

*Full Length Research Paper*

## **Fertilizer source influence on antioxidant activity of lettuce**

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**Consumers are increasingly aware and concerned about their health and therefore demanding for more healthy and nutritious food. Thus, the demand for organic foods has increased recently. Thus the aim of this study was to compare the antioxidant activity and total phenolic content of lettuce produced in organic and conventional systems. The antioxidant activity of ethanol extracts was determined by the DPPH and FRAP assay. The total phenolics content was obtained spectrophotometrically according to the Folin-Ciocalteu method and calculated as gallic acid equivalent. The organic lettuce showed higher effectiveness in antioxidant capacity and higher levels of phenolic compounds than lettuce produced in the conventional system.**

**Key words:** *Lactuca sativa*, antioxidant potential, agricultural management, dpph frap, phenolics total.

### **INTRODUCTION**

The current concern with men's health has led to a demand for healthier foods, thus providing a better life quality. Some natural products present high levels of antioxidants that are responsible for the prevention of various diseases. Antioxidants are molecules that inhibit or decrease the damages provoked by free radicals (Shahidi and Ambigaipalan, 2015). Free radicals can be produced by exogenous sources or by the natural

metabolic reactions.

The human metabolism synthesizes free radicals during its physiological reactions that interact with DNA, RNA, proteins, lipids and polysaccharides causing serious damage as degenerative diseases. Vitamins C and E, phenolic compounds, flavonoids and carotenoids are extremely relevant antioxidants that can be acquired through a balanced diet of vegetable origin (Nizmse and

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Pal, 2015)

The consumption of fruits, cereals and vegetables are effective free radicals that fight foods. According to Borges et al. (2016), some studies indicate that the use of organic fertilizers can influence the synthesis of antioxidant compounds. The organic products are better than the conventional, however the studies comparing foods from both managements still scarce.

Lettuce (*Lactuca sativa* L.), belonging to the Asteraceae family, is native from Mediterranean (Medeiros et al., 2016), but is currently cultivated in innumerable regions and is the most popular and appreciated hardwood vegetable.

The phenolic profile of lettuce is well defined and reported in the literature with caffeic acid derivatives and flavonoids, representing the main component class. However, genetic and environmental factors can influence in the phytochemical profile of plants. Therefore, the variety planted, weather, soil type, the light intensity, irrigation, the supply of nutrients, plage control, the system of cultivation, among others can influence on the chemical content of plants (Sofa et al., 2016; Santos et al., 2014; Perez-Lopez et al., 2018; Durazzo et al., 2014).

Several studies analyzing the antioxidant properties of fruits, vegetables, and medicinal plants have been reported (Talens et al., 2016; Calado et al., 2015; Turati, 2015). However, a few papers show the influence of the cultivation type on antioxidant action of these foods. There is a tendency that organically produced foods have lower nitrate content, highest content of vitamin C and dry matter as well a high content of compounds with antioxidant action, such as flavonoids and carotenoids (Reganold and Wachter, 2016; Baranski et al., 2014; Williams et al., 2016). On the other hand, Aires (2016) showed that the organic foods are similar to conventional ones. However, Baranski et al. (2014) concluded that organic crops present higher concentrations of antioxidants than conventional crops.

It can be seen that there are disagreements about the differences between organic and conventional products on the antioxidants levels. Thus, the aim of this study is to compare the organic and conventional production system over lettuce antioxidant activity.

## MATERIALS AND METHODS

The study was carried out in a greenhouse unit with planting being held in 2.5 L vases with 2 treatments types (conventional and organic fertilizer), using 30 samples for each treatment. The fertilization was performed from the soil analysis according to Table 1.

### Crude extract obtention

The crude extract was obtained by maceration in ethanol, using the leaves of organic and conventional lettuce. Then the material was dried in an oven at 50°C and crushed at a mill knives; purchasing a dry powder which was submitted to maceration with ethanol 99.5%

with filtering every 24 hours for five days. The the material was dried in the 40°C on a rotary evaporator BUCHI Heating Bath B-490. The percentage yield was calculated using the following equation:

$$\text{Yield(\%)} = \frac{\text{Crude extract weight}}{\text{Lettuce Leaves weight}} \times 100 \quad (1)$$

### Determination of total phenol content

The total phenol content of the samples was determined by the Folin-Ciocalteu method (Scherer and Godoy, 2014). 100 µL of the methanolic solution of the sample, 500 µL of the aqueous Folin 1:11 (v/v) solution and 400 µL of the aqueous Na<sub>2</sub>CO<sub>3</sub> solution (7.5%) were added in eppendorf flasks. The solution was stirred in a vortex for 30 seconds. After an aliquot of 250 µL transferred to 96-well plate, and kept in the dark for 2 hours and analyzed using a plate reader spectrophotometer at 740 nm. The results were expressed in mg Gallic Acid equivalent/g dry extract. All experiments were realized in triplicate.

### Antioxidant activity with DPPH

From stock solutions (1.0 mg/mL) of samples of conventional lettuce (CL) and organic lettuce (OL), test solutions were prepared at concentrations of 500 to 6 µg/mL (CL) and the 250 µg/mL (OL). An aliquot of 100 µL of sample test solution were transferred to a 96-well plate and 40 µL of methanolic solution of 2,2-diphenyl-1-picrylhydrazyl (DPPH) 0.3 mM. Blank solution was prepared using 100 µL of sample test solution and 40 µL of methanol for each concentration. Control solution was prepared using methanol (100 µL) and DPPH 0.3 mM. The samples was kept in the dark for 30 minutes and analyzed in a plate reader spectrophotometer at 518 nm. All experiments were realized in triplicate. The antioxidant activity percent was obtained using the follow equation:

$$\text{AAP(\%)} = 100 - \frac{\text{Abs}_{\text{Sample}} - \text{Abs}_{\text{Blank}}}{\text{Abs}_{\text{Control}}} \times 100 \quad (2)$$

### Ferric reducing antioxidant power (FRAP)

The Ferric reducing antioxidant power test was realized as described by Aras et al. (2017). The FRAP test was realized using 30 µL of methanolic sample solution, 90 µL of distilled water and 900 µL FRAP reagent (25 mL of acetate buffer 0.3 M, 2.5 mL of 2,4,6-Tri(2-pyridyl)-s-triazine (TPTZ) 10 mM and 2.5 mL of an aqueous solution of ferric chloride 20 mM). The test solutions were homogenized in the vortex and an aliquot of 250 µL was transferred to 96-well plate and kept at 37°C for 30 min. The sample absorbance was analyzed using a plate reader spectrophotometer at 595 nm. The trolox standard curve was constructed from these test solutions at concentrations of 10 µM a1000 µM. The results were expressed in µM Trolox equivalent/g of extract. All experiments were realized in triplicate (Table 3).

## RESULTS AND DISCUSSION

### Determination of total phenol content

Varying the concentration of gallic acid, the analytical

**Table 1.** Fertilization systems and its composition/amount.

Fertilizer	System	Compounds	Amount
Planting N:P <sub>2</sub> O <sub>5</sub> :K <sub>2</sub> O (30:90:60)	Conventional	Urea	70 kg/ha or 0,2 g/vase
		Simple superphosphate	500 kg/ha or 1,4 g/vase
		Potassium chloride	105 kg/ha or 0,3 g/vase
		Dolomitic limestone	1,4 ton/ha or 1,8 g/vase
	Organic	Organic compound (N:1.98%)	50 ton/ha or 65 g/vase
		Gafsa Phosphate	1 ton/ha or 2,7 g/vase
		Potassium sulphate	120 kg/ha or 0,32 g/vase
		Dolomitic limestone	1,4 ton/há or 1,8 g/vase
15 after planting N:P <sub>2</sub> O <sub>5</sub> :K <sub>2</sub> O (40:0:0)	Conventional	Urea	90 kg/ha or 0,25 g/vase (15 days after planting)
	Organic	—	—

**Table 2.** Total phenol content (mg EAG/100 g) obtained from ethanolic extracts of organic and conventional lettuce.

System	Present study	Kapoulas et al. (2017)	Fontana et al. (2018)
Organic	301.17a	~30-40	27.4
Conventional	204.50b	~40-50	27.1

Means followed by the same letter in the column does not differ at the 5% probability level.

**Table 3.** FRAP (mg of quecetin/g of sample) of conventional and organic lettuce ethanolic extracts.

System	FRAP
Organic	4209.52 <sup>a</sup>
Conventional	3553.33 <sup>b</sup>

Means followed by the same letter in the column does not differ at the 5% probability level.

curve could be obtained, with excellent repeatability and linear correlation coefficient (0.995) (Figure 1).

The total phenols content were obtained from the absorbance values of the samples, using the straight line equation of analytical curve of Gallic acid. The results were expressed as mg Gallic acid equivalent per gram of sample. As seen in Table 1 the values of this study were different from those obtained by Kapoulas et al. (2017) and Fontana et al. (2018) for different lettuce cultivars.

Several studies with bran and refined flour (Mazzoncini et al., 2015), chicory (Sinkovič et al., 2015), lemons (Uckoo et al., 2015) and tomato (Watanabe et al., 2015) also showed that the organic fertilizer sources provided greater quantities of phenolic compounds.

### Ferric Reducing Antioxidant Power (FRAP)

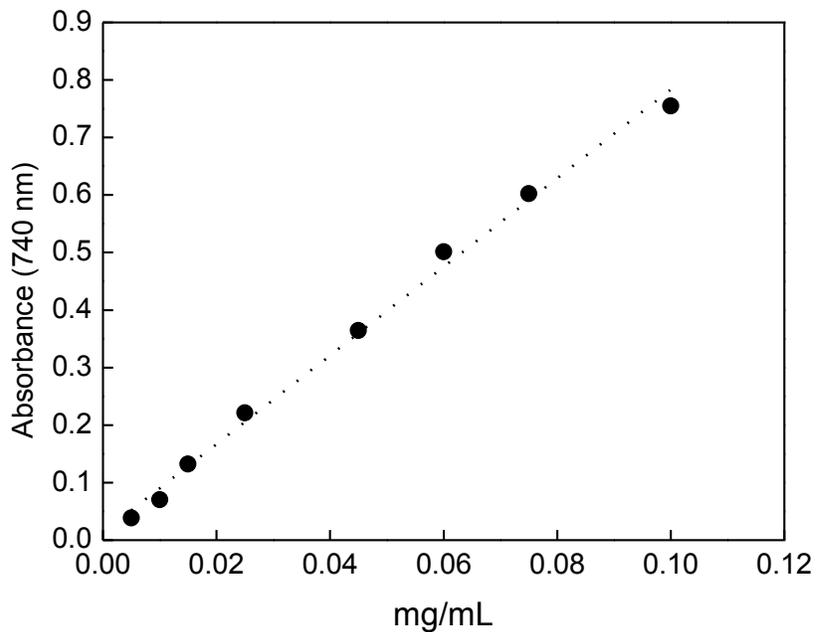
The analytical curve was constructed from the samples

absorbances of trolox (Figure 2).

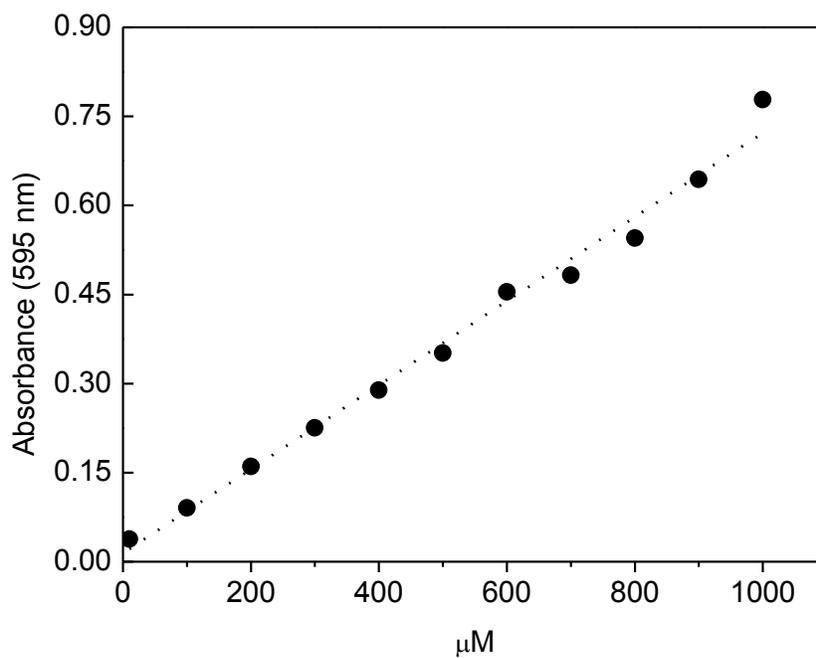
From the absorbance values at 595 nm and using the straight line equation of the analytical curve of trolox, the FRAO levels were obtained (Table 2). It turns out that there was a significant difference ( $P < 0.05$ ) between the lettuces produced in both cropping systems. Lettuce produced with organic fertilizer sources presented 18.47% more antioxidant activity as against that produced with conventional sources. It was verified a significant difference ( $P < 0.05$ ) between both culture systems. Organic lettuce presented 18.47% more antioxidant activity than lettuce produced from conventional sources.

### Antioxidant activity using DPPH method

The antioxidant percent (AAP%) were obtained from the samples' absorbance in different concentrations (Figures 3 and 4).



**Figure 1.** Analytical curve of Gallic acid.



**Figure 2.** Analytical curve of Trolox.

The changes in the antioxidant effect of extracts depending on the concentrations, the results are presented by CE50 value, since this parameter indicates the concentration of the sample needed to promote 50% of AAP%. The lower CE50 value, the higher the antioxidant activity. From the results of absorbance of the

sample, an analytical curve was constructed and through the straight line equation the CE50 was obtained. To calculate the CE50 for conventional and organic lettuce, the equation 3 and 4 were used, respectively.

$$50 = 0.105 \times \text{CE50} + 26.64 \quad (3)$$

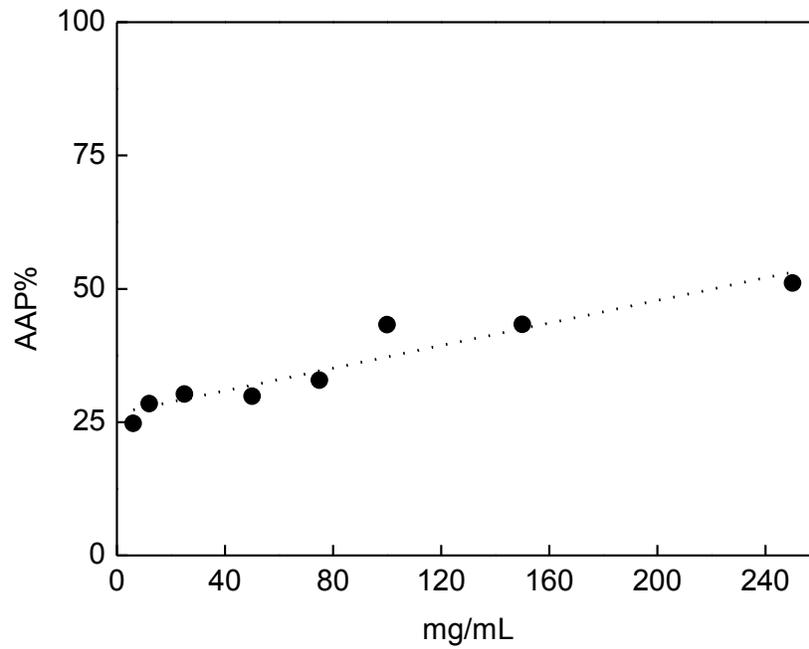


Figure 3. Antioxidant activity of conventional lettuce.

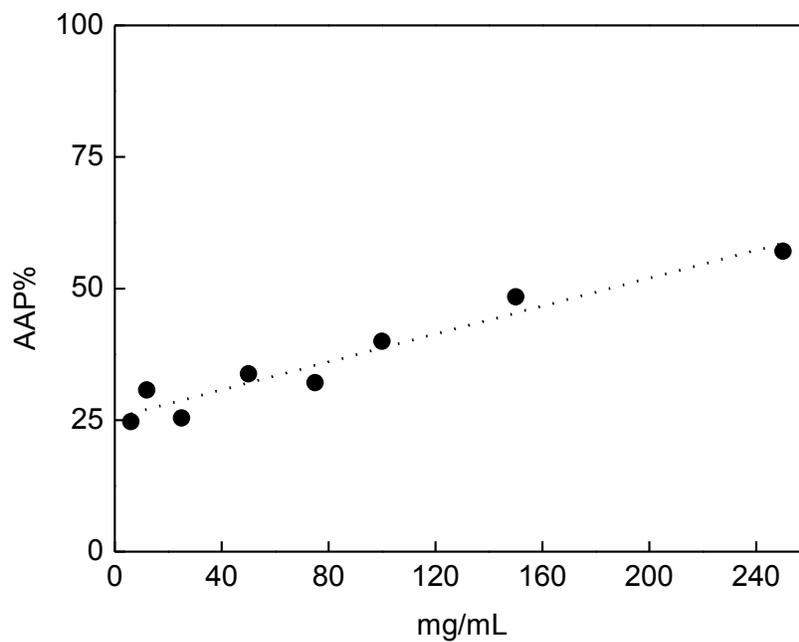


Figure 4. Antioxidant activity of organic lettuce.

$$50 = 0.132 \times CE_{50} + 25.43 \quad (4)$$

The  $CE_{50}$  values are listed in Table 4. It was verified that the organic lettuce showed the highest antioxidant capacity ( $CE_{50} = 0.18$  mg/mL) than the conventional

lettuce ( $CE_{50} = 0.22$  mg/mL). These results are similar to those obtained by Harsha et al. (2013) that detected 0.16 mg/mL in organic lettuce and by Kurubas et al. (2018) who found that lettuce produced in organic systems presented a higher effectiveness in antioxidant capacity

**Table 4.** CE50 values of organic and conventional lettuce.

System	CE50 ( $\mu\text{g/ml}$ )
Organic	185.14a
Conventional	220.77b

Means followed by the same letter in the column does not differ at the 5% probability level.

when compared with the conventional lettuce.

Rigueira et al. (2017) verified that the leaves and stems of collard greens cultivated in organic system had a higher antioxidant activity and levels of phenolic compounds compared to the conventional system.

The difference between these results may be due the use of different cultivars and environmental conditions where the experiments were developed. Similarly, Kim et al. (2018) found that cultivating as well as color of lettuce can change the phenol content and antioxidant activities. Therefore, the levels of phenolic compounds and antioxidant activities can change depending on the genetic material, applied management and climatic conditions used. Thus, Durazzo et al. (2014) showed the importance of cultivating, agronomic practices, edaphoclimatic, harvest and harvest season. According to the authors the variation of these elements represents the role to getting more nutritious, healthy and maximizing levels of bioactive molecules in food.

There was a better performance of organic lettuce when compared with the conventional. Comparative studies between these fertilizing systems explain that the greater effects of antioxidant activity and levels of phenolic compounds are due to biotic and/or abiotic stress that plants are subjected. Baranski et al. (2014) reported that the increase of phenol content in organic lettuce it can be due to environmental stress on plants that have not received pesticides or synthetic fertilizers for your development. Sharma et al. (2017) showed that the highest content of phenolic compounds in organic brassica leaves is probably due to interference of inorganic fertilizers and pesticides with the biosynthetic role of phenolic compounds.

The biosynthesis of phenolic compounds in plants is strongly influenced by cultivar (Aires, 2016), fertilization (Medeiros et al. 2016), temperature, light and seasonal variations (Kapoulas et al., 2017; Kurubas et al., 2018). The presence of higher concentrations of polyphenols in plants could be explained by a greater absorption of phosphorus and a limited nitrogen availability (Fan et al. 2017). The discovery that organic crops have a lower  $\text{N}$ ,  $\text{NO}_3^-$  and  $\text{NO}_2^-$  concentrations support the theory that differences of concentrations of antioxidants/phenolic compounds between organic and conventional crops are influenced by different nitrogen levels supply. This hypothesis is supported by previous studies that have suggested that under high nitrogen availability, plants can allocate photosynthesis carbohydrates to primary

metabolism and rapid growth while producing less amounts of secondary metabolites involved in your defense (Caretto et al., 2015).

This study does not show incidence of plagues, diseases or pesticide application, thus the effects are due the sources of fertilizer used. The increased amounts of phenolic compounds and antioxidant activity are due to the gradual release of nutrients and during the development of plants.

## Conclusions

Organic Lettuce had a better performance as regards phenolic compounds contents and antioxidant potential. The source of organic fertilizer is responsible for the effectiveness in antioxidant capacity and levels of phenolic compounds.

## CONFLICT OF INTERESTS

The authors declare that they have no conflict of interest.

## REFERENCES

- Aires A (2016). Conventional and Organic Farming-Does Organic Farming Benefit Plant Composition, Phenolic Diversity and Antioxidant Properties? Organic Farming-A Promising Way of Food Production. InTech <http://dx.doi.org/10.5772/61367>
- Aras A, Silinsin M, Bingol MN, Bursal E (2017). Identification of bioactive polyphenolic compounds and assessment of antioxidant activity of *Origanum acutidens*. *ILNS* 66:1-8.
- Baranski M, Srednicka-Tober D, Volakakis N, Seal C, Sanderson R, Stewart GB, Benbrook C, Biavati B, Markellou E, Giotis C, Gromadzka-Ostrowska J, Rembialkowska E, Skwarlo-Sonta K, Tahvonen R, Janovska D, Niggli U, Nicot P, Leifert C (2014) Higher antioxidant and lower cadmium concentrations and lower incidence of pesticide residues in organically grown crops: a systematic literature review and meta-analyses. *British Journal of Nutrition* 112:794-811.
- Borges LS, Vieira MCS, Vianello F, Goto R, Lima GPP (2016). Antioxidant compounds of organically and conventionally fertilized jambu (*Acmella oleracea*). *Biological Agriculture and Horticulture* 32:149-158.
- Calado JCP, Albertão PA, de Oliveira EA, Letra MHS, Sawaya ACHF, Marcucci MC (2015) Flavonoid Contents and Antioxidant Activity in Fruit, Vegetables and Other Types of Food. *Agricultural Sciences* 6:426-435.
- Caretto S, Linsalata V, Colella G, Mita G, Lattanzio V (2015). Carbon Fluxes between Primary Metabolism and Phenolic Pathway in Plant Tissues under Stress. *International Journal of Molecular Sciences* 16:26378-26394. doi:10.3390/ijms161125967
- Durazzo A, Azzini E, Lazzé MC, Raguzzini A, Pizzala R, Maiani G, Palomba L, Maiani G (2014), Antioxidants in italian head lettuce (*Lactuca sativa* var. *Capitata*) grown in organic and conventional systems under greenhouse conditions. *Journal of Food Biochemistry* 38:56-61.
- Fan DM, Fan K, Yu CP, Lu YT, Wang XC (2017). Tea polyphenols dominate the short-term tea (*Camellia sinensis*) leaf litter decomposition. *Journal of Zhejiang University-Science B*. 18:99-108.
- Fontana L, Rossi CA, Hubinger SZ, Ferreira MD, Spoto MHF, Sala FC, Verruma-Bernardi MR (2018). Physicochemical characterization and sensory evaluation of lettuce cultivated in three growing systems. *Horticultura Brasileira* 36:20-26.
- Harsha SN, Anilakumar KR, Mithila MV (2013). Antioxidant properties of *Lactuca sativa* leaf extract involved in the protection of biomolecules.

- Biomedicine and Preventive Nutrition 3(4):367-373.
- Kapoulas N, Koukounaras A, Ilić ZS (2017). Nutritional quality of lettuce and onion as companion plants from organic and conventional production in north Greece. *Scientia Horticulturae* 219:310-318.
- Kim DE, Shang X, Assefa AD, Keum YS, Saini RK (2018). Metabolite profiling of green, green/red, and red lettuce cultivars: Variation in health beneficial compounds and antioxidant potential, *Food Research International* 105:361-370.
- Kurubas MS, Maltas AS, Dogan A, Mustafa K, Mustafa E (2018). Comparison of organically and conventionally produced Batavia type lettuce stored in modified atmosphere packaging for postharvest quality and nutritional parameters. *Science Food and Agriculture* <https://doi.org/10.1002/jsfa.9164>
- Mazzoncini M, Antichi D, Silvestri N, Ciantelli G, Sgherri C (2015). Organically vs conventionally grown winter wheat: Effects on grain yield, technological quality, and on phenolic composition and antioxidant properties of bran and refined flour. *Food Chemistry* 175:445-451
- Medeiros CH, Ribeiro LV, Custodio T, Morselli T, Sedrez F (2016). Substratos alternativos para a produção de mudas de alface. *Revista Científica Rural* 18:100-107.
- Nimse SB, Pal D (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Advances* 5:27986-28006.
- Perez-Lopez U, Sgherri C, Miranda-Apodaca J, Micaelli F, Lacuesta M, Mena-Petite A, Quartacci MF, Muñoz-Rueda, A. (2018). Concentration of phenolic compounds is increased in lettuce grown under high light intensity and elevated CO<sub>2</sub>. *Plant Physiology and Biochemistry* 123:233-241.
- Reganold JP, Wachter JM (2016). Organic agriculture in the twenty-first century. *Nature Plants* 2(2):15221.
- Rigueira GDJ, Bandeira AVM, Chagas CGO, Milagres RCRM (2017). Atividade antioxidante e teor de fenólicos em couve-manteiga (brassica oleracea l. var. acephala) submetida a diferentes sistemas de cultivo e métodos de preparo. *Semina: Ciências Biológicas e da Saúde* 37(2):3-12.
- Santos J, Oliveira MBPP, Ibáñez E, Herrero M (2014). Phenolic profile evolution of different ready-to-eat baby-leaf vegetables during storage. *Journal of Chromatography A* 1327:118-131.
- Scherer R, Godoy HT (2014). Effects of extraction methods of phenolic compounds from *Xanthium strumarium* L. and their antioxidant activity. *A Revista Brasileira de Plantas Mediciniais* 16:41-46.
- Shahidi F, Ambigaipalan P (2015). Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects – A review. *Journal of Functional Foods* 18:820-897.
- Sharma, A, Kumar V, Kanwar MK, Thukral AK, Bhardwaj R (2017). Phytochemical profiling of the leaves of Brassica juncea L. using GC-MS. *International Food Research Journal* 24:547-551.
- Sinkovič L, Demšar L, Žnidarčič D, Vidrih R, Hribar J, Treutter D, (2015). Phenolic profiles in leaves of chicory cultivars (*Cichorium intybus* L.) as influenced by organic and mineral fertilizers. *Food Chemistry* 166:507-513.
- Sofo A, Lundegårdh B, Mårtensson A, Manfra M, Pepe G, Sommella E, Nisco M, Tenore GC, Campiglia P, Scopa A (2016). Different agronomic and fertilization systems affect polyphenolic profile, antioxidant capacity and mineral composition of lettuce. *Scientia Horticulturae* 204:106-115.
- Talens P, Mora L, Bramley PM, Fraser PD (2016). Antioxidant compounds and their bioaccessibility in tomato fruit and puree obtained from a DETIOLATED-1 (DET-1) down-regulated genetically modified genotype. *Food chemistry* 213:735-741.
- Turati F, Rossi M, Pelucchi C, Levi F, La Vecchia C (2015). Fruit and vegetables and cancer risk: A review of southern European studies. *British Journal of Nutrition* 113(S2):S102-S110.
- Uckoo RM, Jayaprakasha GK, Patil BS (2015). Phytochemical analysis of organic and conventionally cultivated Meyer lemons (*Citrus meyeri* Tan.) during refrigerated storage. *Journal of Food Composition and Analysis* 42:63-70.
- Watanabe M, Ohta Y, Licang S, Motoyama N, Kikuchi J (2015). Profiling contents of water-soluble metabolites and mineral nutrients to evaluate the effects of pesticides and organic and chemical fertilizers on tomato fruit quality, *Food Chemistry* 169:387-395.
- Williams DJ, Edwards D, Pun S, Chaliha M, Burren B, Tinggi U, Sultanbawa Y (2016). Organic acids in Kakadu plum (*Terminalia ferdinandiana*): The good (ellagic), the bad (oxalic) and the uncertain (ascorbic). *Food Research International* 89:237-244.