# Physiological and quality attributes associated with different centrifugation times of baby carrots

Celso Luiz Moretti<sup>1\*</sup>; Leonora Mansur Mattos<sup>1</sup>; Cristina MM Machado<sup>1</sup>; Ricardo Alfredo Kluge<sup>2</sup>

Embrapa Hortaliças; C. postal 218, 70359-970 Brasília-DF, Brazil; 2USP, ESALQ, 13418-900 Piracicaba-SP, Brazil; moretti@cnph.embrapa.br

### **ABSTRACT**

Centrifugation is one of the most important steps in the freshcut industry. Inadequate centrifugation can lead to increased white blush in baby carrots. The present work was carried out aiming at evaluating the effects of different centrifugation times in baby carrots physiological and quality attributes. Carrot roots cv. Alvorada were harvested and minimally processed as baby carrots. After processing, samples were placed in nylon bags and centrifuged (378 rad. s<sup>-1</sup>) for 0; 30; 60; 90, and 120 seconds. Temperature of baby carrots centrifuged for 120 seconds was 63% higher than the temperature at the beginning of the experiment. Respiratory activity increased 49% when centrifugation time increased from 30 to 120 seconds. Ethylene evolution remained around 1.7 µL kg<sup>-1</sup>h<sup>-1</sup> until 60 seconds, increasing to 3.5 µL kg<sup>-1</sup> h<sup>-1</sup> at 120 seconds of centrifugation. Whiteness index increased 34% and 68% when centrifugation time shifted from 30 s to 60 s and from 30 s to 120 s, respectively. No significant changes in total carotenoids content were observed for the different tested centrifugation intervals. Baby carrots should be centrifuged for 30 seconds in order to maintain the quality and to avoid the development of white blush.

**Keywords:** *Daucus carota*, total carotenoids; color; postharvest; temperature; white blush.

#### **RESUMO**

# Atributos fisiológicos e de qualidade associados com tempos de centrifugação de mini-cenouras

A centrifugação é uma das etapas mais importantes na indústria de processamento mínimo. A centrifugação inadequada pode levar a um aumento do esbranquiçamento em mini-cenouras. O presente trabalho teve como objetivo avaliar os efeitos de diferentes tempos de centrifugação nas características fisiológicas e de qualidade em minicenouras. Raízes de cenouras cv. Alvorada foram colhidas e minimamente processadas como minicenouras. Após o processamento, as amostras foram colocadas em sacos de nylon e centrifugadas (378 rad. s<sup>-1</sup>) por 0; 30; 60; 90 e 120 segundos. A temperatura das minicenouras centrifugadas por 120 segundos foi 63% mais alta que a temperatura do início do experimento. A atividade respiratória aumentou 49% quando o tempo de centrifugação aumentou de 30 s para 120 s. A evolução de etileno permaneceu em torno de  $1.7~\mu L~kg^{-1}~h^{-1}$  até 60~segundos, aumentando para  $3.5~\mu L~kg^{-1}$ <sup>1</sup> h<sup>-1</sup> aos 120 segundos de centrifugação. O índice de esbranquiçamento aumentou 34% e 68% quando o tempo de centrifugação alterou de 30 s para 60 s e de 30 para 120 s, respectivamente. Nenhuma mudança significativa no teor de carotenóides totais foi observada para os diferentes intervalos de centrifugação testados. As minicenouras devem ser centrifugadas por 30 minutos para manter a qualidade e evitar o desenvolvimento do esbranquiçamento.

**Palavras-chave:** *Daucus carota*, carotenóides totais, cor, pós-colheita, temperatura, esbranquiçamento.

## (Recebido para publicação em 16 de julho de 2007; aceito em 19 de novembro de 2007)

The consumption of minimally processed fruits and vegetables has increased significantly during the last years, considering both retail and consumer level. Many factors are associated with this trend, such as the decrease in family size, population aging, decrease in the number of family members, and increase in the foodservice sector (Moretti, 2007).

The obtention of fresh-cut products involves many steps, such as cleaning, washing, trimming, coring, slicing, shredding, and other related operations. The main objective is to provide fresh, healthy, ready-to-eat food that, in most cases, does not need further preparation to be consumed (Rolle & Chism, 1987). Examples of fresh-cut products include

peeled potatoes, shredded cabbage and lettuce, salad mixes, washed and trimmed spinach, broccoli florets and diced onions. These products can maintain their quality for up to 14 days, when stored in the optimum conditions.

The physiology and biochemistry of fresh-cut products is quite similar to the same metabolic events observed in fresh fruits and vegetables that have been mechanically injured. Essentially, the physiology of fresh-cut fruits and vegetables is the physiology of a wounded tissue (Brecht, 1995). This is easily understandable once minimal processing involves the occurrence of mechanical damages during preparation procedures. Different metabolic changes have been reported in mechanically

injured tissues, such as increase in carbon dioxide and ethylene evolution (Moretti *et al.*, 1998), alteration in aroma volatile profiles (Moretti *et al.*, 2002), and increase in the activity of many enzymes related with browning (Ke & Saltveit, 1989).

Among the various steps associated with minimal processing, centrifugation has a major importance, once it is desirable that this process should remove at least the same amount of water retained by the product during sanitation and rinsing. Centrifugation is generally used, although other methods such as vibration screens and forced air tunnels can be used as well. For lettuce products, removal of slightly more moisture (i.e., slight desiccation of the

product) may favor longer postprocessing life (Cantwell, 2000). For most of the centrifuges in the market, the angular speed and time of centrifugation are established according to the type of the product and to the degree of processing (Darezzo, 2000). Setting time and speed of centrifugation is a major problem once overcentrifuged tissues tend to have their commercial quality altered. However, it is not unusual to observe processors under or over centrifuging products what can significantly affect the quality of the final product.

For baby carrots, excessive water removal during centrifugation can cause a severe loss in quality due to the development of white blush. According to different authors, white blush appears in fresh-cut carrots due to the desiccation of cellular remnants on the carrot surface (Tatsumi et al., 1991) and to the synthesis of lignin. In many cases, white blush is the limiting factor in marketing the product, once consumers tend to associate it with decay. Acceptability of baby carrots in grocery stores and salad bars are significantly affected when the sticks have white blush (Tatsumi et al., 1993).

Scientific papers published during the last two decades have focused different physiological and quality changes associated with fresh-cut baby carrots and the possible strategies to circumvent the existing problems. However, there is a lack in the literature concerning the relation among centrifugation time and changes in quality attributes.

The present work was carried out aiming at evaluating the effects of different centrifugation times in freshcut baby carrots physiological and quality attributes.

#### MATERIAL AND METHODS

Carrot roots cv. Alvorada were harvested at commercial fields in Brasilia, Brazil. After harvest, roots were taken to the postharvest laboratory, selected for external blemishes, graded for size (18±2 cm long), and minimally processed as baby carrots.

Roots were pre-washed in tap water, hydro cooled at 5°C, and processed as

baby carrots sticks. Samples of 1.5 kg were placed in nylon bags, sanitized (NaClO, 200 mg L<sup>-1</sup>, at  $5\pm1^{\circ}$ C), rinsed (NaClO, 3 mg L<sup>-1</sup>,  $5\pm1^{\circ}$ C) and centrifuged (378 rad. s<sup>-1</sup>) in a stainless steel centrifuge (radius = 0.14 m; F = 60 Hz) for 0; 30; 60; 90, and 120 seconds. The water tension generated by the centrifuge, considering the parameters described above, at its maximum speed, was equal to -2.8 MPa, according to the equation presented in Tyree (1997).

After each centrifugation time, the following variables were assessed: mass loss, temperature, carbon dioxide and ethylene evolution, color (L\*a\*b\*) and total carotenoids content. Temperature was evaluated in carrots sticks surface with an infrared thermometer. Carbon dioxide and ethylene evolution were determined in a gas chromatograph. Column, injector, and detector temperatures were set at 60; 100 and 140°C, respectively, for CO, analysis and at 60; 100 and 150°C for ethylene quantification. The results were expressed in mL CO, kg-1 h-1 and µL C<sub>2</sub>H<sub>4</sub> kg<sup>-1</sup> h<sup>-1</sup>, respectively. Color was evaluated with a hand colorimeter (L\*a\*b\*) and whiteness index calculated according to Bolin & Huxsoll (1991), using the following equation: Whiteness index =  $100 - [(100 - L)^2 +$  $a^2 + b^2$ <sup>1/2</sup>. Total carotenoids were assayed as described by Lime et al. (1957) and Umiel & Gabelman (1971).

Analysis were performed using a completely randomized design, with five treatments and eight replicates (n = 1.5 kg). Data were subjected to analysis of variance and the least significant difference procedure was carried out. Differences between any two treatments larger than the sum of two standard deviations were always significant (P>0.05).

# RESULTS AND DISCUSSION

Considering the different centrifugation times tested, 30 seconds were enough to remove water in excess, and the mass of the product after centrifugation was similar to the mass after processing. Centrifugation for periods higher than 30 seconds

contributed to excessive tissue dehydration. During the first 30 seconds water removal was done easily when compared to the subsequent intervals (Figure 1A). Water removal in the present work was done easily when compared to the work carried out by Silva et al. (2002). They worked on fresh-cut cabbage and verified that 10 minutes of centrifugation were necessary to remove water in excess, considering a centrifuge with an angular speed of 150 rad s<sup>-1</sup>. This is partially explained by the highest hydraulic conductivity observed in roots (carrots) than leaves (cabbage).

Centrifugation or other procedures are recommended for most fresh-cut items for complete water removal or, in some cases, to cause slight desiccation of the surface (Cantwell & Suslow, 2002). The major objective is primarily to reduce microbial growth. In detached fruits and vegetables desiccation can also induce the production of stress ethylene (Yang, 1985).

Temperature is an important factor governing different postharvest processes. In the present experiment, the temperature of baby carrots showed a tendency to increase as centrifugation times increased (Figure 1B). Carrots sticks centrifuged for 120 seconds showed a temperature that was 63% higher when compared to the temperature of the material at the beginning of the experiment (Figure 1B). This heating probably occurred due to the friction suffered by the processed tissue against the centrifuge walls or even due to the heating of the equipment itself. The observed rise in temperature induced alteration in both carbon dioxide and ethylene evolution.

Similar results were verified by Cantwell (1992) for cabbage and by Artés *et al.* (1999) for tomatoes. The same phenomenon was observed by Watada *et al.* (1996) for distinct freshcut products stored under different temperatures. Increased temperature can also contribute to the loss of nutritional value of fresh-cut product, as observed for vitamin C of vegetable crops stored under different temperatures (Favell, 1998).

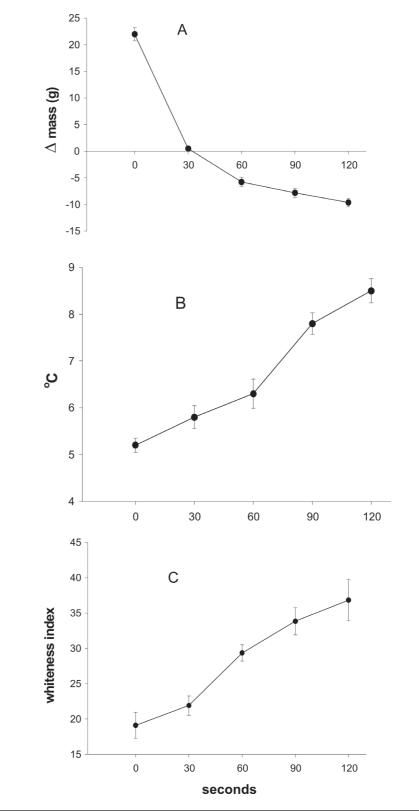
White blush is a serious defect in baby carrots. It was demonstrated in the

present investigation that increasing centrifugation time contributed to a higher level of white blush. In fact, whiteness index increased 34 and 68% when centrifugation time increased from 30 s to 60 s and from 30 s to 120 s, respectively, showing that over centrifuging baby carrots significantly contributed to the increasing of cells desiccation (Figure 1C).

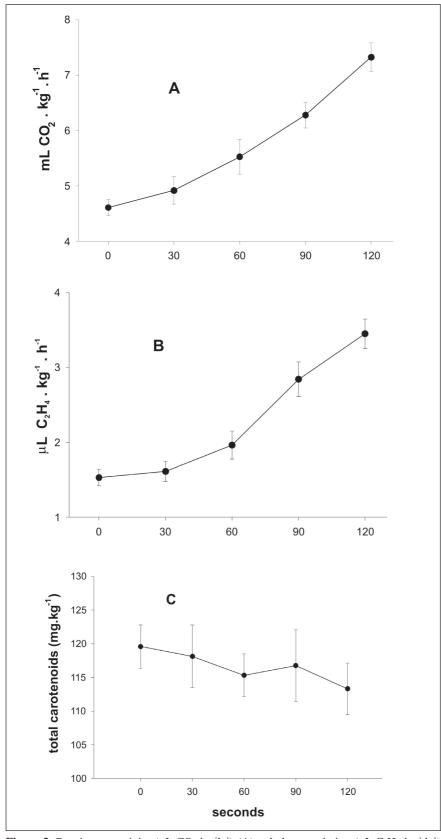
In several published studies, white blush is considered a result of both surface dehydration and enzymatic activity to form lignin (Tatsumi et al., 1991, 1993; Avena-Bustillos et al., 1993). According to different results, white blush formation was reported to be influenced by temperature, relative humidity (Avena-Bustillos et al., 1993), degree of peeling (Bolin & Huxsol, 1991) and type of cutting surface (Tatsumi et al., 1991, 1993; Bolin & Huxsol, 1991). A possible strategy to minimize or to solve the problems related to white blush in baby carrots is the application of edible coatings. The utilization of edible coatings to increase water vapor resistance aiming to reduce white blush have been tested by different authors (Avena-Bustillos et al., 1993; Sargent et al., 1994). Avena-Bustillos et al. (1994) verified that an edible sodium caseinate/stearic acid emulsion also controlled white blush and has also contributed to reduce respiration by about 20% when compared to the uncoated control.

Respiratory activity had a significant rise as centrifugation time increased. Carbon dioxide evolution increased around 49% when centrifugation time shifted from 30 s to 120 s (Figure 2A). A significant rise in CO<sub>2</sub> evolution was observed after 30 seconds, what is probably explained by both the excessive dehydration stress suffered by the tissue (Figure 1A) and the mechanical damage caused by the impacts and compression of baby carrots against the centrifuge walls.

The increase in respiration rate verified in the present investigation is one of the earliest physiological responses to mechanical stresses and can be linked to the induction of the phenolic metabolism and the wound healing response of the tissue. Different signals



**Figure 1.** Mass variation ( $\Delta g$ ) (A); temperature (°C) (B); and whiteness index (C) of freshcut baby carrots centrifuged for different times intervals ( $\Delta g = Mass$  of the product after centrifugation – mass of the product right after processing). Vertical bars mean  $\pm$  SD. (Mass variation ( $\Delta g$ ) (A); temperature (°C) (B); and whiteness index (C) of fresh-cut baby carrots centrifuged for different times intervals ( $\Delta g = Massa$  do produto após centrifugação – massa do produto logo após o processamento). Barras verticais significam  $\pm$  SD). Brasília, Embrapa Hortaliças, 2007.



**Figure 2.** Respiratory activity (μL  $CO_2$  kg<sup>-1</sup>h<sup>-1</sup>) (A); ethylene evolution (μL  $C_2H_4$  .kg<sup>-1</sup>.h<sup>-1</sup>) (B); and total carotenoids content (mg kg<sup>-1</sup>) (C) of fresh-cut baby carrots centrifuged for different times intervals. Vertical bars mean  $\pm$  SD (atividade respiratória (mL  $CO_2$  kg<sup>-1</sup> h<sup>-1</sup>) (A); evolução do etileno (mL  $C_2H_4$  kg<sup>-1</sup> h<sup>-1</sup>) (B); e teor total de carotenóides (mg kg<sup>-1</sup>) (C) de mini-cenouras centrifugadas em diferentes intervalos. Barras verticais significam  $\pm$  SD). Brasília, Embrapa Hortaliças, 2007.

induced by mechanical stresses elicit physiological and biochemical responses in both adjacent and distant tissues (Ke & Saltveit, 1989). Mechanical stresses can also induce increase in respiration rate in some plant tissues, which is probably related to alfaoxidation of fatty acids (Shine & Stumpf, 1974). This reaction oxidizes fatty acids to  $\mathrm{CO}_2$  and it is associated with  $\mathrm{CO}_2$  releasing after potato tubers slicing (Rolle & Chism, 1987).

Ethylene evolution significantly increased during the experiment (Figure 2b), remaining around 1.7  $\mu$ L kg<sup>-1</sup> h<sup>-1</sup> until 60 seconds of centrifugation. The evolution of this hormone for the material centrifuged for 120 seconds was 214% higher than that observed for baby carrots centrifuged for 30 seconds (Figure 2b).

The increase in ethylene evolution observed is directly associated with the mechanical damage suffered by the tissue. Wound induced ethylene is associated with the onset of senescence of different products (Abeles et al., 1992). Different authors verified that ethylene evolution resulting from minimal processing was enough to induce chlorophyll degradation in spinach (Spinacia oleracea L.); however, no effects were verified in broccoli (Brassica oleracea L. var. italica). The observed effects in freshcut spinach may be correlated with an increase in chlorophyllase activity due to the ethylene rise (Watada et al., 1990; Yamauchi & Watada, 1991). The consequent reduction in chlorophyll level allows the revelation of carotenoids pigments, changing the product color (Heaton & Marangoni, 1996).

Carotenoid pigments are relatively stable in their natural environment, but postharvest treatments or processing operations may contribute to increase pigments degradation (Rigal *et al.*, 2000). Possible reasons for carotenoids losses in fresh tissues are autoxidation that occurs when pigments combine with oxygen in the air or are mediated through enzymatic oxidation, which is catalyzed by oxidative enzymes (Gross, 1991). The abrasion of carrot surfaces exposes the phloem, where carotenes are

most concentrated, leading to the direct contact with air and light (Li & Barth, 1998).

No significant changes in total carotenoids content were observed for the different centrifugation intervals tested (Figure 2C), indicating that, apparently, the stress caused by the mechanical damage and desiccation was not severe enough to stimulate significant pigment degradation, considering that baby carrots were evaluated right after centrifuging. Further investigations should focus on the effect of different centrifugation times in the content of total carotenoids during refrigerated storage periods.

Mechanical stresses associated with centrifugation increased product temperature, carbon dioxide and ethylene evolution. Centrifugation for more than 30 seconds significantly contributed to tissue dehydration and the development of white blush. Considering the conditions this experiment was carried, it is suggested that baby carrots should be centrifuged for 30 seconds in order to maintain the quality and to avoid the development of white blush.

#### **ACKNOWLEDGEMENTS**

This research was supported by PRODETAB (Embrapa/World Bank Agreement), Grant n°. 019-01/02.

#### REFERENCES

- ABELES FB; MORGAN PW; SALTVEIT JR ME. 1992. *Ethylene in plant biology*. 2ed. San Diego: Academic Press. 414 p.
- ARTÉS F; CONESA MA; HERNÁDEZ S; GIL MI. 1999. Keeping quality of fresh-cut tomato. *Postharvest Biology and Technology* 17: 153-162.
- AVENA-BUSTILLOS RJ; CISNEROS-ZEVALLOS LA; KROCHTA JM; SALTVEIT ME. 1993. Optimization of edible coatings on minimally processed carrots using response surface methodology. *American Society of Agricole Engineers* 36: 801.
- AVENA-BUSTILLOS RJ; CISNEROS-ZEVALLOS LA; KROCHTA JM; SALTVEIT ME. 1994. Application of casein-lipid edible film emulsions to reduce white blush on minimally processed carrots. *Postharvest Biology and Technology* 4: 319-329.

- BOLIN HR; HUXSOLL CC. 1991. Control of minimally processed carrot (*Daucus carota* L.) surface discoloration caused by abrasion peeling. *Journal of Food Science* 56: 416.
- BRECHT JK. 1995. Physiology of lightly processed fruits and vegetables. *HortScience* 30: 18-22.
- CANTWELL MI. 1992. Postharvest handling systems: minimally processed fruits and vegetables. In: KADER AA (Ed.). Postharvest technology of horticultural crops. 2ed., Davis: University of California, Division of Horticultural and Natural Resources. p 273-281
- CANTWELL MI. 2000. Preparation and quality of fresh-cut produce. In: PUSCHMANN R (Ed.). Encontro Nacional sobre Processamento Mínimo de Frutas e Hortaliças, 2. *Proceedings...* Viçosa: UFV. p. 156-182.
- CANTWELL MI; SUSLOW TV. 2002. Postharvest handling systems: Fresh-cut fruits and vegetables. In: KADER AA (Ed.). Postharvest technology of horticultural crops. 3ed. Publ. 3311. Oakland: University of California, Division of Agriculture and Natural Resources. p. 445-463.
- DAREZZO HM. 2000. Fresh-cut lettuce (*Lactuca sativa* L.). In: PUSCHMANN R (Ed.). Encontro Nacional Sobre Processamento Mínimo de Frutas e Hortaliças, 2. *Palestras...* Viçosa: UFV. p. 117-127.
- FAVELL DJ. 1998. A comparison of the vitamin C content of fresh and frozen vegetables. *Food Chemistry* 62: 59-64.
- GROSS J. 1991. *Pigments in vegetables*. New York: AVI, Van Nostrand Reinhold. 335 p.
- HEATON JW; MARANGONI AG. 1996. Chlorophyll degradation in processed foods and senescent plant tissues. *Trends in Food Science and Technology* 7: 8-15.
- KE D; SALTVEIT ME. 1989. Wound induced ethylene production, phenolic metabolism, and susceptibility to russet spotting in iceberg lettuce. *Plant Physiology* 76: 412-418.
- LI P; BARTH MM. 1998. Impact of edible coatings on nutritional and physiological changes in lightly processed carrots. *Postharvest Biology and Technology* 14: 51-60.
- LIME BJ; GRIFFITHS FP; O'CONNOR RT; HEINZELMANN DC; MCCALL ER. 1957. Spectrophotometric methods for determining pigmentation - beta-carotene and lycopene in ruby red grapefruit. Agricultural and Food Chemistry 5: 941-944.
- MORETTI CL. 2007. Panorama do processamento mínimo de frutas e hortaliças. In: MORETTI CL (Ed.). Manual de Processamento Mínimo de Frutas e Hortaliças. Brasília: SEBRAE. p. 25-40.
- MORETTI CL; BALDWIN E; SARGENT SA; HUBER DJ. 2002. Internal bruising alters aroma volatile profiles in tomato fruit tissues. *HortScience* 37: 378-382.

- MORETTI CL; SARGENT SA; HUBER DJ; CALBO AG; PUSCHMANN R. 1998. Chemical composition and physical properties of pericarp, locule and placental tissues of tomatoes with internal bruising. *Journal of the American Society for Horticultural Science* 123: 656-660.
- RIGAL D; GAUILLARD F; FORGET FL. 2000. Changes in the carotenoid content of apricot (*Prunus armeniaca* var Bergeron) during enzymatic browning: beta-carotene inhibition of chlorogenic acid degradation. *Journal of Science and Food Agriculture* 80: 763-768.
- ROLLE R; CHISM GW. 1987. Physiological consequences of minimally processed fruits and vegetables. *Journal of Food Quality* 10: 157-165.
- SARGENT SA; BRECHT JK; ZOELLNER JJ; BALDWIN EA; CAMPBELL CA. 1994. Edible films reduce surface drying of peeled carrots. *Proceedings of Florida State Horticultural Society* 107: 245-247.
- SILVA EO; MORETTI CL; CARNELOSSI MAG; PUSCHMANN R; CAMPOS RS; CARDOSO RAL. 2002. Quality attributes associated with different centrifugation times in fresh-cut cabbage. *Proceedings of Florida State Horticultural Society* 115: 114-117.
- TATSUMI Y; WATADA AE; WERGIN WP. 1991. Scanning electron microscopy of carrot stick surface to determine cause of white translucent appearance. *Journal of Food Science* 56: 1357-1362.
- TATSUMI Y; WATADA AE; LING PP. 1993. Sodium chloride treatment or waterjet slicing effects on white tissue development of carrot sticks. *Journal of Food Science* 58: 1390-1392.
- TYREE MT. 1997. The Cohesion—Tension theory of sap ascent: current controversies. *Journal of Experimental Botany* 48: 1753-1765.
- UMIEL N; GABELMAN WH. 1971. Analytical procedures for detecting carotenoids of carrot (Daucus carota L.) roots and tomato (Lycopersicon esculentum) fruits. Journal of the American Society for Horticultural Science 96: 702-704.
- YAMAUCHI N; WATADA AE. 1991. Regulated chlorophyll degradation in spinach leaves during storage. *Journal of American Society for Horticultural Science* 116: 58-62.
- YANG SF. 1985. Biosynthesis and action of ethylene. *HortScience* 20: 41-45.
- WATADA AE; ABE K; YAMUCHI N. 1990. Physiological activities of partially processed fruits and vegetables. Food Technology 44: 116-122.
- WATADA AE; KO NP; MINOTT DA. 1996. Factors affecting quality of fresh-cut horticultural products. *Postharvest Biology and Technology* 9: 115-126.