

Research Paper

Development and assessment of a latex agglutination test based on recombinant MSP5 to detect antibodies against *Anaplasma marginale* in cattle

Carlos A.N. Ramos¹, Flávio R. Araújo¹, Rafaelle C. Santos², Elaine S.P. Melo³,
Letícia C. Sousa³, Carlos E.S. Vidal⁴, Neurisvan R. Guerra⁵, Rafael A.N. Ramos⁶

¹Embrapa Gado de Corte, Campo Grande, MS, Brazil.

²Programa de Pós Graduação em Saúde Animal na Amazônia, Universidade Federal do Pará,
Castanhal, PA, Brazil.

³Programa de Pós Graduação em Ciência Animal, Universidade Federal de Mato Grosso do Sul,
Campo Grande, MS, Brazil.

⁴Programa de Pós Graduação em Medicina Veterinária, Universidade Federal de Santa Maria,
Santa Maria, RS, Brazil.

⁵Programa de Pós Graduação em Ciência Animal Tropical, Universidade Federal Rural de Pernambuco,
Recife, PE, Brazil.

⁶Department of Veterinary Medicine, Università degli Studi di Bari, Valenzano, Bari, Italy.

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Abstract

The recombinant protein MSP5 has been established as an important antigen for serological diagnosis of *Anaplasma marginale* by enzyme-linked immunosorbent assay (ELISA). However, due to the high cost of specialized equipment, this technique is not accessible to all laboratories, especially in developing countries in areas where the disease is endemic. The present study describes the standardization of a latex agglutination test (LAT) to detect antibodies against *A. marginale* based on recombinant MSP5. Compared with indirect enzyme-linked immunosorbent assay (iELISA), the relative sensitivity and specificity of the LAT were 95.21% and 91.86% respectively, with an almost perfect agreement between tests (kappa index = 0.863). These results can be considered important for the serological diagnosis of *A. marginale*, as they indicate that the test represents a rapid and low cost alternative to ELISA.

Key words: latex agglutination, rapid test, *Anaplasma marginale*, MSP5.

Introduction

A variety of serological tests, such as the indirect fluorescent antibody test (IFAT) (Lohr *et al.*, 1973), complement fixation (Gainer, 1961), enzyme-linked immunosorbent assay (ELISA) (Luckins, 1977) and more recently, the immunosensor (Silva *et al.*, 2006a) have been used to detect antibodies to *A. marginale*. However, most of these techniques require equipment and specialized technicians, which increase the reaction costs substantially. Thus, the development of a simpler and less expensive method is desirable for laboratories with little infrastructure, especially

in developing countries with areas where the disease is endemic.

In recent years, the major surface protein (MSP) 5 has been established as an important antigen for serological diagnosis of *A. marginale* (Reyna-Bello *et al.*, 1998; Silva *et al.*, 2006a,b; Melo *et al.*, 2007). The use of recombinant antigens in serological assays has favored the development of highly standardized diagnostic tests that are both reproducible and inexpensive (Boonchit *et al.*, 2002).

The aim of the present study was to develop and evaluate a latex agglutination test (LAT) using recombinant MSP5 as antigen to detect antibodies against *A. marginale* in cattle.

Materials and Methods

Serum samples source

All cattle serum samples used in the present study were originally collected for other epidemiological studies between 2005 and 2010, and stored at -20 °C at Embrapa Beef Cattle, Campo Grande, MS, Brazil. The sample selection for assessment by serological methods was based on the convenience for researchers and the epidemiological status of the region of origin to *A. marginale*.

Serum samples were obtained from cattle that had been experimentally infected with *A. marginale* (n = 48) and from cattle kept in a tick-free isolation area of Embrapa Beef Cattle, Campo Grande, MS, Brazil (n = 50). Sera was also obtained from cattle raised in areas of endemic stability of *A. marginale*: from the Brazilian states of Pará (n = 71), Rio de Janeiro (n = 66) and Mato Grosso do Sul (n = 76) (Souza *et al.*, 2000; Guedes Junior *et al.*, 2008); as well as from cattle raised in areas of endemic instability such as the Brazilian states of Rio Grande do Sul (n = 153) and Pernambuco (n = 5); and Costa Rica (n = 54) in central America (Artiles *et al.*, 1995; Oliveira *et al.*, 2011). Serum samples from cattle experimentally infected with *B. bovis* (n = 16) or *B. bigemina* (n = 30) and negative for antibodies against *A. marginale* in iELISA and the immunofluorescent antibody test (IFI) were also used.

Recombinant MSP5 production

Recombinant protein was produced as described by Silva *et al.* (2006b). Briefly, *Escherichia coli* TOP10 with *pTrec-HisTOPO/msp5* was grown in Luria Bertani (LB) broth supplemented with 100 µg/mL ampicillin. Gene expression was stimulated by 1 mM isopropyl-β-D-galactopyranoside (IPTG) and incubation for six hours at 37 °C at 250 rpm. *E. coli* cells were then recovered by centrifugation at 10,000 x g for 10 min, and the recombinant protein was purified in denaturing conditions using the His-Trap HP agarose-nickel resin (GE Healthcare), following the manufacturer's instructions. The purity of rMSP5 was confirmed by sodium dodecyl-polyacrylamide gel electrophoresis (SDS-PAGE) and dialyzed with phosphate saline buffer pH 7.2 (PBS) at 4 °C for 48 hours. Once the proteins became insoluble after dialysis, solubilization with 2% SDS was conducted as described by Lechtzie *et al.* (2002). The protein concentration was determined by SDS-PAGE stained with Coomassie blue by comparison with known concentrations of bovine serum albumin (BSA) using the LabImage v.3.3.2 image analysis software (Loccus, Brazil).

Latex beads/rMSP5 conjugation

Polystyrene latex particle suspension (0.8 µm diameter, Sigma LB-8) was diluted to 1% with phosphate saline buffer (PBS) pH 7.2. The latex particle suspension was centrifuged at 3500 x g for 45 min. After removing the super-

natant, the polystyrene latex particles were resuspended to 1% with previously solubilized rMSP5, diluted in PBS to a final concentration of 0.5 mg/mL. The mixture was incubated for 24 h at room temperature with constant shaking. After incubation, rMSP5-sensitized latex particles were recovered by centrifugation at 3500 x g for 45 min, resuspended in PBS with 0.5 mg/mL of BSA and blocked for one hour at room temperature with constant shaking. The blocked latex particles were washed twice with PBS and centrifuged again at 3500 x g for 45 min. The recovered rMSP5 sensitized latex particles were diluted to 1% with PBS containing 0.01% sodium azide, 0.05 mg/mL of BSA and 5% of glycerol. The rMSP5 latex agglutination test (rMSP5 LAT) antigen was kept at 4 °C until use.

rMSP5 latex agglutination test assessment

The agglutination reactions were performed on black agglutination cards, with antigen and serum at room temperature. The optimal serum/antigen ratio and reaction time were defined by an assessment of three serum samples obtained from cattle kept in a tick-free isolation area of Embrapa Beef Cattle, Campo Grande, MS, and three sera samples from cattle experimentally infected with *A. marginale*. The serum samples used were negative and positive to antibodies against *A. marginale*, respectively, with indirect ELISA (iELISA) with rMSP5 as antigen (Silva *et al.*, 2006b). The selection of optimal parameters was based on naked eye observation of agglutination with positive serum and the absence of agglutination with negative serum. The serum samples were classified only as positive (+) or negative (-).

The relative sensitivity and specificity of the rMSP5 LAT were assessed using the iELISA based on rMSP5 (Silva *et al.*, 2006b) as the standard. Five hundred and sixty-nine bovine serum samples were paired-tested with iELISA and the LAT. An additional assessment of the specificity of the rMSP5 LAT was performed with serum samples from cattle experimentally infected with *B. bovis* (n = 16) or *B. bigemina* (n = 30).

All rMSP5 LAT were performed by a technician without previous information about the status of the sera (positive/negative) as defined by iELISA. This caution was taken to avoid any bias related test interpretation.

Stability and repeatability

The stability of the rMSP5 LAT antigen was assessed by the agglutination of the standard positive serum with sensitized latex beads stored at 4 °C for 8 months. The standard sera used were the same as those used in the standardization reaction (serum/antigen ratio and reaction time).

Repeatability was determined by the analysis of 139 serum samples: 66 positive and 73 negative to antibodies against *A. marginale* with iELISA, using three batches of the rMSP5 LAT antigen (B1, B2 and B3), produced from different inductions and purifications of the recombinant

protein. The serum samples used to assess reproducibility were also selected by convenience. The 139 samples used in this stage of the study were composed of 54 samples from Costa Rica (Central America), 51 samples from the semi-arid of Pernambuco and 34 samples, randomly selected, from Rio Grande do Sul.

Statistical analyses

The agreement between tests (LAT and iELISA) and batches of antigen (B1, B2 and B3) was assessed by the kappa index and interpreted based on the scale described by Landis and Koch (1977). The significance of differences between the tests were obtained by analysis of discordant results by McNemar's χ^2 test. The analyses were realized using the software BioStat 5.0. Differences were considered statistically significant when $p < 0.05$.

Results and Discussion

The rMSP5 sensitized latex and serum agglutinated in the presence of *A. marginale* antibodies but remained evenly dispersed in the absence of antibodies (Figure 1).

The optimal ratio of serum/antigen and reaction time established was 1:4 (10 μ L of serum and 30 μ L of rMSP5 LAT antigen) and three minutes, respectively. The relative comparison between the rMSP5 latex agglutination test and iELISA also based on rMSP5 (Table 1), previously standardized and validated by Silva *et al.* (2006b), resulted in an almost perfect agreement (94.20%, 536/569) with a kappa index of 0.863 (95% CI 0.818-0.908). The relative sensitivity and specificity of the rMSP5 LAT were 95.21%, and 91.86% respectively. No significant difference was observed between the tests ($p = 0.486$). In the assessment of the analytical specificity of the rMSP5 LAT antigen, agglutination was not observed when serum from cattle that had been experimentally infected by *B. bovis* or *B. bigemina* was used.

The rMSP5 LAT antigen was stable during the eight months of assessment. All standard samples showed the same result month to month. Although it was not the objective of the present study to assess the agglutination intensity, a slight reduction was observed over the months. This could compromise the sensitivity of the rMSP5 LAT when performed with antigen stored for a long time. Therefore, a

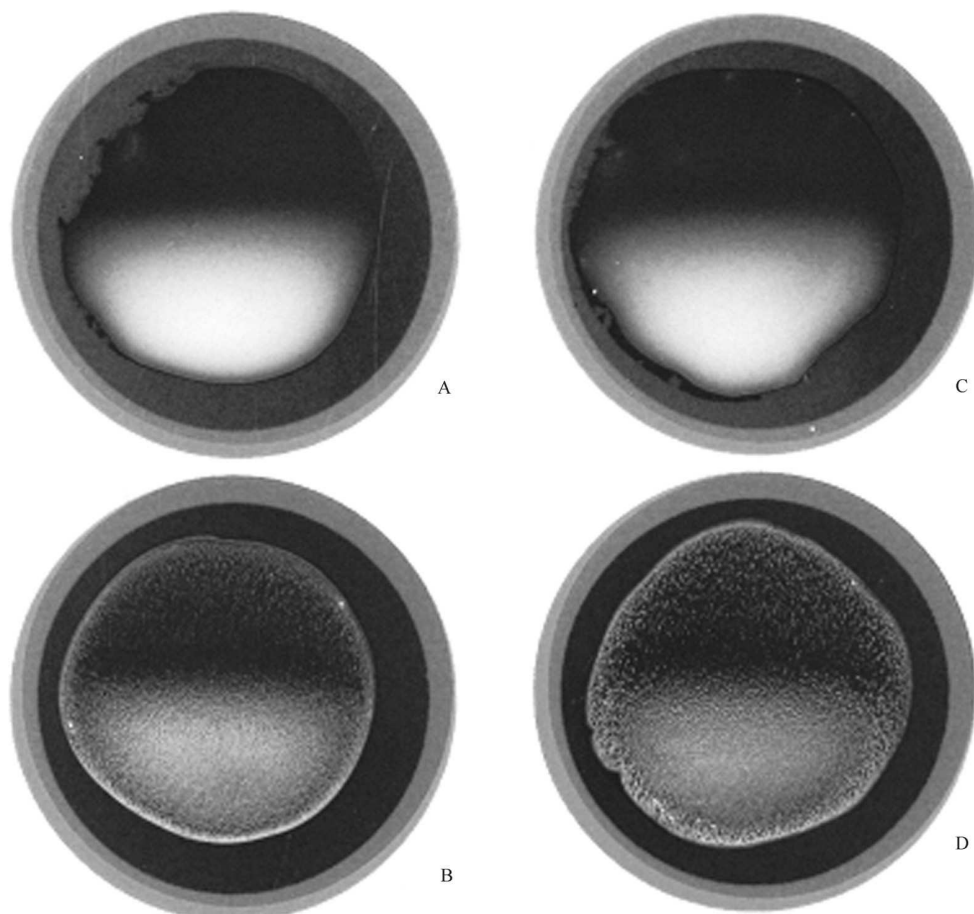


Figure 1 - Latex agglutination test of bovine sera using rMSP5. *Anaplasma marginale* negative (A, C) and positive sera (B, D) were mixed with latex beads coated with rMSP5. Agglutination is clearly observed in panels B and D.

Table 1 - Comparison between the results of iELISA and the rMSP5 LAT to detect antibodies against *A. marginale* in cattle.

rMSP5 LAT	iELISA		Total
	Positive	Negative	
Positive	378	14	392
Negative	19	158	177
Total	397	172	569

more detailed study in relation to LAT antigen stability is necessary. Additional stability assessments of the antigen in other temperature zones were not performed.

The rMSP5 LAT to *A. marginale* showed good repeatability between three different batches of antigen. There was an agreement between: B1 and B2 of 97.12% (kappa 0.941, 95% CI 0.885 - 0.998); B1 and B3 of 97.84% (kappa 0.956, 95% CI 0.907 - 1); B2 and B3 of 99.28% (kappa 0.985, 95% CI 0.956 - 1). The differences found between the batches of antigen were not statistically significant by McNemar χ^2 test (B1 x B2 $P = 0.125$; B1 x B3 $p = 0.250$; B2 x B3 $p = 1$).

Comparisons between seroprevalence obtained by the rMSP5 LAT and iELISA in each region (Pará, Rio de Janeiro, Mato Grosso do Sul, Rio Grande do Sul, semi-arid Pernambuco and Costa Rica) are shown in Table 2. No significant differences were observed between tests. According to the concept of enzootic stability/instability proposed by Mahoney and Ross (1972), no differences were observed in region classification based on the results of the rMSP5 LAT or iELISA.

Agglutination tests to detect antibody against *A. marginale* have been used for decades in capillary tubes, cards or plates (Welter, 1964; Amerault and Roby, 1968; Howarth and Hokama, 1976; Rodgers *et al.*, 1998). However, in all cases the *A. marginale* antigen used is derived from initial body semi-purified from bovine erythrocytes (Wilson *et al.*, 19778) or *in vitro* propagated in tick cell line (Rodgers *et al.*, 1998). These antigens are formed by a complex protein mixture, making it difficult to standardize dif-

ferent batches of antigen, resulting on low reproducibility. In the case of bovine derived antigens, contamination with erythrocytic stroma can contribute to false positive reactions (Barry *et al.*, 1986). In addition, for the production of antigen an experimental infection of cattle is necessary, which increases the production costs and has important ethical implications.

Advances in molecular technologies during the last decade, especially in cloning and gene expression, have enabled the production, with highly reproducible methodologies, of recombinant proteins of a variety of organisms, including *A. marginale* (Silva *et al.*, 2006b; Araújo *et al.*, 2008). Many of these recombinant proteins have been utilized in serodiagnosis of *A. marginale* with excellent results (Reyna-Bello *et al.*, 1998; Melo *et al.*, 2007).

The latex agglutination test appears attractive, since this test is simple and rapid. In the reaction presented in the present study, no more than three minutes were enough to achieve the result with sensitivity and specificity comparable to the other agglutination test of *A. marginale* (Wilson *et al.*, 1978; Rodgers *et al.*, 1998). An additional advantage is that the test does not require previous serum dilution or reading equipment, such as the semi-automated agglutination developed by Rodgers *et al.* (1998), nor does it require a long incubation time before reading as is the case of the capillary agglutination test used by Wilson *et al.* (1978). Furthermore, the LAT does not require bovine serum factor (BSF), unlike the card agglutination assays presented by Amerault and Roby (1968), Howarth and Hokama (1976) and Bastos *et al.* (2000). Another advantage of the rMSP5 LAT is related to the cost and portability. No special equipment is required to perform the test and thus it can be conducted in the field conditions. In Brazil, the cost of the rMSP5 LAT for each sample tested was estimated to be less than US\$ 0.27.

The false positive result of the rMSP5 LAT (15/597) in relation to iELISA can be attributed to immunoglobulin M (IgM) reaction, either from cattle recently infected by *A. marginale* or nonspecific binding (Holliman *et al.*, 1989). Since IgM antibodies appear earlier after infection, it in-

Table 2 - Comparison of the seroprevalence to *Anaplasma marginale* obtained with the rMSP5 LAT and iELISA by region.

Seroprevalence % (Positive/Total)				
Region	rMSP5 LAT	iELISA	p-value ^a	Epidemiological status ^b (rMSP5 LAT/ iELISA)
Pará/Brazil	91.54 (65/71)	90.14 (64/71)	1.00	Stability/ Stability
Rio de Janeiro/Brazil	98.48 (65/66)	100 (66/66)	1.00	Stability/ Stability
Mato Grosso do Sul/Brazil	97.36 (74/76)	97.36 (74/76)	1.00	Stability/ Stability
Rio Grande do Sul/Brazil	60.78 (93/153)	63.39 (97/153)	0.45	Instability/ Instability
Pernambuco/Brazil (semi-arid)	52.94 (27/51)	45.09 (23/51)	0.12	Instability/ Instability
Costa Rica (Central America)	46.29 (25/54)	42.59 (23/54)	0.62	Instability/ Instability

^aMcNemar χ^2 test.^bClassification proposed by Mahoney and Ross (1972).

duces intense agglutination reactions (Janeway Jr. *et al.*, 2005) and is not detected by iELISA used as standard method. On the other hand, most of the false negative reactions may have been due to the difficulty of identifying weak reactions with the naked eye, or due to the prozone effect (Butch, 2000). Some of these problems could easily be minimized by reading automation and sera dilution. However, the test run would be limited to the laboratory.

To assess the hypothesis of the prozone effect, the nineteen iELISA positive and the rMSP5 LAT negative sera samples were diluted serially in PBS (1:4 to 1:16). Six samples showed agglutination that was visible to the naked eye in the 1:4 dilutions, as well as one sample in the 1:8 dilution. No sample agglutinated at 1:16 dilution.

Currently, it is known that MSP5 as the antigen in serodiagnosis is not specific for *A. marginale*. Cross-reactivity with antibodies against *A. centrale*, *A. ovis*, *A. phagocytophilum* and recently a new genotype of *Ehrlichia* (BOV2010), have been reported (Dreher *et al.*, 2005; Al-Adhami *et al.*, 2011). However, in Brazil other species of the blood-borne bacterial family *Anaplasmataceae* infecting cattle does not have been described, except occasional reports of *A. bovis*, limited to the state of Rio de Janeiro (Santos and Carvalho, 2006; Melo Junior *et al.*, 2010). Therefore, although it is not possible to affirm that the specificity of the rMSP5 LAT is restricted to *A. marginale*, in areas in which bovine anaplasmosis is endemic, and other *Anaplasmataceae* do not occur, is a useful epidemiological tool.

In conclusion, the rMSP5 LAT is a simple and economical technique that can be used in the field and is suitable for epidemiological studies, principally in developing countries where the equipment and technology required for ELISA is not always available. Positive results derived from these assay should be interpreted with caution due to the high degree of conservation of the MSP5 among family *Anaplasmataceae*.

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