Biodiversity of Prokaryotes from High-Elevation Grassland Organosols of the Parana State, Brazil

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The High-Elevation Grasslands of the Atlantic Mountain Range, and the First, Second and Third Plateau cover 14% of the Parana State area and contain hydromorphic soils consisting primarily of organic matter, called organosols. In this work, the Atlantic Mountain Range and Second Plateau organosols, with large geographical distance, distinct geological history and similar ecological roles, were selected for prokaryotic diversity investigation. Total DNA of the both soils was extracted and used as template with specific oligonucletides primers for the amplification of *nifH*, a marker gene for diazotrophs and 16S rRNA, for Bacteria. Analysis of RFLP profiles of nifH and 16S rRNA amplicons revealed that the Atlantic organosol had higher total bacterial diversity and lower diazotrophic diversity than the Second Plateu organosol. 16S rRNA gene libraries from both organosols were constructed and sequenced. Analysis with the RDPII database showed the predominance of the Acidobacteria phylum in both environments (~ 45%). Moreover, in the Atlantic Mountain Range's library the following phyla was also found: Proteobacteria (21%), Bacteriodetes (11%), Firmicutes (6%), Actinobacteria (2%) and unclassified (15%). In the Second Plateau's library, the diversity was restricted to Proteobacteria (15%), Choroflexi (1%) and unclassified (39%). The highest diversity in the Atlantic Mountain Range is consistent with the profile obtained in RFLP analysis. These results show that despite the floristic similarities, the bacterial communities of the analyzed organosols present structural differences.

Key words: biodiversity of prokaryotes; high-elevation grassland organosols; *nifH*; 16S rRNA.

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ABSTRACT

The High-Elevation Grasslands of the Atlantic Mountain Range, and the First, Second and Third Plateaus cover 14% of the Paraná State area and contain hydromorphic soils consisting primarily of organic matter, called organosols. In this work, the Atlantic Mountain Range and Second Plateau organosols, with large geographical distance, distinct geological history and similar ecological roles, were selected for prokaryotic diversity investigation. Total DNA of the both soils was extracted and used as templates with specific primers for the amplification of niffH, a marker gene for diazotrophs and 165 rDNA, for bacteria. Cluster analysis of RFLP profiles of niffH and 165 rDNA amplicons revealed that the 20 cm deep sample collected in the Second Plateau (CG20) and the 50 cm deep (CG50) collected in the same region are closer to each other than to the 20 cm deep sample collected in the Atlantic Mountain Range (SM20). 165 rDNA gene libraries of the three samples were constructed and 328 clones were sequenced. The comparison of the clone sequences with the RDP II database of 165 rDNA indicated predominance of Acidobacteria followed by the Proteobacteria phylum in the threes samples. Community analysis using SONS indicated a 5 OTU_{0.03} overlap among the libraries. S-Libshuff analysis yielded no significant differences between the CG20 and CG50 communities (P = 0.2678), which is consistent with the cluster analysis of RFLP profiles. These results showed that, despite the edaphic factor and floristic similarity, the bacterial communities of the sample SM20 and CG30 are structurally different. Moreover, the validation of the clusters by library sequence analysis showed that the molecular marker RFLP is a rapid and efficient tool for studies of prokaryotic community.

GENERAL OBJECTIVES

Investigate the bacterial and diazotrophic diversity of distinct horizon organosols from the High-Elevation Grassland of the Parana State.

SPECIFIC OBJECTIVES

- Extraction of the total DNA of organosols collected 20 cm (SM20) deep in the Atlantic Mountain Range, and collected 20 cm (CG20) and 50 cm (CG50) deep in the Second Plateau;
 Comparison of the bacterial and diazotrophic community
- Comparison of the bacterial and diazotrophic community structures found in SM20, CG20 and CG50 by RFLP;
 Comparison of the bacterial diversity found in SM20, CG20 and
- CG50 by 16S rDNA library sequencing.

MATERIAL AND METHODS

DNA EXTRACTION AND AMPLIFICATION. DNA was extracted using the Ultraclean Soil DNA kit (MO BIO Laboratories, Carlsbad, CA). The PCR conditions and the specific primer sequences for the amplification of 165 rDNA, and nifth were established based on LANE (1991) and ROSCH et al. (2002).

RFLP ANALYSIS. 500ng of the amplified 165 rDNA or nifH from the collected samples was digested with Hinfl, or Haelll restriction enzymes at 37°C for 5 h. Digested products were separated by native polyacrylamide gel electrophoresis (12%) and visualized after ethidium bromide staining under UV light. The similarity dendogram was estimated through a binary matrix of present and absent DNA bands using the computer program NTSYSON (Numerical Taxonomy of multivariate Systems) version 2.1 (Applied Biostatistics Inc). The dendograms were built by the algorithm UPGMA.

1.65 rDNA LIBRARY. 165 rDNA PCR products of 1.5 kb were cloned into the p-GEM-T vector (Promega) and transformed into *E. coli* DH5 α cells, previously treated with 100 mM ice-cold CaCl2. Several transformants were randomly picked, and the plasmids were extracted by alkaline lysis (SAMBAOOK et al., 1989). 576 clones were sequenced in a *MegaBACE* 1000 DNA automated sequencer with the primer 27F.

SEQUENCE ANALYSIS. Nucleotide sequences (reads) were trimmed for base quality. Sequences were aligned with clustal W 1.8 and a distance matrix calculated by the kimura method using DNAdist (both programs running under BIOEDIT 7.08 package). A total of 328 sequences spanning the V2-V3 region of 165 rDNA were obtained from SM20 (131), CG20 (103), and CG50 (94) samples.

OTU DEFINITION and TAXONOMIC AFFILIATION. Sequences of gene libraries were clustered into OTUs using the program DOTUR (SCHLOSS and HANDELSMAN, 2005) and compared against Ribosomal Database Project II database using the SEQUENCEMATCH program for taxonomic affiliation.

LIBRARY COMPARISONS. Comparisons were performed using the program SONS, which implements nonparametric estimators (SCHLOSS and HANDELSMAN, 2006) and S-LIBSHUFF which uses the Cramer von-Mises statistic. S-LIBSHUFF calculates P values by using a random-permutation provisionificance of overlapping co al., 2004).

RESULT

RFLP ANALYSES OF 165 FDNA SHOWED THAT THE CG50 AND CG20 BACTERIAL COMMUNITIES ARE STRUCTURALLY CLOSER TO EACH OTHER THAN SM20



Figure 1 - 185 rDNA fingerprinting of SM20, CG20, and CG90 bacterial communities of organosols obtained after digestion with Hinfi restriction enzyma. The three independent assays are represented. M: 10tho DNA marker.



Figure 2 - Similarity dendrogram of the 165 rDNA Ingerprinting obtained siter digestion with the restriction entrymes Heelii and HMI, generated by program NTSVS pc 2.1, using jaccard's similarity coefficient and the UPSMA algorithm.

RFLP ANALYSES OF *nith* SHOWED THAT THE CG50 AND CG20 DIAZOTROPHIC COMMUNITIES ARE STRUCTURALLY CLOSER TO EACH OTHER THAN SM20



Figure 3 - mfH fingerprinting of SM20, CG20 and CG30 bacterial communities of organisols obtained after digestion with the Haelli restriction enzyme. The three independent assays are represented. M: 100h 0104 merker.



Figura 4 - Similarity dendrogram of nifth fingerprinting obtained after digestion with the restriction enzymes Haelli and Hinfl. generated by program NTSYS pc 2.1. using Jaccard's similarity coefficient and the UPSMA algorithm

THE THREE LIBRARIES OF 165 rDNA GENE SHOWED A HIGH RICHNESS WITH DOMINANCE OF THE ACIDOBACTERIA CLASS

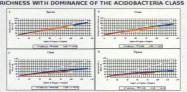


Figure 5 - Rarefaction curves of the 165 rDNA libraries, generated with a cut-off value of 0.03 for species definition (A.). 0.05 for genus (B.). 0.15 for class (C) and 0.2 phylum (D). Data were generated with

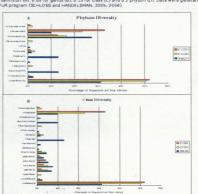


Figure 6 - 165 rDNA gene sequences were compared with sequences in the Ribosomal Databa Project II (released 10 April 3, 2009) for taxonomic affiliation at the level of phylum IA) and class (6 using the program Sequence Match. Sequences showing less than 80% identity to known batten



COMPARISONS OF BACTERIAL COMMUNITY MEMBERSHIPS SHOWED THAT THE 16S IDNA LIBRARIES CG20 AND CG50 ARE STRUCTURALLY CLOSER TO EACH OTHER THAN SM20



	Shwed-Chaol	Issues	Tourse	0	0 4
SM2G-CC20	55.5	0,1935	0,01116	0.159	0,05963
SM20-CCM	13	0.1684	0,1674	0,037	0.02433
C030-C030	192.3	0,3877	6.346	0,100	0,04021

Figure 2 – Venn diagram companing the OTU₄₅₀ memberships found in the SM20 (F = 331), CG20 (in = 103) and CG50 (in = 84) libraries Fixed OTU-6000 (in = 84

COMPARISON OF THE 165 FONA LIBRARIES SHOWED NO STATISTICAL DIFFERENCE BETWEEN SAMPLES CG20 AND CG50

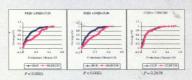


Figure 8 - A partial plot of the companions between 165 rRNA gene libraries generated by the S-LIBS-IUFF program. P values are reported

CONCLUSIONS

- The geographical distance between High-Elevation Grasslands seems to influence the structural differences of bacterial and diazotrophic communities, more than does the sampled horizon depth;
- The three analyzed samples of High-Elevation Grassland organosols showed predominance of the Acidobacteria phylum, followed by Proteobacteria;
- The validation of the clusters by library sequence analysis showed that the molecular marker RFLP is a rapid and efficient tool for studies of prokaryotic community structures.

REFERENCES

- LANE, D. J. 165/23S rRNA sequencing. In STACKEBRANDT, E., GOODFELLOW, M. (Eds.). Nucleic Acid Techniques in Bacterial Systematics: New York (Wiley), p. 115-148, 1991.
 RÖSCH, C.; MERGEL, A.; BOTHE, H. Biodiversity of
- RÖSCH, C.; MERGEL, A.; BOTHE, H. Biodiversity of denitrifying and dinitrogen-fixing bacteria in an acid forest soil. Appl. Environ. Microbiol. v. 68, p. 3818-3829, 2002.
- SAMBROOK, J.; FRITSCH, E.F.; MANIATIS, T. Molecular cloning: a laboratory manual. 2ed. Cold Spring Harbor, New York, Cold Spring Harbor Laboratory Press., 1989.
- SCHLOSS, P. D.; HANDELSMAN, J. Introducing DOTUR, a computer program for defining operational taxonomic units and estimating species richness. Appl. Environ. Microbiol., v. 71, p. 1501-1506, 2005.
 SCHLOSS, P. D.; HANDELSMAN, J. Introducing SONS, a
- SCHLOSS, P. D.; HANDELSMAN, J. Introducing SONS, a tool for operational taxonomic unit-based comparisons of microbial community memberships and structures.
 Appl. Environ. Microbiol., v. 72, p. 6773-9, 2006.
 SCHLOSS, P. D.; LARGET, B. R.; HANDELSMAN, J.
- SCHLOSS, P. D.; LARGET, B. R.; HANDELSMAN, J. Integration of microbial ecology and statistics: a test to compare gene libraries. Appl. Environ. Microbiol., v. 70, p. 5485-5492, 2004.
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