to flavour development of the seeds of the two Theobroma species and compared them with the characteristics of T. cacao.

2. Materials and Methods

2.1 Samples

Ripe fruits of T. cacao of the Forastero variety, T. grandiflorum and T. bicolor were obtained from the experimental field and the surroundings of the German-Brazilian project SHIFT near Manaus, Brazil. The Theobroma seeds were analysed at the Institute of Applied Botany, University of Hamburg, Germany,

2.2 Microscopical Studies

Entire seeds were cut into slices and fixed in 4 % formaldehyde. Tissue samples of seed coats and cotyledons were dehydrated first in 60 %, then in 100 % ethanol and subsequently embedded in LR White ® Medium Grade. Semithin sections (1-2 µm) were fixed on a slide. After incubating in 12 % sodium hypochloride they were dyed with 0,05 % aq. Toluidine Blue and Lugol's Solution to stain protein bodies, starch grains, polyphenols and cell walls (GUTMANN 1995, modified).

The semithin sections were analysed with an Olympus BH-2 light microscope.

2.3 Quantification of Total Phenolics

After removal of the seed coat the embryos were shock frozen and freeze dried. Samples of 5 g were ground with 20 ml hexane until the particle size was reduced to less than 0,04 mm. Most of the residual seed fat was extracted by flushing the solids with petrol ether in a Buchner funnel.

The phenolic compounds were extracted by agitating 0.5 g of the fat-free sample on ice once with 80 % aq. acetone and twice with 60 % aq. acetone (50 ml respectively). After centrifugation the supernatants were combined in a flask containing 5 ml of acetic acid. The acetone was removed by rotary evaporation under partial vacuum at 40 °C. The aqueous residue was brought to 200 ml in a volumetric flask. Total phenolics were determined by the Folin-Ciocalteu procedure (SINGLETON and ROSSI 1965, modified).

2.4 Preparation of Acetone Dry Powder (acdp)

The fat of freeze dried and ground seed material was extracted exhaustively with petrol ether in a Twisselman apparatus. The phenolic compounds were removed by agitating twice with 80 % aq. acetone and once with 70 % aq. acetone containing 5 mm sodium ascorbate. After centrifugation the solids containing the proteins and proteases were dried to an anhydrous powder (acetone dry powder, acdp) by extracting twice with 100 % acetone and subsequent removal of the solvent under vacuum.

2.5 Analysis of the Seed Proteins

Total protein content was measured by Biuret (COOPER, 1981).

In order to obtain the albumin fraction, the acdp was extracted in a low salt buffer (0.01 mol/l NaCl, 0.05 mol/l TRIS/HCl pH 7.5, 2 mmol/l Na₂-EDTA, 5 mmol/l sodium ascorbate, 5 μ g/ml pepstatin A). The precipitate was extracted in a high salt buffer (0.5 mol/l NaCl, 0.2 mol/l TRIS/HCl pH 8.0, 5 μ g/ml pepstatin A) to obtain the globulin fraction.

For SDS gel electrophoresis, portions of 10 mg acdp or fractions were diluted in 1.5 ml SDS sample buffer containing 0.0625 mol/l Tris/HCl, pH 6.8, 2 % SDS, 5 % mercaptoethanol, 10 % glycerine and 0.001 % Bromophenol blue.

The electrophoresis was conducted using BIO-RAD ® 10-20 % polyacrylamide gradient ready gels. The protein bands were stained with Coomassie Brilliant Blue or with silver staining methods.

2.6 Characterization of the Proteases

The aspartic endoprotease was extracted from the acdp according to HANSEN et. *al.* 1998. After centrifugation, aliquots of the supernatant were incubated at 35° C both in presence and absence of 20 µg/ml pepstatin A in McIlvaine buffer, pH 3,5, containing 4 mg/ml BSA as substrate.

The seryl exoprotease was extracted with 20 mmol/l Na₂HPO₄ containing 0,5 mol/l NaCl, 0,5 % Triton X-100 and 20 g/l Pepstatin A (pH 6,8). After centrifugation, aliquots of the supernatant were incubated both in presence and absence of 1 mmol/l phenyl methane sulfonyl fluoride (PMSF) in McIlvaine buffer pH 5,6 containing 5 mmol/l N-benzyl-oxycarbonyl dipeptides as substrates.

The proteolytic reactions were stopped by adding a solution of trichloro acetic acid to a final concentration of 4 % (w/v).

The amount of liberated α -NH₂ groups was measured in the supernatants according to SHUTOV et *al.* 1992. The activities were determined as mol α -NH₂ liberated per minute and g fresh weight.

2.7 Seed Incubation Experiment

Fermentation-like seed incubation was performed during 5 days with 1 % acetic acid in glass vessels in a water bath at 40 °C.

Every day, three seeds of each species were sampled and sliced. The pH of the intersections was measured by means of a pH electrode with a flat membrane (Mettler Toledo InLab ®).

3. Results

3.1 The Structure of the Seed Shells

The *Theobroma* seed shells species consist of two integuments, of which the outer one is much thicker than the inner. Its predominantly collapsed parenchyma contains vascular bundles and, beneath the outer epidermis, large chambers filled with slime. Significant differences between the three examined *Theobroma* species were found in the thickness of the seed shells that ranges between 385 μ m (*T. cacao*) and 1670 μ m (*T. bicolor*).

The parenchyma of the inner integument consists of only a few cell layers. Its outer epidermis is composed of a single layer of thick-walled, lignified cells.

The thickness of the outer epidermis of the inner integument also varies significantly between the three species. In the seed coat of *T. cacao* the epidermal cells form a sclereide layer that is about 9 μ m thick. The homologue palisade layers of the two other examined species are 30 μ m (*T. grandiflorum*) and 130 μ m (*T. bicolor*) thick.

3.2 The Structure of the Cotyledons

The cotyledonar tissue of *Theobroma* seeds mainly consists of storage cells which contain about 60 % fat, 20 % proteins and 5-10 % starch in separated compartments.

Besides the storage cells, polyphenol cells amount to a portion of 8-11 % of the cotyledonar cells of *T. cacao* and *T. grandiflorum*. Due to anthocyanins, these cells cause the purple colour of Forastero-type cocoa seeds but they are colourless in cupuaçu seeds. Apart from a higher frequency beneath the vascular bundles, the polyphenol cells are evenly distributed in the cotyledonar tissues of *T. cacao* and *T. grandiflorum* seeds. In some sections they form small groups and rows which mainly appear close to the upper epidermis of the cotyledons.

The seed tissue of *T. bicolor* is virtually free of polyphenol cells, which appear sporadically beneath the vascular bundles.

3.3 Total Phenolics

While the phenolics of *T. cacao* and *T. grandiflorum* cotyledons amounted to 7-7,5 % of freeze-dried and defatted sample, their portion was only 0,5 % in *T. bicolor*. These results correlate with the few polyphenol cells found in *T. bicolor* seed tissue.

3.4 Seed Proteins

The patterns of seed proteins of all three *Theobroma* species are marked by two predominant globulins and at least one predominant albumin. The molecular weights of the two classes of globulins are similar for all three species $(46,5 \pm 1 \text{ kDa} \text{ and } 30,3 \pm 2 \text{ kDa})$. Considering the quantity of globulins, the seeds of *T. bicolor* are prominent regarding their high content of the 46 kDa storage protein, while only the seeds of *T. grandiflorum* contain more albumins than globulins. However, in all three species the relative quantity of the 30 kDa globulin vary around 15 % of total protein content.

3.5 Incubation Experiment

Before incubation *T. grandiflorum* seeds showed a pH of $5,7 \pm 0,3$ whereas a pH of $5,9 \pm 0,3$ was measured at the intersections of *T. cacao* and *T. bicolor* seeds. During incubation the pH was declining in all of the species. Seed samples drawn after 4 days of incubation indicated pH values of $4,78 \pm 0,1$ (*T. cacao*), $4,71 \pm 0,3$ (*T. grandiflorum*) and $5,1 \pm 0,3$. (*T. bicolor*).

During seed incubation, 70% of the total protein content of T. cacao seed was degraded after two days whereas even after four days a decrease of only 15% resp. 3% was measured in *T. grandiflorum* and *T. bicolor* seed.

3.6 Proteolytic Activities

The seeds of all three examined Theobroma species contain pepstatin-A sensitive endoproteolytic activities, which gives rise to classify the respective enzymes as aspartic endoproteases. All of these activities are considerably high but the one of cocoa is twice as high as of *T. bicolor* and *T. grandiflorum* (fig.). The pH-dependency of the aspartic proteases is similar in all three species with an optimum range at pH 3.4-3.6.

In seeds of *T. bicolor* and *T. grandiflorum* we found PMSF-sensitive carboxy exopeptidases of an activity equal to the respective enzyme of cocoa seeds (Fig.). Thus, the particular enzymes of all three species are to be classified as seryl type carboxypeptidases. The pH optimum of the enzymes ranges from pH 5.4-5.7.



Fig: Proteolytic Activities in Cotyledons of three *Theobroma* **Species a:** Pepstatin-A sensitive activity of aspartic endoproteases incubated at pH 3.5 with 4 mg/ml BSA **b:** PMSF-sensitive activities of seryl carboxypeptidases incubated at pH 5.6 with different z-dipeptides

4. Discussion

Our studies show that *T. bicolor* and *T. grandiflorum* seeds generally meet the biochemical requirements for the generation of aroma precursors comparable to those of cocoa seeds. The quantitative and qualitative characteristics of the seed globulins and the cotyledonar proteases are similar in all three species. The slight differences regarding the proteolytic activities and the globulin contents are not likely to influence the potential yield of aroma precursors. Furthermore, due to broad range of intraspecific variation of *T. grandiflorum* and *T. bicolor*, there should exist proveniences with higher globulin and protease contents. Fermented cupuaçu and *T. bicolor* seeds are of a light colour and so they might be used for the manufacture of light coloured chocolates.

However, simple application of the fermentation practices known from cocoa will not yield sufficient aroma precursors, since structural differences especially of the seed shells may influence the duration in which the acetic acid infiltrates the seeds. In fact, both the decrease in pH and the generation of flavour precursors are delayed in *T. bicolor* seeds which may be due to the comparatively thick seed shell with its massive palisade layer.

Additionally, the distinct contents of phenolic compounds of *T. bicolor* seeds most likely result in a differing flavour development during fermentation.

Our results show that appropriate fermentation procedures need to be developed for the production of chocolate-like products of a high and reproducible quality from seeds of *T. bicolor* and *T. grandiflorum*. First studies are being performed to test new fermentation technologies meeting the particular requirements of seeds of *T. grandiflorum*.

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