Full Length Research Paper

Growth, phosphorus status, and nutritional aspect in common bean exposed to different soil phosphate levels and foliar-applied phosphorus forms

Fabrício William Ávila¹*, Valdemar Faquin¹, Allan Klynger da Silva Lobato², Danielle Pereira Baliza³, Douglas José Marques¹, Alexandre Martins Abdão dos Passos⁴, Carla Elisa Alves Bastos⁵ and Elaine Maria Silva Guedes²

¹Departamento de Ciência do Solo, Universidade Federal de Lavras, Lavras, Brazil.
²Núcleo de Pesquisa Vegetal Básica e Aplicada, Universidade Federal Rural da Amazônia, Paragominas, Brazil.
³Instituto Federal de Educação, Ciência e Tecnologia do Sudeste de Minas Gerais, Rio Pomba, Brazil.
⁴Empresa Brasileira de Pesquisa Agropecuária/Rondônia, Porto Velho, Brazil.
⁵Departamento de Ciência do Solo, Escola Superior de Agricultura Luiz de Queiroz/USP, Piracicaba, Brazil.

Accepted 30 May, 2012

This study aimed to investigate the effect of the foliar application of phosphite and phosphate on growth, phosphorus (P) status, and nutritional aspect of common bean (*Phaseolus vulgaris* cv. Radiante) plants grown under different soil phosphate levels. Experiment was organized in factorial scheme completely randomized using 2 soil phosphate levels (Pi-starved and Pi-sufficient plants), combined with 3 nutrient sources supplied via foliar application (KH₂PO₃, KH₂PO₄, and KCI used as control), and 2 foliar application numbers (single and two applications). In this study were measured root dry weight, shoot dry weight, and root to shoot ratio, as well as shoot P concentration, root P concentration, accumulated P in shoot, accumulated P in root, P uptake efficiency, P utilization efficiency, P translocation, and macro and micronutrients in shoot. Common bean growth under limiting phosphate availability in soil exhibited lower biomass yield and higher concentration of nutrients in shoot tissues. The results exhibit foliar-applied KH₂PO₄ were not sufficient to affect in phosphate-starved common bean. Either one or two foliar sprays of KH₂PO₄ were not sufficient to affect the growth and nutrition of the common bean plants, regardless of soil P status.

Key words: Phaseolus vulgaris, phosphorus, phosphate and phosphite anions, foliar application.

INTRODUCTION

Limited phosphorus (P) availability in Ultisols and Oxisols has been identified as one of the major problems for plant growth in tropical and subtropical regions of the world. High rate P "fixation" and formation of insoluble complexes with aluminum and iron under acid conditions are recognized as an important factor contributing to the low P availability. Thus, application of P-containing fertilizers in these soils is a necessary practice for adequate crop yields in many instances (Vance et al., 2003) and foliarapplied P may increase use efficiency by minimizing soil supply (Girma et al., 2007; Mosali et al., 2006).

Phosphate anions $(H_2PO_4^-, HPO_4^{-2}^- and PO_4^{-3})$ are considered as the main phosphorus forms assimilated by plants and these can induce adequate growth and development with consequences in yield. However, another P form known as phosphite has been widely marketed either as fungicide or as a superior P source for plant nutrition (McDonald et al., 2001; Thao and Yamakawa, 2009; Deliopoulos et al., 2010). Phosphite anions $(H_2PO_3^- and HPO_3^{-2})$ are reduced forms of phosphate anions, in which one hydroxyl group is substituted by hydrogen (Danova-Alt et al., 2008).

Several studies conclusively indicate that phosphite is

^{*}Corresponding author. E-mail: fabriciowilliamavila@yahoo. com.br. Tel: +55 35 38291252.

effective in controlling some important plant diseases caused by pathogens belonging to the class Oomycetes (phylum Oomycota), such as *Phytophthora* sp. Action of phosphite anion is based on two mechanisms: the first is a direct toxic action on the pathogen and the second in indirect action due to phosphite anion activates plant defence responses (McDonald et al., 2001; Wilkinson et al., 2008; Shearer and Fairman, 2007; Orbovic et al., 2008; Cook et al., 2009; Moor et al., 2009). Thus, phosphite has been used as active ingredient in several fungicides.

In terms of plant nutrition, phosphite-based products have been recommended as fertilizers for foliar application, and number of foliar fertilizers containing the phosphite anion has recently increased (Moor et al., 2009). Phosphite salts are recommended as fertilizer because they contain a cation that may be plant nutrient, such as K^+ , NH_4^+ , Ca^{2+} , Mg^{2+} , Cu^{2+} or Zn^{2+} , and the P in form of phosphite anion. However, results of studies that investigated nutritional value of phosphite anion as a P nutrient are inconclusive. In the year 1990, it was reported foliar application of potassium phosphite improved set fruit and yield of avocado, and restored normal growth of phosphate-starved citrus (Lovatt, 1990; Lovatt, 1990). Similarly, positive effects of phosphite on plant P nutrition or crop yields also were demonstrated in other works (Albrigo, 1999; Rickard, 2000; Watanabe, 2005). On the other hand, recent studies have indicated phosphite anion may not be used by plants as a P nutrient, even though it is well absorbed by leaves and roots. Moreover, there are indications phosphite supply causes growth depression in phosphate-starved plants (McDonald et al., 2001; Schroetter et al., 2006; Thao et al., 2008, 2009). In this case, it appears that phosphite inhibits the gene expression related to the responses for overcoming P starvation, such as increased phosphatase activity, synthesis of high affinity transporters for P and elongation of the root system (Varadarajan et al., 2002; Ticconi et al., 2001; Lee et al., 2005).

The aim of this study was to investigate (i) interference produced by different soil phosphate levels, to evaluate (ii) action produced by foliar-applied phosphorus forms (phosphite and phosphate), and to measure (iii) as number of foliar application can act on growth, phosphorus status, and nutritional aspect in common bean (*Phaseolus vulgaris* cv. Radiante) plants.

MATERIALS AND METHODS

Growth conditions, substrate, and plant material

Study was implemented in Departamento de Ciência do Solo of the Universidade Federal de Lavras, Brazil (21°14' S; 45°00' W; 915 m asl). Plants remained in glasshouse environment under natural conditions day/night. Substrate used was composed by low-fertility Oxisol (Typic Haplustox) placed in plastic pots with capacity of 6 L (Table 1). For plant material, common bean (*Phaseolus vulgaris* cv.Radiante) was used.

Substrate preparation

Surface soil with depth from 0 to 20 cm was collected from a noncultivated field with natural Brazilian cerrado vegetation, allowed to dry, crushed to pass through a 4-mm sieve and then mixed with CaCO₃ and MgO (4:1 stoichiometric ratio of Ca:Mg) to raise soil base saturation to 60% of cation exchange capacity at pH 7.0. After 30 days of incubation, a basal nutrient solution was applied and was thoroughly mixed with the soil. Nutrients without P treatments were supplied at the following rates of 90 N, 80 K, 30 S, 5 Zn, 5 Mn, 2 Cu, 1 B and 0.25 Mo mg dm³ of dry soil.

Experimental application

Experiment was organized in factorial scheme completely randomized using 2 soil phosphate levels (Pi-starved and Pi-sufficient), combined with 3 nutrient sources supplied via foliar application (KH₂PO₃, KH₂PO₄, and KCI used as control), and 2 foliar application numbers (single and two applications). For soil phosphate levels, Pi-starved and Pi-sufficient corresponded to 40 and 200 mg of P per dm³ of dry soil, respectively, applied together with the basal nutrient solution. This study had 3 replicates, and each experimental unit consisted of one pot containing two plants, and all variables measured were expressed as mean of two plants.

Nutrient solutions and foliar applications

Solutions of KH_2PO_3 (monobasic potassium phosphite pa), KH_2PO_4 (monobasic potassium phosphate pa) and KCI (potassium chloride pa) were sprayed at concentration of 40 µM, using a manual backpack sprayer. Concentration of P equals the used dose of approximately 3 L of commercial potassium phosphate to 400 L of water, which is usually recommended for growing beans. And KH_2PO_3 was obtained by reaction of H_3PO_3 (phosphorous acid pa) with KOH (potassium hydroxide pa). Single application was implemented when plants presented fourth trifoliate leaf stage, and two applications was carried out in stage of fourth trifoliate leaf and another application in the beginning of flowering stage.

Fertilization as top dressing, irrigation, and harvest

During the soil pot experiment, fertilizations with 240 N, 210 K, and 45 S mg dm⁻³ of dry soil were supplied as top dressing. These fertilizations were split among into three applications throughout the experiment. Soil moisture was maintained at 60% of the total soil pore space occupied by water through daily irrigation. Plants were harvested at full flowering stage and separated into shoot and root. Both shoot and root were rinsed in deionized water and dried at 60°C for 72 h prior to dry weight determination.

Phosphorus determinations

Shoot and root dry mass were ground and analyzed for total P content colorimetrically (Murphy and Riley, 1962) after nitricperchloric digestion of the plant material (Johnson and Ulrich, 1959). Data from shoot and root dry wt and total P concentration were used to calculate the P accumulation, P uptake efficiency (Swiader et al., 1994) (P total accumulation in plant / root dry wt), P utilization efficiency (Siddiqi and Glass, 1981) [(plant dry wt)²/ (P total accumulation in plant)], and P translocation from root to shoot (P total accumulation in plant)

Chemical compositions ⁽¹⁾																
рН	Р	Κ	Zn	Cu	Mn	Fe	EP	Са	Mg	AI	H+AI	Т	m	V	MPAC	
(mg dm ⁻³ of soil)						(mg L ⁻¹)	(mg L^{-1}) (cmol _c dm ⁻³ of soil)					(%)	(mg kg ⁻¹)		
5.4	0.9	22	0.5	0.7	0.4	27.4	20.5	0.1	0.1	0.1	1.7	2	28	13.3	396	
Physical compositions (%) ⁽²⁾																
	Sand Silt					Clay						OM				
60					17		23						0.8			
Mineralogical compositions (g kg ⁻¹ of clay) ⁽³⁾																
SiO ₂	Al ₂ O ₃	Fe	2 0 3	TiO	2	P ₂ O ₅	Fed		Fe₀	C	t	Gb		Ki	Kr	
95.1	97.4	36	6.2	6.2		0.0	10.8		0.1	75	2.0	63.0	0	.98	0.71	

 Table 1. Chemical, physical and mineralogical compositions of Oxisol.

⁽¹⁾ pH in water (1:2.5), P and K by Mehlichl extraction, Mg and Al extractable by 1 M KCl solution (Thomas, 1982); P in the equilibrium solution (EP) according to Alvarez et al. (2000); level of organic matter (OM) according to Anne (1945). T = Cation exchange capacity at pH 7.0; m = Aluminum saturation index; V = Base saturation index and MPAC = maximum P adsorption capacity (Ohtake et al., 1996).⁽²⁾ The soil granulometry was determined by the pipette method of Day (1965). ⁽³⁾ SiO₂, Al₂O₃, Fe₂O₃, TiO₂ and P₂O₅ were determined according to Vettori (1969) with modifications (Embrapa ,1997); Fe_d, according to Mehra and Jackson (1960); Fe_o, according to Schwertmann (1964) and Ct (kaolinite) and Gb (gibbsite) according to Klug and Alexander (1974). Ki = SiO₂ / Al₂O₃ and Kr = SiO₂ / (Al₂O₃ + Fe₂O₃).

Macro and micronutrients

Concentrations of nutrients in shoot were determined after nitric-perchloric digestion as follows: S by turbidimetry; K by flame photometry; Ca, Mg, Cu, Mn, Fe, and Zn by flame atomic absorption spectroscopy. Total N was determined using the Kjeldahl method after sulphuric digestion, and B by colorimetry using the Azomethine-H method after dry digestion, with ash content obtained in muffle furnace by 1 h at 550°C.

Data analysis

Results were submitted to variance analysis (F teste, $p \le 0.05$), and when significant differences occurred were applied to Tukey test at 5% level of error probability ($p \le 0.05$), standard errors were calculated in all evaluated points. The statistical analyses were carried out with the Sisvar software (Ferreira, 2008).

RESULTS AND DISCUSSION

Biomass yield

Most variables in this study were not significantly affected (p > 0.05) by foliar application numbers during single application timing and two application timings (Figure 1). As expected, common bean plants grown under limiting phosphate availability (Pi-starved) showed considerable reductions in the root and shoot dry wt and increased root to shoot ratio. The increase root to shoot ratio by phosphate-starved plants is a mechanism for overcoming P deficiency (Clarkson, 1985).

Foliar application of potassium phosphite (KH_2PO_3) and potassium phosphate (KH_2PO_4) had no significant effect (p > 0.05) on biomass

vield of phosphate-sufficient common bean, when compared with the control (foliar application of potassium chloride). However, for plants grown under limiting phosphate availability, shoot and root dry weight were significantly decreased by foliar-applied potassium phosphite. In addition, root to shoot ratio also was increased with two foliar applications of potassium phosphite (an application in the fourth trifoliate leaf stage and another application in the beginning flowering stage) due to the strong inhibition of shoot dry mass yield. This same behavior also occurred for a single foliar application of potassium phosphite (in the fourth trifoliate leaf stage) but in this case there was no significant difference by Tukey's test (p > 0.05). Hence, our results showed phosphite may not be used by common bean as a P nutrient, and that this anion inhibits biomass yield



Figure 1. Root dry weight, shoot dry weight, and root to shoot ratio in common bean grown in Oxisol under 2 phosphate levels (Pi-starved and Pi-sufficient), 3 nutrient sources supplied via foliar application (KH₂PO₃, KH₂PO₄, and KCl), and 2 foliar application numbers (single and two applications). Averages followed by the same lowercase letter within soil phosphate levels and uppercase letter among foliar application for each phosphate level, do not differ among themselves by the Tukey test at 5% of probability. The bars represent the mean standard error.

under phosphate-deficient conditions.

The inhibiting effect of the phosphite anion on growth of phosphate-starved plants has been reported by different workers (Thao and Yamakawa, 2009). The causes of this effect are not well understood. The most plausible hypothesis to date is that plants do not metabolize phosphite anion, which, after uptake, remains stable in the cell compartments. Furthermore, phosphite anion inhibits some mechanisms involved in overcoming of phosphate deficiency, such as increased synthesis of phosphatases, phosphodiesterases, nucleases, and highaffinity P transporters. Most likely, the molecular mecha-



Figure 2. Shoot P concentration, root P concentration, accumulated P in shoot, and accumulated P in root in common bean grown in Oxisol under 2 phosphate levels (Pi-starved and Pi-sufficient), 3 nutrient sources supplied via foliar application (KH_2PO_3 , KH_2PO_4 , and KCI), and 2 foliar application numbers (single and two applications). Averages followed by the same lowercase letter within soil phosphate levels and uppercase letter among foliar application for each phosphate level, do not differ among themselves by the Tukey test at 5% of probability. The bars represent the mean standard error.

nisms responsible for signaling P deficiency do not discriminate phosphate from phosphite. Thus, there is no expression of genes responsible for proteins involved in P starvation responses (Varadarajan et al., 2002; Ticconi et al., 2001; Lee et al., 2005).

We also found that one and two foliar applications of potassium phosphate had no significant effect (p > 0.05) on biomass yield of phosphate-starved common bean, when compared with the control. Thus, these results suggest that several foliar applications of phosphate may

be necessary to adequately correct a P deficiency, impractical in most cases.

Concentrations and accumulations of P in shoot and root

Common bean plants grown under limiting phosphate availability (Pi-starved) showed decreased concentrations and accumulations of P in shoot and root (Figure 2). Foliar-applied potassium phosphite did not affect P nutrition of phosphate-sufficient plants, but increased concentration of P in shoot of phosphate-starved plants. However, accumulation of P was not significant varied among foliar application treatments (p > 0.05), showing that this increased concentration of P was not due directly to the P from the foliar-applied phosphite, but likely to concentration effect, which is confirmed by the lower shoot dry weight. When biomass yield decreases, this concentration effect for some nutrients may occur (Crusciol et al., 2008; Marschner, 1995), which is the elevation of their concentration in the tissues without there being an alteration in the quantity of nutrient taken up.

In this study, foliar-applied potassium phosphate did not significantly affect (p > 0.05) concentrations and accumulations of P in common bean, when compared with the control. This shows that either one or two foliar applications of phosphate were not sufficient to affect the plant P status. Several attempts to use foliar-applied phosphate in plant nutrition are known, but results are inconclusive. It was recorded that phosphate uptake by leaves after the foliar spray is about 50% (Kannan, 1990). A previous study reported that P concentration in grain of common bean grown under field conditions was not affected by three foliar application timing of phosphate anion (Contee Castro and Boaretto, 2001). On the other hand, Girma et al. (2007) found effect of foliar-applied phosphate on forage and grain P concentrations of maize varied with both applied P levels and plant growth stage. Another study by Mosali et al. (2006) on winter wheat indicated foliar application of phosphate generally increased grain yield, P uptake and P use efficiency, suggesting the authors that low rates of foliar-applied phosphate might correct mid-season P deficiency.

Effects on P uptake efficiency, P utilization efficiency, and P translocation

Either one or two foliar applications of potassium phosphate and phosphite had no significant effects (p >0.05) on P uptake efficiency such as ability to take up P from soil, and also P translocation such as ability to transporter P from root to shoot (Figure 3). Nevertheless, regardless of the foliar-applied treatments, limiting phosphate availability in soil reduced P uptake efficiency and P translocation by common bean. This was principally due to decreased P uptake by phosphate-starved plants according to the Figure 2, although the root dry weight (Figure 1) also was reduced, but in lower magnitude. To reiterate, in this study the P uptake efficiency represent the P-uptake amount per root dry weight unit, and the P translocation from root to shoot represent the ratio between accumulated P in shoot and accumulated P in plant. When the phosphate availability to plants is insufficient, P translocation from root to shoot decreases,

increasing the root growth rate to the detriment of shoot growth rate. Thus, phosphate-starved plants commonly have higher root dry weight to shoot dry weight ratio, a response that enhances the P uptake efficiency (Raghothama, 1999; Schenk, 2006).

Foliar-applied treatments did not affect P utilization efficiency (that is, ability to yield biomass for a given plant P concentration) (Siddigi and Glass, 1981) of phosphatesufficient plants, whereas under limiting phosphate availability in soil, foliar-applied phosphite decreased P utilization efficiency of common bean. This result was due to the inhibitory effect of phosphite on biomass yield of the phosphate-starved plants (according to the Figure 1), since accumulated P of the plants was not affected by the treatments. In contrast, when compared with the control, either one or two application timings of foliar potassium phosphate had no significant effect (p > 0.05) on P utilization efficiency. Other studies had shown phosphatestarved plants exhibit more P utilization efficiency, as response to soil phosphate deficiency, but this more P utilization efficiency varies among cultivars of the same species. Akhtar et al. (2008) found Brassica cultivars differ substantially in P utilization efficiency when grown with sparingly soluble P-forms $(Ca_3(PO_4)_2)$ and Jordan rock-P). These investigators suggested the existence of useful genetic differences among cultivars for mobilization of P from sparingly soluble P-sources. In spite of phosphate-starved plants tend to increase the P utilization efficiency; the results of this study may have been a reflection of the strong P uptake decrease of the plants grown under phosphate deficient Oxisol.

Concentrations of nutrients in shoot

Likewise for shoot P concentration, concentrations of other nutrients in plant shoot tissues were not significantly affected (p > 0.05) by foliar application numbers (Table 2). Apart from the K, common bean grown under limiting phosphate availability exhibited higher concentrations of nutrients in shoot, regardless of the foliarapplied treatments. Foliar application of potassium phosphite also increased concentrations of N, K, Mg, B, Cu, Mn and Fe in shoot tissues of phosphate-starved plants. However, this increased concentration of nutrients coincides with the decreased shoot dry weight (Figure 1), which may suggest that it is involved with the concentration effect, as mentioned above in the presentation of P concentration. Indeed, the applied treatments did not increase accumulation of nutrients in shoot (data not shown), supporting the suggestion above. On the contrary, plants grown under limiting phosphate availability exhibited lower accumulations for all nutrients measured in shoot, regardless of the foliar-applied treatments, due the strong inhibition of shoot biomass vield. Likewise, foliar application of potassium phosphite decreased the accumulation for the majority of nutrients



Figure 3. P uptake efficiency, P utilization efficiency, P translocation in common bean grown in Oxisol under 2 phosphate levels (Pi-starved and Pi-sufficient), 3 nutrient sources (KH_2PO_3 , KH_2PO_4 , and KCI), and 2 foliar supplied via foliar application application numbers (single and two applications). Averages followed by the same lowercase letter within soil phosphate levels and uppercase letter among foliar application for each phosphate level, do not differ among themselves by the Tukey test at 5% of probability. The bars represent the mean standard error.

in shoot of phosphate-starved plant, whereas for others nutrients the foliar treatment effects were not significant (p > 0.05). In general, foliar-applied phosphate has no effect on nutrient concentrations and accumulations in shoot. Hu et al. (2008) also did not find alterations in leaf P concentrations of maize plants submitted to foliar NPK applications. In this study the concentrations of nutrients in roots (data not shown) were not affected by application of foliar potassium phosphate and phosphite as well as by foliar application numbers, whereas the limiting phosphate availability in soil increased concentration and decreased accumulation of some nutrients.

Application		Foliar treatments		Macron	utrients ((g kg ⁻¹)		Micronutrients (mg kg ⁻¹)					
numbers	Soll P status		N	К	Са	Mg	S	В	Zn	Cu	Mn	Fe	
		KCI	47 ^{bA}	19 ^{bA}	13 ^{aA}	6 ^{bA}	2 ^{aA}	32 ^{abA}	57 ^{aA}	6 ^{bA}	61 ^{bA}	347 ^{bA}	
	Pi-starved	Phi	57 ^{aA}	22 ^{aA}	15 ^{aA}	9 ^{aA}	2 ^{aA}	37 ^{aA}	61 ^{aA}	8 ^{aA}	77 ^{aA}	511 ^{aA}	
A single		Pi	46 ^{bA}	18 ^{bA}	14 ^{aA}	6 ^{bA}	2 ^{aA}	30 ^{bA}	58 ^{aA}	5 ^{bA}	58 ^{bA}	381 ^{bA}	
foliar applicat	ion												
timing		KCI	33 ^{aB}	17 ^{aA}	9 ^{aB}	4 ^{aB}	1 ^{aB}	17 ^{aB}	30 ^{aB}	4 ^{aB}	39 ^{aB}	131 ^{aB}	
	Pi-sufficient	Phi	33 ^{aB}	17 ^{aB}	9 ^{aB}	4 ^{aB}	1 ^{aB}	17 ^{aB}	29 ^{aB}	3 ^{aB}	42 ^{aB}	183 ^{aB}	
		Pi	31 ^{aB}	18 ^{aA}	8 ^{aB}	3 ^{aB}	1 ^{aB}	16 ^{aB}	35 ^{aB}	4 ^{aB}	37 ^{aB}	167 ^{aB}	
		KCI	46 ^{bA}	20 ^{bA}	14 ^{aA}	6 ^{bA}	2 ^{aA}	31 ^{abA}	52 ^{aA}	5 ^{bA}	55 ^{bA}	334 ^{bA}	
	Pi-starved	Phi	55 ^{aA}	23 ^{aA}	16 ^{aA}	8 ^{aA}	2 ^{aA}	36 ^{aA}	53 ^{aA}	7 ^{aA}	73 ^{aA}	559 ^{aA}	
Two		Pi	45 ^{bA}	19 ^{bA}	13 ^{aA}	6 ^{bA}	2 ^{aA}	29 ^{bA}	51 ^{aA}	5 ^{bA}	62 ^{abA}	310 ^{bA}	
foliar applicat	ion												
timings		KCI	29 ^{aB}	18 ^{aA}	9 ^{aB}	3 ^{aB}	1 ^{aB}	17 ^{aB}	32 ^{aB}	4 ^{aB}	40 ^{aB}	186 ^{aB}	
C	Pi-sufficient	Phi	34 ^{aB}	19 ^{aB}	8 ^{aB}	4 ^{aB}	1 ^{aB}	16 ^{aB}	34 ^{aB}	3 ^{aB}	38 ^{aB}	176 ^{aB}	
		Pi	32 ^{aB}	19 ^{aA}	7 ^{aB}	4 ^{aB}	1 ^{aB}	16 ^{aB}	36 ^{aB}	3 ^{aB}	41 ^{aB}	148 ^{aB}	
Source of varia	ation:												
Foliar application numbers (A)			ns	ns	ns	ns	ns	ns	ns	ns	ns	Ns	
Soil P status (B)			***	***	***	***	***	***	***	***	***	***	
Foliar treatments (C)			***	*	ns	***	ns	**	ns	***	*	***	
A×B			ns	ns	ns	ns	ns	ns	ns	ns	ns	Ns	
A×C			ns	ns	ns	ns	ns	ns	ns	ns	ns	Ns	
B×C			*	*	ns	**	ns	*	ns	***	*	***	
A×B×C			ns	ns	ns	ns	ns	ns	ns	ns	ns	Ns	

Table 2. Concentrations of nutrients in common bean shoot grown in Oxisol under 2 phosphate levels (Pi-starved and Pi-sufficient), 3 nutrient sources supplied via foliar application (KH₂PO₃, KH₂PO₄, and KCl), and 2 foliar application numbers (single and two applications).

In each number of foliar application (a single application timing and two application timings), lower case compare the foliar application products (KCI, Phi and Pi) for each soil phosphate level (Pi-starved and Pi-sufficient), and upper case compare the soil phosphate levels for each foliar application product. Means followed by same letter are not different by Tukey's test ($p \le 0.05$). *, **, ***, and ns corresponding to $p \le 0.05$, $p \le 0.01$, $p \le 0.001$, and non-significant, respectively, by F test.

Conclusion

Common bean growth under limiting phosphate availability in soil exhibited lower biomass yield and higher concentration of nutrients in shoot tissues, which may be due to concentration effect since accumulation of nutrients in shoot was not increased. Either one or two foliar sprays of potassium phosphate were not sufficient to affect the growth and nutrition of the common bean, regard-less of soil P status. However, the results exhibit foliar-applied potassium phosphite causes harmful effects in phosphate-starved common bean, but no effect is observed in phosphatesufficient common bean, confirming earlier investigations with other plant species. Our results indicate phosphite anion may not be recommended as a P source for nutrition of common bean, but it is suitable to be used for other purposes that requires an optimum soil phosphate status.

ACKNOWLEDGEMENTS

The authors are grateful to the CNPq, CAPES and FAPEMIG, all from Brazil, for the financial support. F.W. Ávila also thanks CNPq for a postgraduate fellowship.

REFERENCES

- Akhtar MS, Oki Y, Adachi T (2008). Genetic variability in phosphorus acquisition and utilization efficiency from sparingly soluble p-sources by *Brassica* cultivars under P-stress environment. J. Agron. Crop Sci., 194(5): 380-392.
- Albrigo LG (1999). Effects of foliar applications of urea or Nutriphite on flowering and yields of Valencia orange trees. Proc. Fla. State Hort. Soc., 112(1): 1-4.
- Alvarez VVH, Novais RF, Dias LE, Oliveira JA (2000). Determination and phosphorus use available. Boletim Informativo da Sociedade Brasileira de Ciência do Solo, 25(1): 27-32.
- Anne A (1945). Sur le dosage rapide du carbone organique des sols. Ann. Agron., 2(1): 161-172.
- Clarkson DT (1985). Factors affecting mineral nutrient acquisition by plants. Ann. Rev. Plant Physiol., 36(1): 77-115.
- Barbara: University of California, California. Agric. Exp. Stat. Bull., 766: 1956.
- Contee CAM, Boaretto AE (2001). Foliar fertilization of bean with nutrients, b1 vitamin and methionine. Sci. Agraria, 2(1): 117-121.
- Cook PJ, Landschoot PJ, Schlossberg MJ (2009). Inhibition of *Pythium* spp. and suppression of Pythium Blight of turfgrasses with phosphonate fungicides. Plant Dis., 93(8): 809-814.
- Crusciol CAC, Arf Ö, Soratto RP, Mateus GP (2008). Grain quality of upland rice cultivars in response to cropping systems in the Brazilian Tropical Savanna. Sci. Agrícola, 65(5): 468-473.
- Danova-Alt R, Dijkema C, Dewaard P, Köck M (2008). Transport and compartmentation of phosphite in higher plant cells-kinetic and ³¹P nuclear magnetic resonance studies. Plant Cell Environ., 31(10): 1510-1521.
- Day PR (1965). Particle fractionation and particle-size analysis. In: Black CA (ed.). Methods of soil analysis, Part 1, Madison, Wisconsin, USA: Am. Soc. Agron., pp. 545-566.
- Deliopoulos T, Kettlewell PS, Hare MC (2010). Fungal disease suppression by inorganic salts: A review. Crop Prot., 29(10): 1059-1075.
- Embrapa-Empresa Brasileira de Pesquisa Agropecuária (1997). Methods of soil analysis. 2 Ed. Centro Nacional de Pesquisa de Solos/EMBRAPA, Rio de Janeiro, RJ, Brazil.
- Ferreira DF (2008). Sisvar: a program for statistical analysis and teaching. Revista Sympos., 6(1): 36-41.
- Girma K, Martin KL, Freeman KW, Mosali J, Teal RK, Raun WR, Moges SM, Arnall DB (2007). Determination of optimum rate and growth stage for foliar-applied phosphorus in corn. Commun. Soil Sci. Plant Anal., 38(9): 1137-1154.
- Hu Y, Burucs Z, Schmidhalter U (2008). Effect of foliar fertilization application on the growth and mineral nutrient content of maize seedlings under drought and salinity. Soil Sci. Plant Nutr., 54(1): 133-141.
- Johnson CM, Ulrich A (1959). Analytical methods for use in plant analysis. California Agric. Exp. Station Bull., p. 766.
- Kannan S (1990). Role of foliar fertilization on plant nutrition. In: Baligar VC, Duncan RR (eds.). Crops as enhancers of nutrient use. Academic Press, San Diego, USA. pp. 313-348.

- Klug HP, Alexander LE (1974). X-ray diffraction procedures for polycrystalline and amorphous materials, John Wiley and Sons, New York, NY, USA.
- Lee TM, Tsai PF, Shyu YT, Sheu F (2005). The effects of phosphite on phosphate starvation responses of *Ulva lactuca* (Ulvales, chlorophyta). J. Phycol., 41(5): 975-982.
- Lovatt CJ (1990). Foliar phosphorus fertilization of citrus by foliar application of phosphite. In: Citrus Research Advisory Committee (ed.). Summary of Citrus Research. University of California, Riverside, USA, pp. 25-26.
- Lovatt CJ (1990). A definitive test to determine whether phosphite fertilization can replace phosphate fertilization to supply P in the metabolism of 'Hass' on 'Duke 7'. California Avocado Society Yearbook, 74(1): 61-64.
- Marschner H (1995). Mineral nutrition of higher plants. Academic Press, London, England.
- McDonald AE, Grant BR, Plaxton WC (2001). Phosphite (Phosphorous acid): Its relevance in the environment and agriculture and influence on plant phosphate starvation response. J. Plant Nutr., 24(10): 1505-1519.
- Mehra OP, Jackson ML (1960). Iron oxide removal from soils and clays by a dithionite-citrate system buffered with sodium bicarbonate. Clays Clay Miner., 7(1): 317-327.
- Moor U, Põldma P, Tõnutare T, Karp K, Starast M, Vool E (2009). Effect of phosphite fertilization on growth, yield and fruit composition of strawberries. Sci. Hortic., 119(2): 264-269.
- Mosali J, Desta K, Teal RK, Freeman KW, Martin KL, Lawles JW, Raun WR (2006). Effect of foliar application of phosphorus on winter wheat grain yield, phosphorus uptake, and use efficiency. J. Plant Nutr., 29(12): 2147-2163.
- Murphy J, Riley HP (1962). A modified single solution method for the determination of phosphate in natural waters. Anal. Chim. Acta, 27(1): 31-36.
- Ohtake H, Wu H, Imazu K, Anbe Y, Kato J, Kuroda A (1996). Bacterial phosphonate degradation, phosphite oxidation and polyphosphate accumulation. Resour. Conserv. Recycl., 18(1): 125-134.
- Raghothama KG (1999) Phosphate acquisition. Annu. Rev. Plant Physiol., Plant Mol. Biol., 50(1): 665-693.
- Rickard DA (2000). Review of phosphorus acid and its salts as fertilizer materials. J. Plant Nutr., 23(2): 161-180.
- Schenk MK (2006). Nutrient efficiency of vegetable crops. Acta Hortic., 700(1): 21-34.
- Schroetter S, Angeles-Wedler D, Kreuzig R, Schnug E (2006). Effects of phosphite on phosphorus suplly and growth of corn (*Zea mays*). Landbauforsch Völkenrode, 56(1): 87-99.
- Schwertmann U (1964). Differenzierung der eisenoxide des bondes durch extraktion mit ammonium-oxalat-losung. Zeitschrift Pflanzenernähr Düng Bodenkd, 105(1): 194-202.
- Shearer BL, Fairman RG (2007) A stem injection of phosphite protects *Banksia* species and *Eucalyptus marginata* from *Phytophthora cinnamomi* for at least four years. Australas Plant Pathol., 36(1): 78-86.
- Siddiqi MY, Glass ADM (1981). Utilization Index: A modified approach to the estimations and comparison of nutrient utilization efficiency in plants. J. Plant Nutr., 4(3): 289-302.
- Swiader JM, Chyan Y, Freiji FG (1994). Genotypic differences in nitrate uptake and utilization efficiency in pumpkin hybrids. J. Plant Nutr., 17(10): 1687-1699.
- Thao HTB, Yamakawa T, Shibata K, Sarr PS, Myint AK (2008). Growth response of komatsuma (*Brassica rapa* var. peruvirids) to root and foliar applications of phosphate. Plant Soil, 308(1): 1-10.
- Thao HTB, Yamakawa T, Shibata K (2009). Effect of phosphitephosphate interaction on growth and quality of hydroponic lettuce (*Lactuca sativa* L.). J. Plant Nutr. Soil Sci., 172(3): 378-384.
- Thao HTB, Yamakawa T (2009). Phosphite (phosphorous acid): Fungicide, fertilizer or bio-stimulator? Soil Sci. Plant Nutr., 55(2): 228-234.
- Ticconi CA, Delatorre CA, Abel S (2001). Attenuation of phosphate starvation responses by phosphite in Arabidopsis. Plant Physiol., 127(3): 963-972.
- Thomas GW (1982). Exchangeable cations. In: Page AL, Miller RH and Keeney DR (eds.). Methods of soil analysis, Part 2, Madison,

- Wisconsin, USA: American Society of Agronomy, Soil Sci. Soc. Am., pp. 159-165.
- Vance CP, Uhde-Stone C, Allan DL (2003). Phosphorus acquisition and use: critical adaptations by plants for securing a nonrenewable resource. New Phytol., 157(3): 423-447.
- Varadarajan DK, Karthikeyan AS, Matilda PD, Raghothama KG (2002). Phosphite, an analog of phosphate suppresses the coordinated expression of genes under phosphate starvation. Plant Physiol., 129(3): 1232-1240.
- Vettori L (1969). Methods of soil analysis. Ministério da Agricultura, Rio de Janeiro, RJ, Brazil, p. 24.
- Watanabe K (2005). A new fertilizer for foliar application, phosphite fertilizer. Fertilizer, 101(1): 91-96.
- Wilkinson CJ, Holmes JM, Dell B, Tynan KM, Mccomb JA, Shearer BL, Orbović V, Syvertsen JS, Bright D, Van Clief DL, Graham JH (2008). Citrus seedling growth and susceptibility to root rot as affected by phosphite and phosphate. J. Plant Nutr., 31(4): 774-787.